Capturing the Diversity of Wild *Malus orientalis* from Georgia, Armenia, Russia, and Turkey

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ABSTRACT. Seeds from wild *Malus orientalis* trees were collected during explorations to Armenia (2001, 2002), Georgia (2004), Turkey (1999), and Russia (1998). Seedling orchards with between eight and 171 individuals from each collection location were established at the U.S. Department of Agriculture–Agricultural Research Service Plant Genetic Resources Unit (PGRU) in Geneva, NY. Genotypic (seven microsatellite markers) and disease resistance data were collected for the 776 *M. orientalis* trees. The genetic diversity of the 280 individuals from Armenia and Georgia was compared with data previously published for the *M. orientalis* individuals from Russia and Turkey. A total of 106 alleles were identified in the trees from Georgia and Armenia and the average gene diversity ranged from 0.47 to 0.85 per locus. The genetic differentiation among sampling locations was greater than that found between the two countries. Six individuals from Armenia exhibited resistance to fire blight (*Erwinia amylovora*), apple scab (*Venturia inaequalis*), and cedar apple rust (*Gymnosporangium juniperi-virginianae*). The allelic richness across all loci in the individuals from Armenia and Georgia was statistically the same as that across all loci in the individuals from Russia and Turkey. A core set of 27 trees was selected to capture 93% of the alleles represented by the entire PGRU collection of 776 *M. orientalis* trees. This core set representing all four countries was selected based on genotypic data using a modified maximization algorithm. The trees selected for the *M. orientalis* core collection will be added to the main field collection at the PGRU.

Wild *Malus orientalis* and *Malus sieversii* populations from the Middle East and Central Asia may be primary progenitor species to the domesticated apple (*Malus ×domestica*) (Buttner, 2001; Watkins, 1995). Populations of *M. sieversii* in Kazakhstan include individuals that most closely resemble the dessert apples consumed today (Luby et al., 2001). During the domestication process, apples that were brought from Central Asia to Europe may have undergone some hybridization with *M. orientalis* trees found wild in regions surrounding the Black Sea (Buttner, 2001; Ercisli, 2004).

Plant explorations have targeted wild *M. sieversii* and *M. orientalis* with the goal of enhanced germplasm representation and characterization in gene banks (Luby et al., 2001). Materials in gene banks are accessible to plant breeders. However, the quantity and extent of accessions available can be overwhelming in light of the amount of extant diversity. Genes from wild

crop relatives can most easily be incorporated into breeding programs using either marker-assisted breeding or through the use of transformation. Both of these methods are dependent on having materials that possess the traits of interest. Development and use of core collections whereby large numbers of accessions are represented by a subset of significantly fewer individuals makes germplasm characterization more feasible and provides an entry point into a more complex collection. Maximization strategies (Gouesnard et al., 2001) have been successful for the development of core collections in *Arabidopsis* (McKhann et al., 2004), *Medicago* (Ellwood et al., 2006; Ronfort et al., 2006), *Cacao* (Zhang et al., 2009), and *Glycine* (Tavaud-Pirra et al., 2009), and *Zea* (Franco et al., 2006).

Malus orientalis germplasm possesses traits that are of potential interest to breeding programs (Aldwinckle et al., 2002). We previously identified 20 individuals that exhibited resistance to apple scab, fire blight, and cedar apple rust from Turkey and Russia (Volk et al., 2008). In addition, Georgian wild *M. orientalis* is valued as a drought- and rodent-resistant rootstock for production of propagated trees of named fruiting cultivars (J. Postman, personal communication). Individual genotypes identified to have resistance to disease can be used both directly in breeding programs and as a starting point for allele mining in related lineages within the collection (Borejsza-Wysocka et al., 1999; Gianfranceschi et al., 1996; Norelli et al., 1998, 2003).

