Characterization of Lateral Root Development at the Onset of Storage Root Initiation in ‘Beauregard’ Sweetpotato Adventitious Roots

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Abstract. This study characterized lateral root (LR) development attributes during the onset of storage root (SR) initiation stage in ‘Beauregard’ sweetpotato. SR initiation has been defined as the appearance of cambia around the protoxylem and secondary xylem elements. Our results showed that 20-day-old adventitious roots (ARs) classified as SRs had 53% and 85% greater mean LR count than pencil roots (PRs) and lignified roots (LGs), respectively. SRs had 53% and 78% greater mean LR density relative to PRs and LGs, respectively. SRs had 66% and 130% greater mean total LR length than PRs and LGs, respectively. SRs had lower mean main root (MR)/LR length ratio compared with PRs (~38%) and LGs (~60%). SRs had 70% and 134% greater mean surface area than PRs and LGs, respectively. SRs had lower mean MR/LR surface area ratio compared with PRs (~42%) and LGs (~62%). The plot of the first and second principal components revealed the presence of a gradient between extreme LG and SR clusters, suggesting a developmental transition between LGs and SRs with PRs representing an intermediate developmental stage. Although AR architecture is not the sole determinant of SR formation, our data help provide a basis for integrating AR architecture attributes with other factors that are known to influence SR initiation. Growth substrate moisture variability influenced LR development during the critical SR initiation period. Relative to the control treatments, water deprivation 10 to 20 days after transplanting (DAT) reduced mean LR count, length, and surface area by 49%, 103%, and 94%, respectively. Saturated conditions 10 to 20 DAT reduced mean LR count, length, and surface area by 75%, 81%, and 77%, respectively. These results represent the first evidence for the association between anatomical cues of SR initiation and root architecture and provide corroborating data that soil moisture variability 10 to 20 DAT directly influences SR yield potential through AR architecture modifications that are associated with diminished SR formation. This information can be used to further optimize SR yield by identifying agroclimatic and management variables that are associated with desirable LR development during the critical SR initiation stage.

Koshimizu and Nishida (1949) measured LR branching in sweetpotatoes and postulated that LR branching may be very important “to supply the internal growth elements” for storage root formation. Pardales and Yamauchi (2003) studied sweetpotato LR development during the establishment period, which they defined as 4 weeks after transplanting and suggested a link between LR variability and the onset of SR initiation in ARs. Recent molecular data have provided further evidence for the presumptive link between LR development and SR initiation. Specifically, IbmMADS1 has been documented as related to SR initiation in sweetpotato (Ku et al., 2008). They have shown that IbmMADS1 transcripts were localized to emerging LR primordia and immature meristematic cells such as the protoxylem and protophloem within the AR stele (Ku et al., 2008). SR initiation has previously been defined as the appearance of cambia around the protoxylem and secondary xylem elements (Togari, 1950; Wilson and Lowe, 1973). Togari (1950) also suggested that once stele lignification occurred, SR initiation did not proceed. LRs originate from the pericycle and the radial location of LRs is correlated with the internal architecture of the stele (Laskowski et al., 1995). In radish (Raphanus sativus) and Arabidopsis, which have diarch steles, LRs arise from pericycle cells located in files adjacent to a xylem pole (Laskowski et al., 1995). Thus, in sweetpotato ARs, the number of LR rows corresponds to the number of protoxylem poles (Yasu, 1944). In ‘Beauregard’ sweetpotato, the number of protoxylem poles varies from five to 10, but the majority of ARs show almost equal occurrence of five (pentarch) or six (hexarch) poles (Villordon et al., 2009a). Previous reports of the morphological and anatomical characterization of SR initiation did not provide LR development data (Belehu et al., 2004; Togari, 1950; Villordon et al., 2009a; Wilson and Lowe, 1973). Pardales and Yamauchi (2003) quantified LR count and total length response to soil moisture variability among greenhouse-grown sweetpotato genotypes but did not present data for SR initiation. Recent data have showed that LR initiation and development were detected as early as 9 DAT among in situ field-grown ‘Beauregard’ plants (Villordon et al., 2011). We have previously reported that SR initiation was detected as early as 13 d in field-grown ‘Beauregard’ plants (Villordon et al., 2009b). In model species, LRs have been referred to as a major determinant of root architecture and contribute to water use efficiency and facilitate the extraction of micro- and macronutrients from the soil (Casimiro et al., 2003). Thus, the presumptive role of LR development in SR initiation represents a knowledge gap in the current research on internal and external factors that influence SR initiation.

