

# Vegetative Phase Transition and Corn Borer Resistance of *shrunken2* versus *sugary1* Sweet Corn Near-isogenic Inbred Lines

Pedro Revilla<sup>1</sup>

Misión Biológica de Galicia, Spanish Council for Scientific Research, Apartado 28, 36080 Pontevedra, Spain

William F. Tracy

Department of Agronomy, University of Wisconsin-Madison, Madison, WI 53706

Pilar Soengas, Bernardo Ordás, Amando Ordás, and Rosa Ana Malvar

Misión Biológica de Galicia, Spanish Council for Scientific Research, Apartado 28, 36080 Pontevedra, Spain

ADDITIONAL INDEX WORDS. *Zea mays*, vegetative phases, *Ostrinia nubilalis*, *Sesamia nonagrioides*

**ABSTRACT.** The genes *sugary1* (*su1*) and *shrunken2* (*sh2*) are commonly used to produce sweet and super-sweet corn (*Zea mays* L.), respectively. In this work we compare corn borer [European corn borer (ECB) (*Ostrinia nubilalis* Hbn.) and pink stem borer (PSB) (*Sesamia nonagrioides* Lef.)] susceptibility in seven pairs of *su1* and *sh2* near-isogenic sweet corn inbreds (101t, C23, C40, C68, Ia453, Ia5125, and P39) and the relationship between corn borer resistance and vegetative phase transition. The seven pairs of near-isogenic inbreds were evaluated under corn borer infestation during 3 years in northwestern Spain. Differences among inbreds were significant for most of the traits, although resistance was partial. Ia5125*su1* and C40*su1* were the most resistant inbreds. Differences between a few pairs of near-isogenic *su1* and *sh2* strains were significant for some vegetative phase change and corn borer damage-related traits. Generally *su1* strains flowered earlier, had a shorter juvenile phase, fewer PSB, and more ECB larvae than *sh2* strains. However *su1* and *sh2* strains did not differ significantly for most traits related to phase transition and corn borer damage; notably ear damage was not significantly different between *su1* and *sh2* strains. These results suggest that theoretical and practical results of sweet corn (*sugary1*) breeding for corn borer resistance could be capitalized for super-sweet corn (*shrunken2*) breeding.

Sweet corn varieties used in temperate areas are primarily homozygous for *sugary1* (*su1*), while super-sweet corn is homozygous for *shrunken2* (*sh2*) and is currently the second most widely used endosperm type after *su1* (Tracy, 2000). There are several differences between *su1* and *sh2*, with sweetness the most evident for consumers (Tracy, 2000). From the agronomic perspective, *sh2* has worse seed quality (germination and field emergence), cold tolerance, root and stalk quality, ear type, plant color, husk protection, and disease resistance than *su1*, although these statements have not always been adequately tested (Tracy, 1997).

The main insect pest of sweet corn in temperate areas is corn borer [European corn borer (ECB) and other Lepidopteran species such as pink stem borer (PSB)] (Cordero et al., 1998; Malvar et al., 2002). Even in the Mediterranean area, where PSB is the major insect pest, ECB is still one of the most common insects in sweet corn ears (Velasco et al., 2002).

Variability for resistance to ECB among sweet corn inbreds has been known since the 1970s (Andrew and Carlson, 1976a). Resistance to ECB and PSB has been studied among *sugary1* genotypes (Malvar et al., 2002; Velasco et al., 1999, 2002). The inheritance of corn borer resistance in sweet corn is partial and

mainly additive (Velasco et al., 2002). However, little has been published about the resistance of *sh2* sweet corn to corn borers. In order to know if the knowledge and germplasm developed for improving corn borer resistance in *su1* sweet corn can be useful for improving super-sweet (*sh2*) corn, it is essential to determine if *sh2* affects corn borer resistance, compared to *su1*. For example, variability for antixenosis among sweet corn and popcorn has been inversely associated to sweetness, lack of pubescence, and earliness (Andrew and Carlson, 1976b).