The *Malus* field collection at the U.S. Department of Agriculture (USDA)–Agricultural Research Service (ARS)–National Plant Germplasm System (NPGS)–Plant Genetic

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Resources Unit (PGRU) in Geneva, NY, field collection includes 776 *M. orientalis* genotypes collected from Russia, Turkey, Armenia, and Georgia. Previously, we used seven microsatellite markers to determine the levels of genetic diversity and disease resistance in the individuals from Russia and Turkey (Volk et al., 2008). In this work, we compare the diversity of the more recently acquired *M. orientalis* individuals from Georgia and Armenia with that of the individuals from Russia and Turkey. Using the entire *M. orientalis* data set of 776 individuals, we propose a core collection of 27 individuals that captures 93% of the allelic diversity of the PGRU *M. orientalis*. We recommend the use of this core collection for future detailed phenotypic evaluations of *M. orientalis*.

Materials and Methods

PLANT MATERIALS. Seeds of *M. orientalis* trees in Russia (28 accessions in 1998), Turkey (57 accessions in 1999), Armenia (seven accessions in 2001, 2002), and Georgia (nine accessions in 2004) were collected during NPGS-sponsored explorations (Table 1; Fig. 1). Accessions from Georgia and Armenia that were collected within the same year and within a 75-km radius were assigned to locations (Fig. 1). Seeds within an accession from the Georgia and Armenia collections were sometimes collected from several trees and thus are not necessarily maternally related. Seeds were established in the greenhouse and then planted in field collections at the PGRU. Leaf tissue was collected from each individual and sent to the USDA-ARS–National Center for Genetic Resources Preservation in Ft. Collins, CO, and kept at –80 °C for DNA extraction.

DISEASE RESISTANCE. Potted seedling plants were inoculated with mixed conidial suspensions of North American Races 1 to 5 of *V. inaequalis*. Plants were incubated at 100% relative humidity in a mist chamber and infection severity was evaluated (number and types of lesions) after 4 weeks' growth in the greenhouse. Seedlings scored as resistant to apple scab had no symptoms, pinpoint lesions, chlorotic lesions, or

necrotic lesions. Seedlings that produced conidia and exhibited signs of sporulation were considered susceptible.

Cedar apple rust susceptibility was determined based on the presence of pycnidia on seedling tissues 8 weeks after inoculation with basidiospores of *Gymnosporangium juniperi-virginianae*.

Greenhouse plants were inoculated with *Erwinia amylovora* strain Ea273. The two youngest actively growing leaves were inoculated by transverse bisection with a pair of scissors dipped in *E. amylovora* (1×10^9 cfu/mL). Necrotic lesion lengths were expressed as a percentage of the current season's shoot length and plants with less than 20% of shoot length blighted were characterized as resistant.

MOLECULAR ANALYSIS. Genomic DNA was extracted from 280 duplicate *M. orientalis* leaf tissue samples using DNeasy 96 plant kits (Qiagen, Valencia, CA) as previously described (Volk et al., 2008). Seven microsatellite regions were amplified using primers for unlinked microsatellite regions (GD12, GD15, GD96, GD100, GD142, GD147, GD162) (Hemmat et al., 2003; Hokanson et al., 1998). Cultivars Golden Delicious, Law Rome Beauty, and Cox's Orange Pippin were standard cultivar controls. Fragments were amplified as described previously (Volk et al., 2005). Replicate products were separated on denatured acrylamide sequencing gels (model 4200; LI-COR, Lincoln, NE) and manually scored using the Saga™ software (LI-COR). Data were considered "missing" if allele calls were not identical for the duplicate DNA samples.

MOLECULAR DATA ANALYSIS. Genetic diversity was assessed for genotypes of *M. orientalis* individuals grown from seeds collected in Georgia and Armenia by calculating indices of average gene diversity and allelic richness (Goudet, 1995). Average differentiation (F_{st}) between *M. orientalis* samples collected in Georgia and Armenia as well as between collection locations within countries was estimated using the software package GDA (Lewis and Zaykin, 2001).

