

Sweet Corn Crop Nitrogen Status Evaluation by Stalk Testing

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Abstract. Sweet corn (*Zea mays* L.) growers evaluating new practices for N management, such as the presidedress soil nitrate test (PSNT), are interested in relating observations about crop performance at time of harvest to their N fertility program. For this purpose, the concentration of nitrogen (N) in the lower portion of sweet corn stalks was examined on the day of harvest as a basis for evaluating the crop N status. Sweet corn stalk tissue was collected from N-rate experiments by cutting a stalk section at 15 and 35 cm aboveground and removing leaf material from the resulting 20-cm segment. Samples were dried and analyzed for total Kjeldahl N. Relationships between crop yield and stalk N concentration indicated that concentrations <11 g·kg⁻¹ are N deficient and underfertilized; N concentrations between 11 and 16.5 g·kg⁻¹ are marginally deficient; and between 16.5 and 21 g·kg⁻¹ the N status is optimum. Concentrations of N >21 g·kg⁻¹ are above optimum and indicate that sweet corn was overfertilized with N. When soil nitrate concentrations (PSNT >25 mg NO₃-N per kilogram) indicated sufficient N at time of sidedressing, stalk N concentrations generally indicated N sufficiency at harvest.

Plant and soil tests provide key information for nutrient management planning that serves to minimize environmental impact and maintains the economic viability of crop production. The presidedress soil nitrate test (PSNT) and end-of-season stalk N test are testing procedures that were originally developed to improve N fertility management for field corn production (Binford et al., 1990; Magdoff, 1991). Use of the PSNT has already been successfully extended to sweet corn (Heckman et al., 1995) and other vegetable crops (Hartz et al., 2000; Heckman et al., 2002). The development of a stalk N testing technique for sweet corn could direct growers to improved N management as it has for field corn (Hooker and Morris, 1999; Sims et al., 1995).

Sweet corn growers evaluating levels of N application or new N management practices, such as the PSNT, may be particularly inter-

ested in whether their N fertility program was appropriate. This fits the function of the stalk N test, which in the case of field corn, can validate the current N fertility program or can determine if the N application was insufficient or excessive. Collecting stalk samples at time of sweet corn harvest should allow observations about current crop performance to be related to the sufficiency of the N fertility program. Furthermore, this should allow growers to learn from that experience, and to direct efforts to improve N management in future growing seasons.

Corn stalk N testing is based on the assumption that “nitrate (NO₃⁻) tends to accumulate in the lower portion of mature corn stalks when abundant amounts of N are available in soils” (Binford et al., 1990; Hanway and Englehorn, 1958; Hoffer, 1926). Sweet corn, which is harvested at a physiologically immature growth stage, may behave differently than field corn with respect to NO₃-N accumulations in stalk tissue. A recent study by Hooker and Morris (1999), however, showed that stalk NO₃-N testing was an effective method of defining excess N availability to silage corn, which is harvested about 3 weeks before physiological maturity. In very young corn plants (4 weeks after emergence) stalk NO₃-N was, however, found to be a poor predictor of soil N availability as a result of the sensitivity of stalk NO₃-N concentrations at this early growth stage to the influence of solar radiation (Iversen et al., 1985) and soil moisture availability (Fox

et al., 1989). Because sweet corn is generally irrigated, the effect of soil moisture on stalk NO₃-N concentration may be less of a factor than for field corn. Stalk total Kjeldahl N (TKN) may be a better indicator of sweet corn N status because it measures both NO₃-N and reduced-N in the stalk tissue, and it may minimize the importance of solar radiation on NO₃ reduction.

This study was conducted to evaluate stalk N testing as a means to determine if the N management program used for sweet corn provided insufficient, optimum, or excessive plant-available N. Stalk N testing was evaluated both by measuring TKN and NO₃-N concentrations. The relationship between the PSNT and stalk N test results was examined. The results of stalk N testing both from sweet corn cultivar trials and a survey of commercial sweet corn fields are also reported.