The knowledge of the association between root architecture and SR initiation will benefit current and future studies in further understanding the mechanism of SR initiation in sweetpotatoes. Such information will also lead to further clarity in understanding how external stimuli such as soil moisture variability and nutrient availability promote or hinder SR initiation and can lead to modifications or enhancements of current management practices. Togari (1950) has previously reported that the growing environment during the first 20 d of AR development directly influenced SR initiation. Togari (1950) also reported that “dryness” and “shortage of O2” during the first 20 d favored lignification and the development of PRs. Pardales and Yamauchi (2003) have previously documented that five sweetpotato genotypes (PRES-118, PRS-74, PRS-46, VSP-4, and VSP-2) subjected to continuous deficient soil moisture showed an overall reduction in LR number and total length. Pardales and Yamauchi (2003) also reported that root development was suppressed among plants subjected to continuous “waterlogged” conditions. In a previous 32-d destructive sampling study, we have documented that the majority of greenhouse-grown ‘Beauregard’ AR samples showed evidence for SR initiation at 19 d (Villordon et al., 2009b). The primary objective of this work was to provide quantitative morphological evidence of the link between LR development, a major determinant of root architecture, and anatomical cues associated with the onset of SR initiation stage in ‘Beauregard’ sweetpotato. A secondary objective was to document
the relationship between soil moisture variability and LR development during the first 20 d of growth.

Materials and Methods

Plant materials. The greenhouse experiments were conducted in Chase, LA (lat. 32°6’ N, long. 91°42’ W). In each of 15 Oct. 2010 and 1 Oct. 2011, virus-tested ‘Beauregard’ Generation 1 SRs were bedded in washed river sand and served as a source of transplants. One set of experiments generated samples for anatomical observations and associating LR attribute with anatomical cues of SR initiation. The second set of experiments investigated the influence of growth substrate moisture content on LR development. Washed river sand was used as a growth substrate for all experiments. In each experiment, cuttings were set in 10-cm-diameter polyvinyl chloride (PVC) pots (height = 30 cm) with detachable plastic bottoms. Each plastic bottom had five drain holes (2 mm in diameter). In addition, each PVC pot had four rows of side drain holes (2 mm in diameter; 3 cm apart within row) that were located diametrically opposite each other. Fertilizer application consisted of 0.76 g of 5N–20P–20K per pot. The greenhouse temperature regime was 29 °C for 14 h (day) and 18 °C for 10 h (night). Photosynthetic photon flux (PPF) for all experiments ranged from 150 to 1300 μmol·m⁻²·s⁻¹. Supplementary lighting was provided using white fluorescent lights (≈42 μmol·m⁻²·s⁻¹ PPF) for 14 h d⁻¹. PPF was measured at the canopy level with a quantum sensor (Model QSO-S; Decagon Devices Inc., Pullman, WA). The relative humidity (RH) averaged 60%. Temperature and RH were monitored at the canopy level using an integrated temperature and RH sensor (Model RHT; Decagon Devices Inc.). Unless otherwise indicated, the moisture of the growing substrate was maintained ≈50% of field capacity (≈7% volumetric water content [VWC]). Growth substrate moisture was measured with EC25 soil moisture sensors (Model EC-5; Decagon Devices Inc.) inserted vertically at the 2- to 7-cm depth. These growth conditions were very similar to those used in previous experiments that characterized AR development and the timing of SR initiation in ‘Beauregard’ sweet-potato (Villordon et al., 2009a, 2009b). In experiments that studied the effect of moisture levels on LR development, a uniform watering regime was imposed during the first 10 d. This was to ensure that all cuttings were established (initiated ARs) and that ARs underwent initial LR development. After 10 d, the following moisture treatments were imposed for an additional 10 d: control (moisture was maintained near 50% of field capacity; ≈200 mL distilled water per pot per irrigation event, up to three irrigation events until harvest), dry (water deprivation 10 DAT), and saturated (400 mL water per irrigation event, up to five irrigation events until harvest). Growth substrate moisture readings for the control treatment ranged from 5% to 8% VWC. The ranges for dry and saturated treatments were 1% to 5% VWC and 8% to 12% VWC, respectively. All sensors were connected to a data logger that was configured to collect measurements every 60 min (Model EM50R; Decagon Devices Inc.).