A core collection for *M. orientalis* was identified using the genotypic data for all *M. orientalis* accessions collected in

Table 1. The numbers of accessions and individuals for each collection region in Armenia, Georgia, Turkey, and Russia.^z

Country	Location ^y	Year	Total accessions (no.)	Total individuals (no.)	Resistant individuals (no.) ^x			
					Apple scab	Fire blight	Cedar apple rust	Apple scab, fire blight, and cedar apple rust
Georgia	GA	2004	1	8	7	1	N/A	N/A
Georgia	GB	2004	3	62	55 (2)	36	N/A	N/A
Georgia	GC	2004	1	14	14	1	N/A	N/A
Georgia	GD	2004	3	58	57	11	N/A	N/A
Georgia	GE	2004	1	17	17	4	N/A	N/A
Armenia	AA	2002	4	80	18	56 (13)	23 (48)	4
Armenia	AB	2001	2	27	20	12 (10)	11 (14)	2
Armenia	AC	2002	1	14	2	4 (7)	0 (4)	0
Russia	RA	1998	6	27	12	19	3	0
Russia	RB	1998	22	171	137	94 (6)	15 (3)	0
Turkey	TA	1999	5	44	31	13 (1)	22 (1)	0
Turkey	TB	1999	22	102	3	84 (3)	11 (63)	0
Turkey	TC	1999	17	93	39	47 (15)	6 (24)	0
Turkey	TD	1999	13	59	30	26 (10)	16 (12)	0
			101	776				

^zNumbers of individuals exhibiting apple scab, fire blight, and cedar apple rust resistance are provided.

^yLocation codes per Figure 1.

^xNumber of individuals for which disease resistance data were not available is provided in parentheses.

N/A = not available.

Fig. 1. Map of countries from which *Malus orientalis* seeds were collected. Individual seedlings from locations within these four countries were included in analyses to identify a core collection.

Georgia, Armenia, Russia, and Turkey. Sample identification and microsatellite data for the Russian and Turkish samples was published previously (Volk et al., 2008). The number and selection of individuals for inclusion in the *M. orientalis* core set was identified using a modified MSTRAT algorithm (Gouesnard et al., 2001) that allows alleles within a diploid genotype to be considered jointly during core assembly and standardizes the contributions of individual loci to the overall diversity during core optimization (C. Richards, personal communication).

Results

Our data suggest that *M. orientalis* accessions from Georgia and Armenia contribute valuable novel germplasm to the NPGS collection. Disease resistance is a key attribute desired in apple breeding programs. A total of six individuals from Armenia were resistant to apple scab, fire blight, and cedar apple rust (Table 1). Two were introduced to the NPGS from an Armenia exploration in 2001 (GMAL 4597.f and GMAL 4598.a) and four individuals were introduced from Armenia in 2002: GMAL 4601.a, GMAL 4601.i, GMAL 4601.p, and GMAL 4602.g (Table 1). Although cedar apple rust resistance was not determined for individuals resulting from seeds collected in Georgia, 49 individuals from Georgia were resistant to both fire blight and apple scab (data not shown).

Genetic diversity, based on microsatellite analyses, was assessed for a total of 280 individuals from 16 accessions from Georgia and Armenia (Table 1). These data are complementary to that previously reported for 496 *M. orientalis* individuals from Russia and Turkey. A total of 106 alleles were identified within the 280 individuals sampled from Armenia and Georgia (Table 2). The number of alleles from each of the seven loci varied from two (GD15) to 22 (GD12) with five of the seven loci each yielding 16 or more alleles. The average gene diversity of the loci ranged from 0.47 (GD15) to 0.85 (GD96).

Allelic richness was calculated for *M. orientalis* individuals collected from Armenia and Georgia and classified according to location (Fig. 1; Table 3). Overall, the samples that originated in Armenia had more private alleles than those from Georgia, but allelic richness levels were comparable (Table 3). Genetic differentiation (F_{st}) was low between the countries of Armenia and Georgia. The level of differentiation among sampling locations (nested within countries) was greater than that found between countries (Table 4).