Materials and Methods

This study used 24 of the 61 N-response trials that were conducted in New Jersey from 1991 to 1994 to calibrate the PSNT for sweet corn (Heckman et al., 1995). At planting, starter fertilizer was applied with N at 22 kg·ha⁻¹, and additional N was sidedressed at 0, 45, 90, 135, and 180 kg·ha⁻¹. The PSNT soil sampling was performed when plants were 30 cm tall at the whorl by collecting eight cores (2 cm in diameter × 30 cm deep) between the rows of each control plot. Harvesting procedures were as previously described (Heckman et al., 1995). Two additional trials were conducted in 1996 using the above methods. Irrigation was applied at all sites as needed to avoid drought stress.

Stalk samples were collected from the sweet corn N response trials using procedures similar to those for field corn (Binford et al., 1990) except that in this study the stalk samples were collected on the day of harvest for sweet corn. The 20-cm segments were collected by cutting the stalk at 15 and 35 cm aboveground from 10 randomly selected plants within the two center rows of six row plots. All leaf tissues were removed from the stalk samples. The samples were dried at 70 °C and ground in a Wiley mill to pass a 1-mm screen. Stalk NO₃-N concentrations were determined by extraction with 2 M KCL (1:10, w:v) and analyzed for NO₃-N colorimetrically using a Technicon Auto-analyzer (Technicon, Tarrytown, N.Y.). Total Kjeldahl N was determined by using the permanganate-reduced Fe method to include NO₃-N (Bremner and Mulvaney, 1982).

The Rutgers integrated crop management program began conducting the stalk N test for commercial sweet corn growers in 1997. Between 1997 and 1999, stalk samples were collected from 55 fields and the data were summarized as a survey of the N status of New Jersey sweet corn acreage. In addition to the sweet corn N response trials, stalk samples were also collected from cultivar trials that were conducted during 1998 to 2000. The cultivar trials were conducted using a randomized complete-block design with four replications. They were established with a population

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of 64,000 plants/ha and uniformly fertilized with N at 180 kg·ha⁻¹. The cultivars Calico Belle, Brilliance Film Coat, Sensor, and Silverado were sampled for stalk N testing each year at time of harvest. Concentrations of TKN and NO₃-N in stalk samples were compared among cultivars by ANOVA.

Relative yields were calculated as the yield of marketable ears produced with N at 0, 45, 90, or 135 kg·ha⁻¹ sidedress N, expressed as a percentage of the yield observed for N at the 180 kg·ha⁻¹ rate of applied sidedress N. The relationship between stalk TKN or NO₃-N concentration and relative yield was examined using Cate-Nelson analysis (Cate and Nelson, 1965) by partitioning data into N-responsive and nonresponsive N rates. Setting the horizontal line at an acceptable relative yield level of 92% modified this procedure. The vertical line drawn according to Cate-Nelson analysis was used to determine the critical stalk TKN and NO₃-N concentration, which effectively establishes the lower value of the optimal range. The relationships between the PSNT NO₃-N concentration and the stalk TKN or NO₃-N concentration were also examined by Cate-Nelson analysis.

Results and Discussion

The relationships between relative yield and stalk TKN or NO₃-N concentration (Fig. 1 a and b) suggest that stalk testing for sweet corn at time of crop harvest is useful for diagnostic purposes. Clearly, very low concentrations of TKN or NO₃-N in stalk tissue are associated with low relative yields and very high concentrations are associated with near maximum yield. Cate-Nelson graphic analysis for the database using TKN or NO₃-N results in critical levels of TKN at 16.5 g·kg⁻¹ or NO₃-N at 11 g·kg⁻¹. We selected these critical levels to minimize the number of outliers but also to ensure that not more than 5% of the errors would fall into the lower right quadrant. This balance of errors is similar to that used by Fox et al. (2001) to ensure acceptability of the test to growers who have a low tolerance of incorrect predictions of N sufficiency when, in fact, N supply was deficient. There were nearly the same number of errors in the upper left quadrant for the stalk test measured as TKN (11.7%) as for NO₃-N (13.0%). Error rates suggest that stalk testing using TKN concentrations may be better at predicting sweet corn N sufficiency than NO₃-N.