In all experiments, plants were grown for 20 d after which near-intact root systems were collected. At harvest, the detachable plastic bottoms were removed and the pot was tilted and the growth substrate was gradually removed using a stream of water. The planting dates for AR anatomical and LR attribute characterization experiments were 2 Mar. 2011, 12 Nov., 2011, and 28 Nov. 2011. The harvest dates were 22 Mar. 2011, 2 Dec. 2011, and 18 Dec. 2011. The planting dates for the growth substrate moisture experiments were 13 Apr. 2011 and 19 Dec. 2011. The harvest dates were 3 May 2011 and 8 Jan. 2012. All experiments were arranged as a randomized complete block repeated across planting dates. There were five replicates (one plant per pot = one replicate) in each of the LR characterization experiments and in each of the substrate moisture experimental treatments. As a result of the variability of the number of adventitious roots per plant, there were unequal subsample sizes of adventitious roots that met the criteria for scanning and analysis in all experiments. In

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Fig. 1. Adventitious root samples harvested at 20 (A) and 40 (B) d after transplanting and representative micrographs used to classify 20-d-old adventitious roots into various stages in ‘Beauregard’ sweet-potato: initiated storage root (C), pencil root (D), and lignified root (E). Adventitious roots (A–B) were floated on waterproof trays and images were acquired using a scanner equipped with the WinRhizo positioning system. (A) Location of tissue sections used for anatomical examination for each 20-d-old adventitious root sample. Samples C–E were derived from the 4.5-cm section of the respective 20-d-old adventitious root specimens. LR = lateral root; MR = main root; SR = initiated storage root; PR = pencil root; LG = lignified root; AC = anomalous cambium; PX = protoxylem; CMC = central metaxylem cell; X = protoxylem element remains connected to CMC; LS = lignified stele; OR = origin of lateral root. Scale bars for A–B = 1 cm. Scale bars for C–E = 0.15 mm.
each experiment, extra replicates were grown for validation of visible storage root formation at 40 d.

Root image acquisition and analysis. Preliminary calibration experiments were performed to determine optimal settings for image acquisition and analysis parameters. These experiments included comparison of scanner-based and manual measurements using representative AR samples. Based on these preliminary experiments, the image acquisition parameter was set to “medium” accuracy (400 dpi; image size = 8 MB), whereas analysis precision was set to “high.” In all experiments, intact ARs that were 20 cm or greater in length were floated on waterproof trays and scanned using a specialized Dual Scan optical scanner (Regent Instruments Inc., Quebec, Canada). The acquisition and image analysis software was WinRHIZO Pro (Version 2009; Regent Instruments Inc.). Debris removal among scanned images was performed manually using the WinRhizo Pro Edition working mode. Debris consisted mainly of images of sand particles and occasional broken root segments (1 cm or less in length). Root types were automatically classified based on root diameter, which was in turn based on predetermined size intervals. In our work, the intervals were set to 0 to 0.9 and 0.9 to 20 mm. The first interval classified LRs, whereas the second interval classified the main AR (main root). These intervals were based on prior calibration experiments on ‘Beauregard’ ARs. Wilson and Lowe (1973) have previously reported that around the onset of SR initiation, the MRs were ~1 mm. LR attributes that were measured from scanned images included LR count, LR length (total length of all LRs), and LR surface area (total surface area of all LRs). MR length was verified using Image-Tool (Univ. of Texas Health Science Center at San Antonio, available at ftp://maxrad6.uthscsa.edu). LR density was calculated by dividing LR count by MR length. The following ratios were calculated: MR length/LR length (MR/LR length ratio) and MR surface area/LR surface area (MR/LR surface area ratio).

Anatomical observations. AR samples for anatomical observations were prepared using free-hand sectioning and staining with toluidine blue (Eguchi and Yoshida, 2008). Samples were prepared by placing a proximal segment of an AR (~7 to 10 cm) in a petri dish containing distilled water followed by the excision and staining of three separate tissue sections (2.5, 3.5, and 4.5 cm from the proximal end) (Fig. 1A). The thickness of transverse sections ranged from 8 to 15 μm. Digital micrographs were taken with a Motic Cam 1000 (Motic Instruments Inc., British Columbia, Canada). In previous SR initiation studies, we routinely observed that anatomical cues of SR initiation (anomalous cambia) occurred within the 2.5 to 5 cm proximal end of an AR and this section typically became swollen after 30 d when grown in conditions described previously (Fig. 1B) (Villordon et al., 2009a, 2009b). Wilson and Lowe (1973) documented that this zone of thickening initially did not extend over the entire region of initiation.