Besides genetic factors, plant age and environment affect corn borer damage (Gardner et al., 2001). Plant age and pubescence are strongly affected by variation in vegetative phases. Maize has two vegetative phases, juvenile and adult. Juvenile and adult leaves, internodes, and axillary buds differ in anatomy and physiology (Bongard-Pierce et al., 1996; Lawson and Poethig, 1995; Poethig, 1988). Compared to juvenile leaves, adult leaves are wider and longer, have trichomes and lack epicuticular wax; adult nodes do not produce adventitious roots, and axillary buds may develop into ear primordia. A short juvenile phase has been associated with resistance to European corn borer, fall armyworm (*Spodoptera frugiperda* J.E. Smith), and southwestern corn borer (*Diatraea grandiosella* Dyar.) in maize because adult tissue is more resistant to insects than juvenile tissue (Abedon et al., 1999; Williams et al., 1998, 2000). However nothing has been published about vegetative phase transition in *sh2* germplasm.

Our objective was to compare relative susceptibility and vegetative phase transition between *su1* and *sh2* near-isogenic sweet corn inbreds, in order to indicate the applicability of *su1* research results to *sh2* breeding.

Received for publication 1 July 2003. Accepted for publication 24 June 2004. Research supported by the National Plan of Research and Development of Spain (Project Cod. AGL2000-0944) and Exema. Diputación Provincial de Pontevedra, Spain.

<sup>1</sup>To whom reprint requests should be addressed; phone: 34 986 854800; fax 34 986 841362; E-mail: previlla@mbg.cesga.es

## Materials and Methods

The sweet corn inbred lines 101t, C23, C40, C68, Ia453, Ia5125, and P39 were used by Soberalske and Andrew (1978, 1980) to develop near-isogenic inbred lines homozygous for either *sugary1* or *shrunken2*. The 14 near-isogenic inbreds were evaluated under corn borer infestation during 2000, 2001, and 2002 at Pontevedra (northwestern Spain). Trials followed a randomized complete-block design with four replications. Each plot consisted of two rows with 15 plants per row. Plants were spaced 0.21 m apart and rows were spaced 0.80 m, corresponding to a density of  $\approx 60,000$  plants/ha. Hills were over planted and thinned after emergence.

After half of the plants in each trial had flowered, five plants per plot were artificially infested with two ECB egg masses of  $\approx 40$  eggs. One of the egg masses was laid between the stem and the third leaf above the main ear, and the other mass at the third leaf below the main ear, in order to assure infestation in both juvenile and adult stem. ECB eggs were supplied by the Centre de Recherches de Poitou-Charentes (Institut National de la Recherche Agronomique, Le Magneraud, France). Infestation was made with ECB because this is a general pest in temperate regions worldwide, but natural infestation with PSB in Pontevedra is so high that usually 100% of the plants are infested by harvest (Cordero et al., 1998). At harvest time (October) in an adjacent field, 95% of maize plants were naturally infested. There were 0.8 PSB larvae/stem, and 0.2 ECB larvae/stem, averaged over 2000, 2001, and 2002.

In each plot, the following data were taken during plant development: days to pollen and silking, and number of adult leaves (without epicuticular wax) below the main ear. At dry seed stage, the ears infested with ECB were harvested. Main ear damage was rated according to a linear scale from 1 (= completely damaged ear) to 9 (= nondamaged ear). The infested stems were divided in juvenile and adult phases by the node corresponding to the last leaf with epicuticular wax. The proportion of juvenile stem was calculated as the ratio between length of juvenile stem and total length. In each part of the stem, the following traits were measured: proportion of adult and juvenile stem damaged (%), and number of PSB larvae and number of ECB larvae per meter of adult and juvenile stem.