The linear correlation between stalk TKN concentrations and stalk NO₃-N concentrations had an r² of 0.67, and the regression equation was:

$$\text{NO}_3\text{-N} = 0.65 \text{ TKN} - 1.6$$

This equation indicates that as stalk tissue TKN concentration increases, an increasing percentage of this tissue N is present in the form of NO₃-N. The critical levels for stalk NO₃-N concentration established by Binford et al. (1990) for field corn was 0.25 g·kg⁻¹, and by Hooker and Morris for silage corn was 0.5 g·kg⁻¹. These values are substantially lower

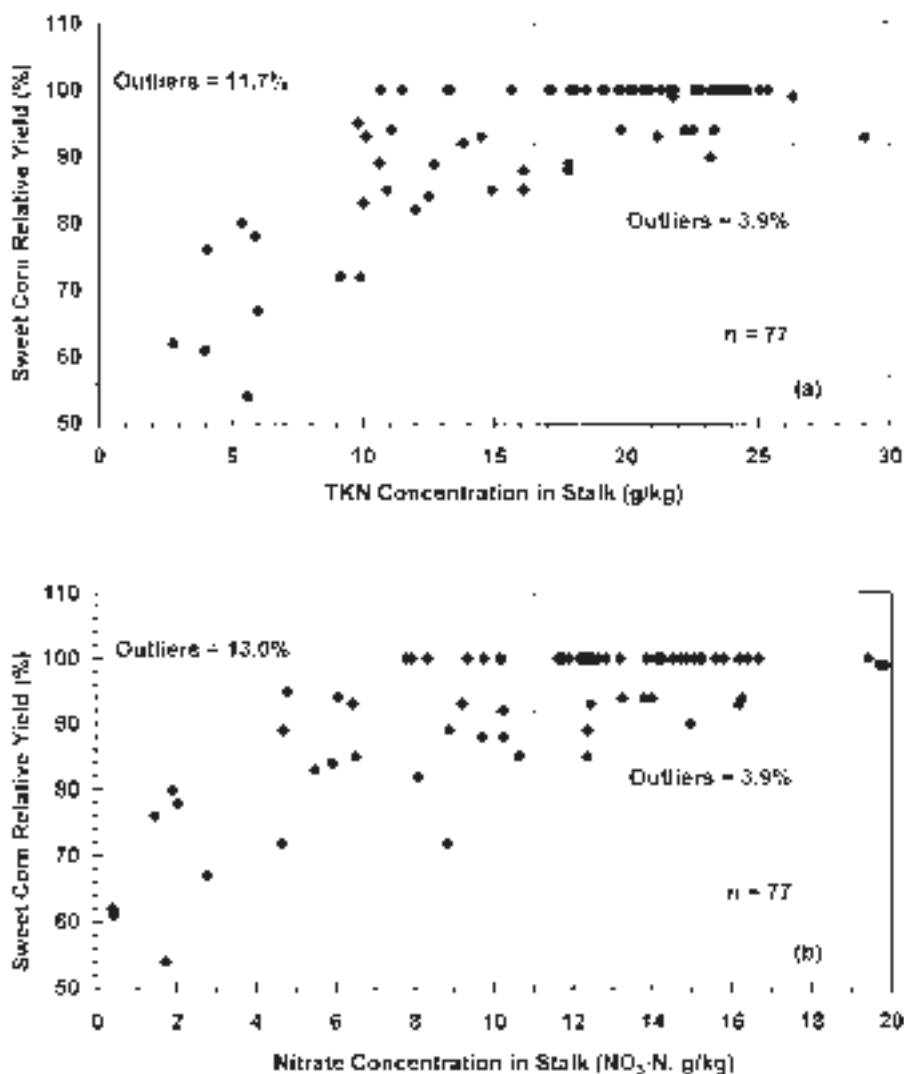


Fig. 1. Relationship between at-harvest stalk total Kjeldahl N (a) or NO₃-N (b) concentrations and relative yield of sweet corn.