Classification of adventitious root development stage. We used the criteria described by Wilson and Lowe (1973) in classifying the AR developmental stage: initiated SRs, PRs, and LGs. According to this scheme, ARs were classified as initiated when circular anomalous cambia were detected around the central metaxylem cell as well as around each of the discrete protoxylem elements in one or all of the proximal sections (Fig. 1C). For initiated ARs without a central metaxylem cell, primary cambia were associated with meristematic activity in the pith cells and the formation of anomalous cambia around the protoxylem elements. PRs were ARs that showed some meristematic activity around the central metaxylem cell, but one or more of the protoxylem elements remained connected to the central cell (Fig. 1D). LGs were ARs that eventually developed into mature thickened roots (up to 5 mm in diameter) with a heavily lignified stele, xylem rays, a broad secondary cortex, and limited secondary phloem (Fig. 1E).

Data collection and statistical analysis. A total of 106 ARs from three anatomical characterization experiments were unambiguously classified as SR (n = 25), PR (n = 32), and LG (n = 49). LR attribute data from these experiments were pooled and the combined data set was used for LR development attribute analysis (n = 106; AR anatomical characterization data set). Similarly, LR attribute data from the two growth substrate moisture experiments were pooled (n = 136; substrate moisture data set). The number of ARs that met the criteria for data acquisition varied among the moisture treatments: control (n = 42), dry (n = 39), and saturated (n = 55). The number of ARs per plant ranged from two to eight. All numeric data were transformed using log 10 and square root transformation, respectively. Ratios were arcsine-transformed. The unbalanced data sets were analyzed using SAS Proc Mixed (SAS Version 9.1; SAS Inc., Cary, NC). Fisher’s least significant difference test at the 0.05 P level indicated that main effects were statistically significant for each of the AR anatomical characterization and substrate moisture data sets. The data presented were means and confidence intervals.

Fig. 2. Adventitious roots that represent the shortest, intermediate, and longest cumulative lateral root length for each of the following stages in ‘Beauregard’ sweetpotato: initiated storage root (SR), pencil root (PR), and lignified root (LG). LR = lateral root. Scale bar = 2 cm.
calculated from balanced data sets (n = 60) that were generated by stratified resampling in IBM SPSS Statistics (Version 19; SPSS Inc., Chicago, IL). Bootstrapped confidence intervals were presented for all LR attribute data. Principal components analysis (PCA) (type = correlation) was performed on the balanced AR anatomical characterization data set using SAS PRINCOMP (SAS Version 9.1; SAS Inc.).

**Results**

**Relationship between lateral root development and adventitious root stage.** Representative images of LR development representing shortest, intermediate, and longest total LR lengths for each of SR, PR, and LG stages are shown in Figure 2. ARs classified as SRs had 53% and 85% greater mean LR count than PRs and LGs, respectively (Fig. 3A). SRs had 53% and 78% greater mean LR density than PRs and LGs, respectively (Fig. 3B). There were no significant differences in mean LR count and density between PRs and LGs. SRs had 66% and 130% greater mean total LR length than PRs and LGs, respectively (Fig. 3C). Mean LR length in PRs was 38% greater than LGs (Fig. 3C). When compared with SRs, the mean MR/LR length ratio was increased by 38% and 60% in PRs and LGs, respectively (Fig. 3D). The mean MR/LR length ratio among LGs was 38% greater than PRs. SR/LR mean surface area was 70% and 134% greater than PRs and LGs, respectively (Fig. 3E). PR/LR surface area was 37% greater than LG surface area. Relative to SRs, mean MR/LR surface area ratio was increased by 42% and 62% in PRs and LGs, respectively (Fig. 3F). The mean MR/LR surface area ratio among LGs was 35% greater than PRs. PCA showed that the first two principal components (PCs) accounted for 96% of the total variance (Table 1). PC1 accounted for 86% of the variance among samples with positive loadings of LR length, LR surface area, LR density, and LR count. 