Analyses of variance (ANOVAs) were made, including as sources of variation years, replications within years, genes (*su1* and *sh2*), inbred lines (101t, C23, C40, C68, Ia453, Ia5125, and P39), and the appropriate interactions. All sources of variation, except genes, inbred lines, and their appropriate interactions were considered random. Since near-isogenic inbreds are actually different genotypes, ANOVAs were performed, including years, replications within years, the 14 genotypes (101t*su1*, C23*su1*, C40*su1*, C68*su1*, Ia453*su1*, Ia5125*su1*, P39*su1*, 101t*sh2*, C23*sh2*, C40*sh2*, C68*sh2*, Ia453*sh2*, Ia5125*sh2*, and P39*sh2*), and the appropriate interactions. Besides, for the three traits that were recorded separately in juvenile and in adult tissue (stem damaged and number of PSB and ECB larvae), further ANOVAs were made using vegetative phase (juvenile and adult) as an additional source of variation. Therefore, the sources of variation in the ANOVAs for damage-related traits were years, replications within years, genes, vegetative phase (juvenile vs. adult), inbred lines, and the appropriate interactions. All sources of variation, except vegetative phase, genes, inbred lines, and their appropriate interactions were considered random. All analyses were performed using the SAS program (SAS Inst., Cary, N.C.).

## Results and Discussion

The combined ANOVAs, considering seven inbred lines and two genes per inbred, showed significant year  $\times$  gene  $\times$  inbred line and year  $\times$  gene interactions for four of the 13 traits (Table 1). The year  $\times$  inbred line and the gene  $\times$  inbred line interactions were not significant for any trait. Genes did not differ significantly for any trait and inbreds differed for four traits.

ANOVA considering the 14 genotypes (seven pairs of near-isogenic inbreds) showed significant year  $\times$  inbred interactions for half of the traits (data not shown). Inbreds were significantly different for days to pollen and silking, adult leaves, and PSB larvae in juvenile and adult stem, and for ECB larvae per ear (Table 2). Differences between *su1* and *sh2* isogenic pairs of each inbred were nonsignificant for most traits, particularly for ear-related traits (Table 2).

The *su1* inbreds tended to have fewer days to pollen and silking than their corresponding *sh2* versions; these differences were significant for four of the seven pairs of isogenic inbreds (Table 2). Concerning phase transition-related traits, the number of adult leaves below the main ear did not follow any particular pattern among *su1* and *sh2* inbreds, and the only significant difference was for C23, for which the *su1* version had more adult leaves below the main ear than the *sh2* version (Table 2). The ANOVAs for stem damage, using vegetative phase as an additional source of variation, showed significant differences between genes for PSB larvae (Table 3). Differences between juvenile and adult phases were not significant for any trait. Within the juvenile and adult phases, *su1* and *sh2* did not differ significantly for stem damage-related traits. When means across inbreds and vegetative phases were compared, *sh2* had similar tunnel length to *su1*.

Comparisons among the 14 inbreds showed that inbreds Ia5125*su1* and C40*su1* had lower proportion of tunnels in adult stem than 101t*su1*, 101t*sh2*, C23*su1*, and P39*su1*. Velasco et al. (1999) identified Ia5125 as one of the inbreds less damaged by PSB and P39 as one of the most damaged inbreds. These authors also concluded that resistance of sweet corn inbreds to PSB was partial and needed to be improved. Velasco et al. (2002) also found that differences in resistance among sweet corn inbreds for ECB were significant although not very clear, because most inbreds had intermediate resistance. Significant differences were found between *su1* and *sh2* strains of Ia453 and Ia5125 for number of PSB larvae in juvenile stem, and between strains of C40 for number of PSB larvae in adult stem, but other differences between pairs were nonsignificant (Table 2). However some trends could be observed, particularly *su1* strains had fewer PSB and more ECB larvae than *sh2* strains, and this relationship was similar for juvenile and adult stem (Table 3). The number of PSB larvae per meter of juvenile stem varied from zero in P39*sh2* to 5.8 in Ia5125*sh2* (Table 2). These data suggest that a reduction in juvenile tissue could result in fewer larvae per stem. However, an alternative explanation is that the proportion of juvenile tissue in fully developed plants is so small that the chances of finding larvae are very low. Differences among inbreds for number of larvae in adult stem were not important, and only C40*sh2* had significantly more larvae than any other inbred (Table 2). Nevertheless, the number of larvae cannot be used as an indication of resistance because larvae migrate and their abundance depends on the appropriate developmental stage of the plant.