CORE COLLECTION. All 776 *M. orientalis* individuals available from Georgia, Armenia, Russia, and Turkey were considered for inclusion in the *M. orientalis* core set. This set was selected based on the alleles identified for seven microsatellite markers, because the disease resistance data were incomplete. The methods used to select the core set were modified from

Table 2. Descriptive statistics for each of seven microsatellite loci based on genotypic data from 280 *Malus orientalis* individuals that originated from seeds collected in Armenia and Georgia.

Marker	Total alleles (no.)	Avg gene diversity	Minimum allele size (bp)	Maximum allele size (bp)
GD12	22	0.80	146	192
GD15	2	0.47	142	145
GD96	21	0.85	150	198
GD100	8	0.64	221	235
GD142	17	0.79	126	164
GD147	16	0.72	117	157
GD162	20	0.80	212	258
Total	106			
Avg (no./locus)	15.1	0.72		

Table 3. Allelic richness was calculated for 280 *Malus orientalis* individuals collected from Armenia and Georgia and grouped according to collection location.^z

Locus	Georgia					Armenia		
	GA	GB	GC	GD	GE	AA	AB	AC
GD12	5.00	6.62	4.33	6.55	5.61	7.05	6.99	4.23
GD15	2.00	1.99	2.00	2.00	2.00	2.00	2.00	2.00
GD96	7.60	7.30	4.49	6.45	6.53	7.53	7.51	4.73
GD100	2.00	4.45	3.35	4.69	5.30	5.08	4.60	1.86
GD142	5.24	5.87	2.64	7.14	6.33	7.01	6.00	4.72
GD147	3.49	6.07	4.79	5.57	5.64	5.80	4.87	2.68
GD162	3.00	5.94	3.96	6.66	6.19	6.25	7.27	4.60
Mean/location	4.05	5.46	3.65	5.58	5.37	5.82	5.60	3.54
Private alleles	2	1	0	1	1	6	3	2
N	9	62	14	58	17	68	39	14

^zLocation codes per Figure 1.

Table 4. Levels of *Malus orientalis* genetic differentiation as estimated by average differentiation (F_{st} ; with confidence intervals) calculated between Armenia and Georgia (country) and within Armenia (three locations) and Georgia (five locations).

Source	F_{st}	F_{st} confidence interval
Country	0.005	−0.02–0.03
Locations (country)	0.09	0.06–0.13

methods used for previous *M. sieversii* core set determinations (Richards et al., 2009; Volk et al., 2005).

The proposed *M. orientalis* core set of 27 trees includes individuals from each of the four countries (Table 5). Individuals selected based on genotypic data obtained for the set of seven microsatellite markers represent 10 of the 14 collection locations. Two individuals in the core collection are resistant to apple scab, fire blight, and cedar apple rust. Four additional individuals in the core collection are resistant to both apple scab and fire blight (Table 5).

The 27 individuals capture 123, or 93%, of the total 132 alleles in the entire *M. orientalis* data set. Allele frequencies were plotted to compare representation in the core versus the entire *M. orientalis* datasets (Fig. 2). The frequency relationship was linear with a slope of 0.94 and an r^2 of 0.84, suggesting that the modified core selection process was representative.

Discussion

Malus orientalis is likely a progenitor species of domesticated apples that has yet to be actively included in breeding

programs. Candidate genes from *Malus floribunda* and *M. sieversii* that offer apple scab resistance have been identified (Boudichevskaia et al., 2009; Bus et al., 2005; Szankowski et al., 2009). *M. orientalis* individuals exhibiting apple scab resistance may offer novel candidate genes that could be of use to breeding programs, particularly as major resistance genes are overcome by the fungus (Soufflet-Freslon et al., 2008). The 20 disease-resistant individuals previously identified from Russia and Turkey (Volk et al., 2008) and the six disease-resistant Armenian individuals identified here may be of particular interest to breeding programs because they offer resistance to apple scab, fire blight as well as cedar apple rust. The proposed core collection includes two individuals with resistance to all three diseases.