### Table 1. Eigenvectors and eigenvalues for the first six principal components estimated from six lateral root attributes of ‘Beauregard’ sweetpotato adventitious roots grown in Louisiana.

<table>
<thead>
<tr>
<th>LR attributes</th>
<th>PC 1</th>
<th>PC 2</th>
<th>PC 3</th>
<th>PC 4</th>
<th>PC 5</th>
<th>PC 6</th>
</tr>
</thead>
<tbody>
<tr>
<td>Length</td>
<td>0.432369</td>
<td>-0.041885</td>
<td>0.447946</td>
<td>0.280676</td>
<td>-0.261731</td>
<td>0.680709</td>
</tr>
<tr>
<td>MR/LR length ratio</td>
<td>-0.410678</td>
<td>0.421374</td>
<td>0.260281</td>
<td>-0.266416</td>
<td>0.564967</td>
<td>0.442579</td>
</tr>
<tr>
<td>Surface area</td>
<td>0.430035</td>
<td>-0.027707</td>
<td>0.500011</td>
<td>0.188038</td>
<td>0.557471</td>
<td>-0.467076</td>
</tr>
<tr>
<td>MR/LR area ratio</td>
<td>-0.396377</td>
<td>0.512477</td>
<td>0.259509</td>
<td>0.592851</td>
<td>-0.312789</td>
<td>-0.252186</td>
</tr>
<tr>
<td>Count</td>
<td>0.384339</td>
<td>0.593236</td>
<td>0.127786</td>
<td>-0.573943</td>
<td>-0.345425</td>
<td>-0.187873</td>
</tr>
<tr>
<td>Density</td>
<td>0.393242</td>
<td>0.453167</td>
<td>-0.630804</td>
<td>0.366063</td>
<td>0.290483</td>
<td>0.153964</td>
</tr>
<tr>
<td>Eigenvalue</td>
<td>5.136686</td>
<td>0.642359</td>
<td>0.171238</td>
<td>0.031302</td>
<td>0.017535</td>
<td>0.001494</td>
</tr>
<tr>
<td>Cumulative</td>
<td>0.856</td>
<td>0.9631</td>
<td>0.9916</td>
<td>0.9968</td>
<td>0.9998</td>
<td>1.0000</td>
</tr>
</tbody>
</table>

*LR = lateral root; MR = main root; PC = principal component. Principal component analysis was performed on standardized LR attribute data (subtraction of mean followed by division by SD). Description of experimental procedures is found in “Materials and Methods.”*
PC2 accounted for an additional 11% of the variation with LR count, MR/LR area ratio, LR density, and MR/LR length ratio loading positively. Proceeding from negative to positive values of PC1, it was evident that PRs classified as LGs and SRs occupied extreme locations with a gradient existing between these samples (Fig. 4A). Within this gradient, PRs represented an intermediate stage with significant overlapping from LGs and SRs. The extreme SR and LG specimens are shown in Figures 2C and 2H, respectively. The PR specimen located at the extreme “bottom” of the horseshoe-shaped plot is shown in Figure 2E. The plot of PC1 and PC3 showed a general increase in scatter (variance) among PRs as well as the overlapping LGs and SRs on PC3 (Fig. 4B). This increased scatter among specimens reflects the fact that ARs with intermediate LR counts and MR/LR area ratios scored low on PC2.

**Relationship between growth substrate moisture variability and lateral root development.** Representative images of ARs showing shortest, intermediate, and longest total LR length for each of three moisture regimes (control, dry, saturated) 10 to 20 DAT from the total data set are shown in Figure 5. Relative to the control treatments, water deprivation 10 to 20 DAT reduced MR/LR mean count, length, and surface area by 49%, 103%, and 94%, respectively (Figs. 6A, 6C, and 6E). Overwatering 10 to 20 DAT reduced MR/LR mean count, length, and surface area by 86%, 81%, and 77%, respectively (Figs. 6A, 6C, and 6E). Relative to the control treatment, dry and overwatering regimes reduced MR/LR mean density by 45% and 92%, respectively (Fig. 6B). Relative to the control treatment, mean MR/LR length ratio was increased by 103% and 81% in dry and overwatering regimes, respectively. Mean MR/LR surface area ratio was increased by 46% in dry and saturated conditions, respectively. There were no statistical differences for all AR attributes between the dry and saturated treatments.