In conclusion, *su1* inbreds flower earlier than *sh2* inbreds. Some relationship between corn borer damage, phase transition, and sweet corn genes (*sh2* or *su1*) can be postulated, although

Table 1. Mean squares from the analyses of variance for 13 traits in seven pairs of isogenic sweet corn inbred lines *sugary1* vs. *shrunken2*, grown for 3 years under artificial infestation with european corn borer (ECB) and natural infestation with pink stem borer (PSB) in northwestern Spain.

Genotype	df <sup>z</sup>	Days to	Days to	Adult	Juvenile	Juvenile stem			Adult stem			Ear		
		pollen	silking	leaves	stem	Tunnels	PSB	ECB	Tunnels	PSB	ECB	Damage	PSB	ECB
Years (Y)	2	126.95	172.33	7.33	703.43	3995.77	1.77**	33.58	4028.1	0.71	11.53	72.78	1.56	0.038
Reps. / Y	7	20.74*	28.40*	0.20	62.83	322.90**	3.71	10.01	696.9**	6.18	0.11	5.37**	1.13*	0.127*
Gene (G)	1	604.30	580.95	3.05	412.35	678.73	35.82	146.29	5.2	38.77	0.70	0.08	0.19	0.050
Y × G	2	103.21*	105.95*	2.70*	300.51	164.08	2.05	46.16	477.8*	2.44	3.77	3.52	0.04	0.114
Line (L)	6	61.25	65.79	9.06**	241.28*	2263.99	13.97	18.32	347.4*	16.76	3.87	7.83	0.32	0.118*
Y × L	12	61.38	31.75	1.58	55.92	989.12	4.92	37.63	111.0	9.95	3.57	2.90	0.58	0.031
G × L	6	16.22	7.64	1.80	81.77	455.84	10.64	40.07	167.3	23.57	3.46	3.10	0.12	0.099
Y × G × L	12	19.41*	19.66	0.71	146.34	1103.39	6.92	35.52	97.5	8.74*	3.23*	5.85**	0.57	0.034
Error	93	9.61	10.97	0.70	107.73	3701.56	4.12	38.75	143.4	3.73	0.35	1.25	0.46	0.056

<sup>z</sup>Degrees of freedom for Y × G × L were 11 for adult leaves, tunnels, PSB, and ECB in juvenile stem, and ear traits, and 10 for days to pollen and silking. Degrees of freedom for the error term were 92 for adult leaves, 91 for juvenile stem, and tunnels, PSB, and ECB in adult stem, and 90 for days to silking, 88 for ear traits, and 58 for tunnels, PSB, and ECB in juvenile stem.

\*,\*\*Significant at  $P = 0.05$  and  $0.01$ , respectively

Table 2. Means for 13 traits in juvenile and adult stem tissue and ears of seven pairs of isogenic sweet corn inbreds *sugary1* vs. *shrunken2*, grown for 3 years under artificial infestation with european corn borer (ECB) and intense natural infestation with pink stem borer (PSB) in northwestern Spain.