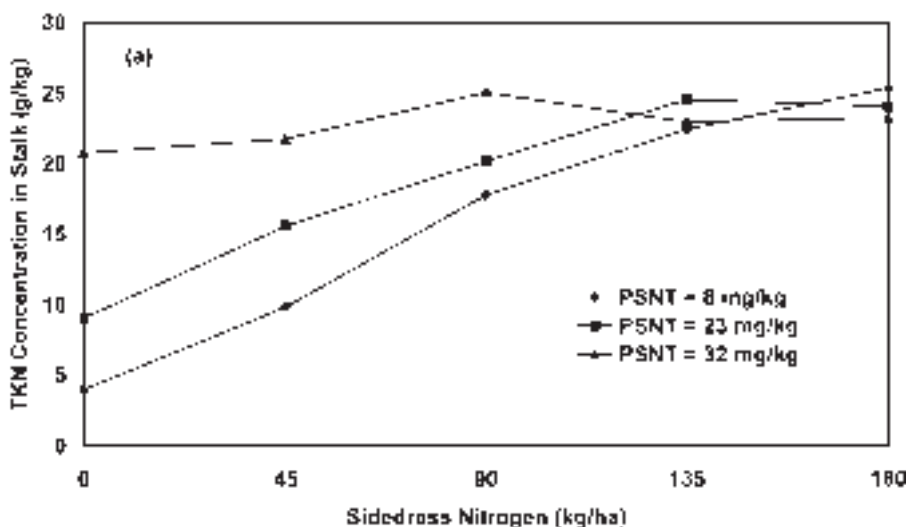


Fig. 2. Concentrations of total Kjeldahl N (TKN) in sweet corn stalk tissue at harvest in response to N fertilizer rate at three field sites with different PSNT levels.

than the $\text{NO}_3\text{-N}$ at 11 $\text{g}\cdot\text{kg}^{-1}$ critical level that was established in our study for sweet corn sampled at time of fresh harvest. The magnitude of this difference is probably largely a result of field corn harvest occurring at a much later physiological growth stage such that much of the $\text{NO}_3\text{-N}$ stored in the lower stalk tissue has been remobilized for grain fill. Because sweet corn harvest and stalk sampling occurs at an earlier physiological growth stage than even silage, remobilization of stalk $\text{NO}_3\text{-N}$ is apparently limited for sweet corn. This may account for the higher error rate found for sweet corn stalk $\text{NO}_3\text{-N}$ critical levels as compared to the stalk TKN test or the lower error rates of the field corn stalk $\text{NO}_3\text{-N}$ test (Fox et al., 2001).

Sweet corn stalk tissue TKN concentrations increase with N fertilizer rate, but this is also influenced by the level of $\text{NO}_3\text{-N}$ available in soil as measured by the PSNT (Fig. 2). A Cate-Nelson graphical analysis of the relationship between PSNT soil $\text{NO}_3\text{-N}$ concentrations and stalk TKN concentrations show that most of the 24 field sites were partitioned into two quadrants (Fig. 3). Data points in the lower left quadrant represent field sites that test below the PSNT critical level of $\text{NO}_3\text{-N}$ at 25 $\text{mg}\cdot\text{kg}^{-1}$ (Heckman et al., 1995) at the appropriate time for sidedressing, but when not given sidedress N, result in stalk samples collected at harvest time that indicate N deficiency. Data points in the upper right quadrant represent field sites that test above the PSNT critical level of 25 $\text{mg}\cdot\text{kg}^{-1}$ in soil and, when not given sidedress N, result in stalk samples collected at harvest that test primarily above the critical level of TKN at 16.5 $\text{g}\cdot\text{kg}^{-1}$ (established in Fig. 1). Overall there is good agreement between sufficiency predictions as indicated by the PSNT and by stalk TKN testing.

The difficulty with attempts to establish a critical stalk TKN concentration is that the transition between deficiency and sufficiency is not sharply defined. Sweet corn cultivar differences in stalk TKN or $\text{NO}_3\text{-N}$ concentration may partially account for this (Table 1). In sweet corn cultivar trials conducted over a 3-year period, the cultivar Sensor had significantly lower stalk TKN and $\text{NO}_3\text{-N}$ concentrations than the other three cultivars.

Based on the current study findings, we propose the interpretations as listed in Table 2 for stalk N testing in sweet corn. Although interpretations are provided for both TKN and $\text{NO}_3\text{-N}$ concentrations, TKN is the preferred measurement because it made fewer incorrect predictions about sweet corn N status.