**Discussion**

Root growth and architecture are important factors that influence plant performance and yield survival yet are frequently overlooked in horticultural research (Wright and Wright, 2004). Our results in part support the hypothesis, first suggested by Koshimizu and Nishida (1949) and reiterated by Pardales and Yamauchi (2003), that postulates a relationship between root system architecture and SR yield potential in sweetpotato. In general, ARs that showed evidence for SR initiation had higher values for all measured LR-related attributes and lower values for calculated MR/LR-related ratios relative to PRs and LGs. PCA integrated these LR attributes and the “horseshoe”-shaped plot of PC1 and PC2 showed a gradient between the extreme LG and SR clusters with PRs appearing to represent an intermediate stage. Reymert (1991) has attributed the horseshoe pattern in PCA to a characteristic of data sets in which the correlations among variables are very high and approximately equal. The horseshoe pattern may also be caused by nonlinear data structures (Podani and Miklos, 2002). Hughes and Chapman (1995) observed a similar pattern in PCA of growth attributes in Silurian proetide trilobites (Aulacopleura Konincki) and attributed this pattern to the ontogenetic transition of A. Konincki. Wilson and Lowe (1973) referred to PRs as “intermediate structures” with some anomalous production of parenchyma cells in the central root axis. In ‘Beauregard’ sweetpotato, this horseshoe pattern appears to represent a growth trajectory where ARs with optimum LR number (Fig. 2E–F) but intermediate LR length possesses the competency to become SRs by undergoing further LR development. On the other hand, ARs (Figs. 2D, 2G, and 2H) with low LR number and density do not appear capable of becoming SRs unless additional LRs develop. In Arabidopsis, Dubrovsky et al. (2006) have presented evidence that LR primordia development occurred strictly acropetally and de novo initiation events were found between already developed LRs or LR primordia. Arrested or slowly developing LR primordia were found between developed LRs and it was concluded that there was a narrow development window for LR initiation (Dubrovsky et al., 2006). Thus, an elucidation of factors that influences LR number and density in sweetpotato will further enhance our knowledge of the presumptive association between root architecture plasticity and yield variability in this crop species. In the PCA plots, the overlapping of SRs and LGs indicates that LR development is not the sole determinant of SR formation. Current evidence shows that sucrose (Eguchi and Yoshida, 2008; Li and Zhang, 2003), cytokinin (Eguchi and Yoshida, 2008; Tanaka et al., 2008), and nitrogen (Villagarcia et al., 1998) are involved in the process of SR initiation. Our results provide a basis for further understanding how these variables influence SR initiation. Our data also provide evidence for the potential link between quantifiable morphological LR attributes.
intense interplant competition.'’ In other

in comparison with the control treatment. These
data provide evidence that soil mois
ture variability during the first 20 DAT, i.e.,
optimum soil moisture followed by drought
or saturated conditions, is directly related

to the determination of yield potential in ‘Beauregard’ sweetpotato. This helps to
explain earlier results reported by Togari (1950)
абout the yield-limiting effects of extreme
soil moisture conditions during the first 20 d.
Pardales and Yamauchi (2003) reported vari
able and significant reductions in LR count
(–44% to –66%) and length (–59% to –77%)
at 28 DAT among sweetpotato genotypes
subjected to continuous drought conditions.
The magnitudes of these differences were
similar to our results (Figs. 6A and 6C).
Pardales and Yamauchi (2003) stated that
drought was the “most limiting factor” in LR
development during the “establishment stage,”
which they defined as the first 4 weeks of
growth. However, our results showed that
saturated conditions also hindered LR de
velopment within this timeframe. This helps
to underscore the importance of properly
calibrating irrigation frequency and amount
for specific soil types in sweetpotato pro
duction. Our findings are also consistent with
experimental results in other species. Seo and
Park (2009) reported that drought conditions
suppressed Arabidopsis LR development.
Zook et al. (1986) have previously demon
strated that flooding conditions suppressed
LR growth in alfalfa (Medicago sativa). In
Arabidopsis, it has been shown that flooding
conditions resulted in increased ethylene
synthesis that inhibited lateral root initiation
and root growth (Ivanchenko et al., 2008).
This work can be used as a methodological
model for conducting follow-up studies to
determine if the sweetpotato root system
possesses a similar mechanism for determin
ing response to saturated conditions. Our
findings can also be used to enhance current
breeding and selection strategies, especially
for low-input environments. For example,
Zhu and Lynch (2004) documented that enhanced
LR rooting under phosphorus stress can be
harnessed as a useful trait for the selection
and breeding of more phosphorus-efficient
maize genotypes. Pardales and Yamauchi
(2003) documented that imposition of opti
mum soil moisture conditions after dry or
saturated conditions led to resumption of LR
development and concluded that root archi
tecture response to saturated conditions. Our
findings are also consistent with