Genotype	Days to	Days to	Adult	Juvenile	Juvenile stem			Adult stem			Ear		
	pollen	silking	leaves	stem	Tunnels	PSB	ECB	Tunnels	PSB	ECB	Damage	PSB	ECB
	(d)	(d)	(no.)	(%)	(%)	(larvae/m)		(%)	(larvae/m)		(1-9) <sup>z</sup>	(no. larvae)	
I01tsh2	77 bc <sup>y</sup>	79 bc	2.3 def	17.7	62	3.1 bc	0.5	33 a	0.6 b	0.6	4.8	0.33	0.04 cd
I01tsul	72 ef	73 fg	1.8 fg	6.9	62	0.3 d	0.6	30 a	1.1 b	0.8	4.9	0.35	0.12 b-d
C23sh2	76 cd	78 cd	3.0 b-d	10.3	45	0.9 cd	0.0	27 ab	1.3 b	1.8	4.5	0.08	0.33 ab
C23sul	76 cd	77 c-e	4.1 a	8.1	38	0.7 d	7.1	33 a	1.2 b	0.6	5.9	0.30	0.11 b-d
C40sh2	78 bc	80 a-c	3.9 a	4.0	53	0.7 d	0.0	24 ab	6.3 a	0.1	6.5	0.88	0.06 cd
C40sul	76 cd	78 b-d	3.7 ab	8.4	25	2.0 b-d	0.2	18 b	0.4 b	0.7	5.6	0.25	0.41 a
C68sh2	80 ab	81 ab	3.0 b-d	9.2	42	1.1 b-d	0.0	24 ab	1.2 b	0.3	6.6	0.74	0.24 a-c
C68sul	73 de	75 d-f	3.6 ab	4.8	33	0.2 d	2.2	24 ab	0.7 b	0.4	6.1	0.46	0.23 a-c
I453sh2	77 bc	79 a-c	2.0 e-g	13.0	39	3.3 b	0.1	26 ab	1.0 b	0.6	6.2	0.55	0.24 a-c
I453sul	71 ef	74 e-g	2.6 c-e	7.7	47	0.1 d	0.7	31 a	1.1 b	1.0	5.9	0.40	0.19 a-d
I5125sh2	81 a	82 a	1.5 fg	19.4	58	5.8 a	0.0	27 ab	1.5 b	0.2	3.8	0.50	0.00 d
I5125sul	79 ab	80 a-c	1.2 g	18.9	40	1.6 b-d	4.2	17 b	0.3 b	0.4	4.9	0.40	0.03 cd
P39sh2	77 bc	78 cd	3.4 a-c	7.8	32	0.0 d	1.2	26 ab	1.3 b	0.5	4.9	0.35	0.10 cd
P39sul	70 f	72 g	3.4 ab	3.1	49	0.3 d	1.6	30 a	0.6 b	0.5	5.8	0.26	0.10 cd
LSD <sub>0.05</sub>	3	3	0.8			2.3		11	1.8				0.23

<sup>z</sup>Ear damage was estimated by using the scale 1 = completely damaged ear to 9 = no damaged ear.

<sup>y</sup>Mean separation within columns by LSD,  $P = 0.05$ .

Table 3. Means for three corn borer damage-related traits caused by pink stem borer (PSB) and european corn borer (ECB) larvae in juvenile and adult stem tissue and ears of seven pairs of isogenic sweet corn inbreds *sugary1* vs. *shrunken2*, grown for 3 years under artificial infestation with PSB in northwestern Spain.

Factor	Class	Tunnels	PSB	ECB
		(%)	(larvae/m)	
Phase	Juvenile	44	1.4	1.5
	Adult	27	1.2	0.6
Juvenile	<i>sh2</i>	46	2.1	0.4
	<i>su1</i>	42	0.6	2.6
Adult	<i>sh2</i>	27	1.7	0.5
	<i>su1</i>	26	0.8	0.6
Gene	<i>sh2</i>	36	1.9 a <sup>z</sup>	0.5
	<i>su1</i>	33	0.7 b	1.5

<sup>z</sup>Mean separation within columns and factors by LSD,  $P = 0.05$ .

environmental effects and interactions were large, reducing significant differences among classes for each of these factors. However, *su1* and *sh2* near-isogenic inbreds do not differ significantly for most of the traits related to phase transition and corn borer damage; particularly ear damage was nonsignificant different between *su1* and *sh2* inbreds. These results suggest that theoretical and practical results of sweet corn (*sugary1*) breeding

for corn borer resistance could be capitalized for super-sweet corn (*shrunken2*) breeding.