The 280 individuals from Armenia and Georgia complement the diversity described for the 496 NPGS Turkish and Russian *M. orientalis* trees. The 106 total alleles identified from Armenia and Georgia was similar in number to the 126 alleles identified from Russia and

Turkey (Volk et al., 2008). When private alleles were identified for the entire *M. orientalis* collection, two alleles were unique to Armenia, one allele was unique to Georgia, and three and 10 alleles were unique to Russia and Turkey, respectively. In the combined *M. orientalis* data set, the allelic richness across all loci in the individuals from Armenia and Georgia (15.07) was statistically the same as that across all loci in the individuals from Russia and Turkey (17.13). Allelic richness was selected as the diversity measurement used to compare among locations to account for the differences between sample sizes. Genetic differentiation was higher among locations within Armenia and Georgia (0.09) relative to those among locations in Turkey and Russia (0.046). Genetic differentiation between the combined groups of individuals from Georgia–Armenia and Turkey–Russia was small but significant (F_{st} 0.021; bootstrapped 95% confidence interval, 0.014 to 0.031). For the Georgian and Armenian samples, the original seeds were not separated by mother; therefore, sampling was not performed in a manner such that estimates about family structure could be made.

Three core collections with a total number of 112 individuals have previously been proposed to represent the diversity of 949 *M. sieversii* accessions from Kazakhstan (Richards et al., 2009; Volk et al., 2005). These core collections were identified based on both phenotypic and genotypic data using the original MSTRAT algorithm (Gouesnard et al., 2001). Based on our findings, this algorithm likely overestimated the number of individuals needed to capture diversity because alleles at co-dominant loci were not considered jointly. The modified core assembly algorithm, which treats loci equally and codominant data correctly, resulted in a core for *M. orientalis* that was

Table 5. Passport information and disease resistance of 27 individuals included in *Malus orientalis* core collection.