and an LR-specific molecular indicator of SR
initiation as described by Ku et al. (2008).

In general, SRs had increased LR count,
length, and surface area relative to LGs and
PRs. This suggested a competitive advantage
relative to other ARs in terms of access to soil
moisture and nutrients. Green et al. (2005)
used the concept of “interlateral root com
petition” in describing the reduction of soil
volume exploited by developing Mediterranea
nean shrub (Pistacia lentiscus) roots with
shortened internode lengths. LR competition
within and among plants may help to deter
mine SR competency among ARs, especially
in marginal environments. In the PCA plots,
the gradient between extreme LG and SR
samples indicated a quantitative component
in the determination of the developmental
fate of ARs, i.e., LG, PR, or SR. Morphologi
cal plasticity in response to competition
effects has been investigated in sweetpotato.
For example, Sulaiman and Sasaki (2001)
observed a decrease of root and shoot dry
weight per hill with increased planting den
sities among the sweetpotato genotypes that
were studied and attributed this response to
“intense interplant competition.” In other

species, LR plasticity has been documented
to be involved in adaptive responses to the
soil environment. For example, Borch et al.
(1999) documented that low phosphorus
availability reduced LR number and sus
tained MR elongation, resulting in decreased
LR density in phosphorus-deficient common
bean (Phaseolus vulgaris). Arredondo and
Johnson (2011) proposed that the prolifer
ation of fine roots deployed into enriched soil
sites did not appear to require a precise
placement of LRs but rather rapid initiation
and elongation of multiple roots. In contrast,
the appropriate strategy under marginal soil
nutrient conditions appeared to involve the
emergence of fewer and spatially distant
roots that delayed LR competition and fa
vored the improvement of root foraging pre
cision (Berntson, 1994).

We demonstrated ‘Beauregard’ sweetpo
tato AR architecture plasticity in response to
growth substrate moisture variability 10 to 20
DAT and provided corroborating data to
an earlier report by Pardales and Yamauchi
(2003). Under low and excessive moisture
conditions 10 to 20 DAT, LR count, length,
and surface area were significantly reduced

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vored the improvement of root foraging pre
cision (Berntson, 1994).

We demonstrated ‘Beauregard’ sweetpo
tato AR architecture plasticity in response to
growth substrate moisture variability 10 to 20
DAT and provided corroborating data to
an earlier report by Pardales and Yamauchi
(2003). Under low and excessive moisture
conditions 10 to 20 DAT, LR count, length,
and surface area were significantly reduced

et al., 2008; Lopez-Bucio et al., 2002). Environmental variables include soil nutrients such as nitrate (Zhang and Forde, 1998), phosphate (Johnson et al., 1996), and sulfate (Kutz et al., 2002). In many cases, nutrient variability appears to influence hormone sensitivity. For example, phosphorus deficiency alters *Arabidopsis* root architecture by changing sensitivity to auxins (Lopez-Bucio et al., 2002). The information about the relationship between LR development and sweetpotato SR initiation helps to provide a basis for integrating the knowledge gained about internal and external cues that influence LR initiation and development in model systems.

**Conclusion**

We have documented the association between LR attributes and anatomical cues associated with onset of SR initiation in ‘Beauregard’ sweetpotato. We have also provided corroborating evidence that soil moisture variability 10 to 20 DAT directly influenced LR development and demonstrated the importance of optimizing soil moisture during the critical SR initiation period. Our data indicate that AR architecture is not the sole determinant of SR initiation. However, these findings can be used to further optimize management and environmental variables during the critical SR initiation period to further optimize SR yield in sweetpotato.

**Literature Cited**


Eguchi, T. and S. Yoshida. 2008. Effects of application of sucrose and cytokinin to roots on