### Literature Cited

- Abedon, B.G., L.L. Darrah, and W.F. Tracy. 1999. Developmental changes associated with divergent selection for rind penetrometer resistance in the MoSCSSS maize synthetic. *Crop Sci.* 39:108-114.
- Andrew, R.H. and J.R. Carlson, Jr. 1976a. Evaluation of sweet corn inbreds for resistance for european corn borer. *J. Amer. Soc. Hort. Sci.* 101:97-99.
- Andrew, R.H. and J.R. Carlson, Jr. 1976b. Preference differences of egg laying european corn borer adults among maize genotypes. *Hort-Science* 11:143.
- Bongard-Pierce, D.K., M.M.S. Evans, and R.S. Poethig. 1996. Heteroblastic features of leaf anatomy in maize and their genetic regulation. *Intl. J. Plant. Sci.* 157:331-340.
- Cordero, A., R.A. Malvar, A. Butrón, P. Revilla, P. Velasco, and A. Ordás. 1998. Population dynamics and life-cycle of corn borers in south Atlantic European coast. *Maydica* 43:5-12.
- Gardner, J., M.P. Hoffmann, M.E. Smith, and M.G. Wright. 2001. Influence of plant age and genotype on resistance to european corn borer in sweet corn. *Maydica* 46:111-116.
- Lawson, E.J.R. and R.S. Poethig. 1995. Shoot development in plants: Time for a change. *Trends Genet.* 11:263-268.
- Malvar, R.A., P. Revilla, P. Velasco, M.E. Carrea, and A. Ordás. 2002.

- Insect damage to sweet corn hybrids in the south Atlantic European coast. *J. Amer. Soc. Hort. Sci.* 127:693–696.
- Poethig, R.S. 1988. Heterochronic mutations affecting shoot development in maize. *Genetics* 119:959–973.
- Soberalske, R.M. and R.H. Andrew. 1978. Gene effects on kernel moisture and sugars of near isogenic lines of sweet corn. *Crop Sci.* 18:743–746.
- Soberalske, R.M. and R.H. Andrew. 1980. Gene effects on water soluble polysaccharides and starch of near-isogenic lines of sweet corn. *Crop Sci.* 20:201–204.
- Tracy, W.F. 1997. History, genetics, and breeding of supersweet (*shrunk2*) corn. *Plant Breeding Rev.* 14:189–236.
- Tracy, W.F. 2000. Sweet corn, p.155–199. In: A.R. Hallauer (ed.). *Specialty corns*. 2nd ed. CRC, Boca Raton, Fla.
- Velasco, P., R.A. Malvar, A. Butrón, P. Revilla, A. Ordás. 1999. Ear feeding resistance of sweet corn inbreds to pink stem borer. *J. Amer. Soc. Hort. Sci.* 124:268–272.
- Velasco, P., P. Revilla, A. Butrón, B. Ordás, A. Ordás, and R.A. Malvar. 2002. Ear damage of sweet corn inbreds and their hybrids under corn borer infestation. *Crop Sci.* 42:724–729.
- Williams, W.P., F.M. Davis, P.M. Buckley, P.A. Hedin, G.T. Baker, and D.S. Luthe. 1998. Factors associated with resistance to fall armyworm (Lepidoptera:Noctuidae) and southwestern corn borer (Lepidoptera:Crambidae) in corn at different vegetative stages. *J. Econ. Entomol.* 91:1471–1480.
- Williams, W.P., P.M. Buckley, and F.M. Davis. 2000. Vegetative phase change in maize and its association with resistance to fall armyworm. *Maydica* 45:215–219.