The usefulness of stalk N testing was shown by the results of stalk samples collected on the day of harvest from 55 commercial sweet corn fields in New Jersey during the summers of 1997 to 1999. Stalk TKN concentrations ranged from 6 to 37 $\text{g}\cdot\text{kg}^{-1}$. Both the mean and median stalk TKN concentrations were 19 $\text{g}\cdot\text{kg}^{-1}$. Based on the interpretations in Table 1, 18% of the samples would be classified as N deficient, 16% as marginal, 31% as optimal and 35% as excessive in N status. Providing this information to sweet corn producers enables them to make adjustments in their N fertility program

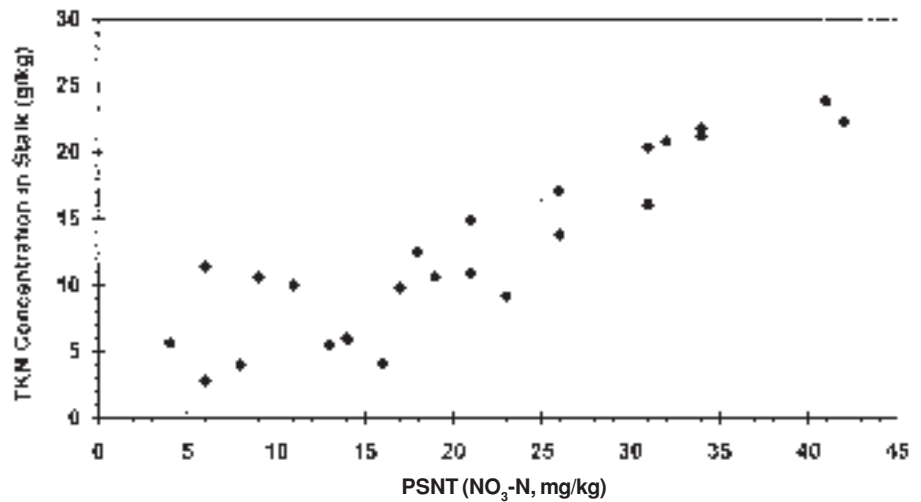


Fig. 3. Relationship between the presidedress soil nitrate test and the stalk total Kjeldahl N concentration of sweet corn at harvest.

Table 1. Concentrations of total Kjeldahl N and $\text{NO}_3\text{-N}$ in stalk samples collected from sweet corn cultivar trials at time of harvest, 1998–2000.

Cultivar	Total	
	Kjeldahl N	NO ₃ -N
	----- g·kg ⁻¹ -----	
1998		
Calico Belle	16.9	9.6
Brilliance Film Coat	16.8	10.5
Sensor	11.5	6.1
Silverado	17.2	7.8
LSD _{0.05}	1.5	1.2
1999		
Calico Belle	18.8	10.6
Brilliance Film Coat	20.7	12.9
Sensor	13.7	7.1
Silverado	20.9	10.0
LSD _{0.05}	1.8	1.5
2000		
Calico Belle	19.2	9.7
Brilliance Film Coat	16.3	9.9
Sensor	13.1	6.1
Silverado	15.2	7.1
LSD _{0.05}	2.7	1.0
<i>P</i> > <i>F</i>		
Cultivar (C)	0.0001	0.0001
Year (Y)	0.0001	0.0001
C × Y	0.01	0.16

Table 2. Proposed Interpretations of the at-harvest stalk N test for sweet corn.

Total Kjeldahl N	$\text{NO}_3\text{-N}$	Interpretation
g·kg ⁻¹		
<11.0	<6.0	N-deficient, underfertilized
11.0 to 16.5	6.0 to 11.0	Marginal, may be underfertilized
16.5 to 21.0	11.0 to 14.0	Optimal range, N sufficient
>21.0	>14.0	Excessive, overfertilized

in subsequent growing seasons. Thus, on average, nearly twice as many growers currently using the Stalk TKN test would be encouraged to reduce N application rates as would be encouraged to increase N application. Just as the end-of-season stalk N test for field corn has been found to be a useful tool to improve N management (Binford et al., 1992; Fox et al., 2001; Hooker and Morris, 1999; Sims et

al., 1995) we believe stalk testing for sweet corn, even with the uncertainty of the proposed interpretations, will provide information to help producers better manage N.

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