Accession no.	Country	Location ²	Collection year	North latitude	East longitude	Elevation (m)	Site details	Apple scab resistance	Fire blight resistance	Cedar apple rust resistance
GMAL 4469.m	Russia	RA	1998	44.92	38	100	5 km south of Krymsk	Resistant	Susceptible	Resistant
GMAL 4471.h	Russia	RA	1998	44.92	38	100	5 km south of Krymsk	Resistant	Susceptible	Susceptible
GMAL 4472.p	Russia	RA	1998	44.92	38	100	5 km south of Krymsk	Susceptible	Resistant	Resistant
GMAL 4473.b	Russia	RB	1998	44.43	40.15	400	3 km southwest of Shuntuk	Resistant	Susceptible	Susceptible
GMAL 4475.b	Russia	RB	1998	44.43	40.15	400	3 km southwest of Shuntuk	Susceptible	Resistant	Susceptible
GMAL 4478.d	Russia	RB	1998	44.43	40.15	400	3 km southwest of Shuntuk	Resistant	Susceptible	Susceptible
GMAL 4479.h	Russia	RB	1998	44.43	40.15	400	3 km southwest of Shuntuk	Susceptible	Resistant	Resistant
GMAL 4484.s	Russia	RB	1998	44.41	40.12	500	7 km southwest of Shuntuk	Resistant	Susceptible	Susceptible
GMAL 4488.f	Russia	RB	1998	44.45	40.21	300	3 km east of Shuntuk along the Bileyi River	Resistant	Resistant	Susceptible
GMAL 4493.s	Russia	RB	1998	44.23	40.18	700	25 km south of Shuntuk	Resistant	N/A ³	Susceptible
GMAL 4510.a	Turkey	TA	1999	41.293	41.51	750	Artvin	Resistant	Susceptible	Susceptible
GMAL 4513.m	Turkey	TA	1999	40.219	42.381	1470	Artvin	Resistant	Susceptible	Resistant
GMAL 4539.af	Turkey	TC	1999	40.548	36.628	1030	Avlunar, Tokat	Susceptible	Resistant	Susceptible
GMAL 4545.f	Turkey	TC	1999	40.934	35.385	1320	Merzifon, Amasya	Susceptible	Resistant	N/A
GMAL 4556.j	Turkey	TD	1999	41.659	33.586	1120	Agli, Kastamonu	Resistant	Resistant	Susceptible
GMAL 4556.p	Turkey	TD	1999	41.659	33.586	1120	Agli, Kastamonu	Resistant	Resistant	Resistant
GMAL 4566.k	Turkey	TD	1999	41.751	33.693	1080	Camil Koyu, Kastamonu	Susceptible	N/A	Susceptible
GMAL 4568.f	Turkey	TD	1999	41.79	33.664	1040	Catak, Kastamonu	Susceptible	Susceptible	Resistant
GMAL 4597.a	Armenia	AB	2001	40.367	44.267	1930		Susceptible	Resistant	N/A
GMAL 4597.i	Armenia	AB	2001	40.367	44.267	1930		Resistant	Susceptible	N/A
GMAL 4598.a	Armenia	AB	2001	40.617	44.983	1924		Resistant	Resistant	Resistant
GMAL 4601.m	Armenia	AA	2002	40.752	44.4712	1794	Vanadzor Valley, Lori Marz province	Susceptible	Resistant	N/A
GMAL 4602.n	Armenia	AA	2002	40.7797	44.4788	1526	Vanadzor Valley, Lori Marz province	Susceptible	Resistant	N/A
GMAL 4602.s	Armenia	AA	2002	40.7797	44.4788	1526	Vanadzor Valley, Lori Marz province	Susceptible	Susceptible	Resistant
GMAL 4681.k	Georgia	GB	2004	42.442	44.483	1700	Mtuleti province	Resistant	Resistant	N/A
GMAL 4682.i	Georgia	GE	2004	41.308	43.308	1587	Satere, Kartli province	Resistant	Resistant	N/A
GMAL 4685.m	Georgia	GD	2004	41.983	43.221	832	Vani, Imereti province	Resistant	Susceptible	N/A

²Location codes per Figure 1.³N/A = not available.

Fig. 2. Relationship between allelic frequency in the proposed *Malus orientalis* core collection of 27 individuals and the entire 776 National Plant Germplasm System-Plant Genetic Resources Unit (NPGS-PGRU) *M. orientalis* collection of individuals in the field.

substantially smaller than the core produced using the original MSTRAT algorithm (C. Richards, personal communication).

We have selected 27 *M. orientalis* individuals to represent the diversity of 776 *M. orientalis* individuals originating from Russia, Turkey, Armenia, and Georgia. The individuals were selected based on genotypic data because comprehensive phenotypic data were not available. We have shown that the allelic frequencies in the core set are representative of that of the *M. orientalis* trees in the PGRU collection. Narrowing the large population down to a smaller representative sample makes the *M. orientalis* collection more accessible to the breeding and research communities. Despite limited descriptive information, a conservation genetic approach that uses estimates of differentiation and a maximization strategy can produce core sets with a minimum amount of redundancy. Development of core sets can be justified when their use can be seen as an efficient entry point in the larger diversity of the collection.

Phenotypic data can be more easily collected from the core set of individuals and that data will be made publicly available on the GRIN database (USDA, 2009). The core set of individuals will also be repropagated and added to the main *Malus* field collection in Geneva, NY. Additional individuals that are not part of this core may also be maintained after further evaluation. Dormant buds from these individuals will be cryopreserved for long-term preservation (Forsline et al., 1998; Towill et al., 2004).

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