

# Application of Calcium to Soil and Cultivar Affect Elemental Concentration of Watermelon Leaf and Rind Tissue

W. Dennis Scott<sup>1</sup>, B. Dean McCraw<sup>2</sup>, James E. Motes<sup>2</sup>, and Michael W. Smith<sup>2</sup>

Department of Horticulture and Landscape Architecture, Oklahoma State University, Stillwater, OK 74078-0481

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**Abstract.** Field experiments were conducted to quantify the effect of Ca supplied as gypsum in factorial combination with watermelon [*Citrullus lanatus* (Thumb) Matsum and Nakai] cultivars Charleston Gray, Crimson Sweet, and Tri-X Seedless on yield and the elemental concentration of leaf and rind tissue. Also, the effect that ontogenetic changes and sectional differences had on the elemental concentration in rind tissue was investigated. The experiments were conducted at two locations in Oklahoma. Yield was not affected by Ca; however, mean melon weight was reduced at 1120 kg Ca/ha. Leaf Ca concentration increased linearly in response to Ca rate. 'Tri-X Seedless' had lower leaf Ca and higher K concentrations than did 'Charleston Gray' or 'Crimson Sweet'. Fruit ontogeny (days from anthesis) and melon section (blossom or stem-end) interacted to affect elemental concentrations in the rind tissue. There was also a significant genotypic effect on elemental concentration in rind tissue. Increasing rates of Ca applied to soil reduced the incidence of blossom-end rot (BER) in 'Charleston Gray' melons. Calcium treatment did not affect flesh redness or soluble solids concentration (SSC) of watermelon.

Adequate Ca nutrition is essential for normal plant growth and development (Kirkby and Pilbean, 1984). Soil water status, root vigor, relative humidity, wind, high substrate soluble salt concentration, and cultivar selection can affect Ca nutrition (Wiersum, 1979). Several Ca deficiency disorders have been described, including BER of pepper (*Capsicum annuum* L.), tomato (*Lycopersicon esculentum* Mill.), and watermelon (Shear, 1975). These disorders are related more to inefficient partitioning of Ca within the plant than to limited uptake (Bangerth, 1979). Low transpiring organs, such as fruits and enclosed tissue, accumulate less Ca than leaves from the same plant (Bangerth 1976).

In field trials, watermelon cultivars with cylindrical or subcylindrical fruit generally are highly susceptible to BER, while spherical-fruited cultivars are completely or highly resistant (Cirulli, 1974; Cirulli and Ciccarese, 1981).

Studies of the effects of lime on watermelon yield have been conducted with variable results (Bradley and Fleming, 1959; Jones et al., 1975; Lacascio and Lundy, 1962). Increasing rates of Ca, supplied as gypsum (CaSO<sub>4</sub>), were shown to reduce watermelon yield (Sundstrom and Carter, 1983). Watermelons grown in sand culture provided progressively more Ca, supplied as CaCl<sub>2</sub>, and showed increased concentrations of Ca in leaf and fruit tissue (Walters and Nettles, 1961). Data were not available for Ca accumulation in watermelon fruit grown in field plots or respondent Ca status in developing watermelon rind tissue.

Objectives of this study were to 1) determine if the rate of Ca applied to soil affects the accumulation of Ca in leaf and rind tissue,

2) determine the influence of growth stage on watermelon fruit Ca concentration, 3) evaluate phenotypic response of selected watermelon cultivars to increasing soil Ca content, and 4) determine if Ca application rate affects the incidence of BER.

## Materials and Methods

The field experiments were conducted at the Vegetable Research Station, Bixby, Okla., on a Sevim fine sandy loam [coarse-silty, mixed (calcareous), thermic Typic Udifluvents] and at the Research Nursery and Teaching Arboretum, Stillwater, Okla., on a Norge loam [fine-silty, mixed, thermic Udic Paleustolls] during the growing seasons of 1989 and 1990. Nitrogen was preplant-incorporated to supply 34 kg·ha<sup>-1</sup> at both locations, and plants were sidedressed to supply 34 kg N/ha at 4 weeks after transplanting. The top 20 cm of soil had a water pH ranging from 6.1 to 6.4 at Bixby and 5.5 to 5.9 at Stillwater. Soil tests showed that native levels of P and K were adequate. Black polyethylene mulch (1.2 m wide × 0.08 mm thick) and trickle irrigation hose (Bi-wall, Hardie Irrigation, Luguna Niguel, Calif.) with 0.38 mm diameter holes 30 cm apart were mechanically laid in rows on 5 m centers.

Three-week-old transplants grown in 100 cm<sup>3</sup> peat pots containing commercial peat-lite mix were in the three leaf stage when planted five per plot, 1.2 m apart. Transplanting was accomplished on 6 and 9 May 1989 and 19 and 23 May 1990 at Bixby and Stillwater, respectively. Soil water potential was maintained between 20 to 30 kPa with the aid of tensiometers installed 30 cm deep (Bhella, 1985).

2,6-dinitro-N,N-dipropyl-4-(trifluoromethyl) benzenamine (trifluralin) at 840 g·ha<sup>-1</sup> was incorporated between rows at the time of transplanting. Accepted commercial foliar insecticides were used including methyl-N-[(methylamino) carbonyl] oxylethanimidothioate (methomyl) and (S)-cyano(3-phenoxyphenyl)-methyl-(S)-4-chloroalpha-(1-methylethyl) benzenoacetate (fenvalerate).

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<sup>2</sup>Professor.

Abbreviations: BER, blossom-end rot; SSC, soluble solids concentration.

Table 1. Influence of soil applied gypsum (CaSO<sub>4</sub>) on the yield of watermelon.<sup>z</sup>

Ca rate (kg·ha <sup>-1</sup> )	Bixby	Stillwater
	Melon wt (kg)	Melon wt (kg)
0	9.06	10.17
280	9.18	10.00
560	9.56	10.22
1120	9.22	10.14
Linear	NS	NS
Quadratic	*	NS

<sup>z</sup>Means of 2 years, three cultivars.

<sup>NS</sup>, \*Nonsignificant or significant at  $P = 0.05$ .

Gypsum treatments in factorial combination with three cultivars were incorporated into a 1.5 m wide band and 18 cm deep at 0, 280, 560, and 1120 kg·ha<sup>-1</sup> Ca 1 week before transplanting. The cultivars, chosen to provide a range in susceptibility to BER, were 'Charleston Gray' (highly susceptible), 'Crimson Sweet' (intermediate), and 'Tri-X Seedless' (resistant). The experimental design was a split-split-split-plot, with three replications in 1989 and four in 1990. Gypsum was the main-plot, cultivar the subplot, the stage of development the sub-sub-plot, and the watermelon rind section the sub-sub-sub-plot.

Flowers were tagged at anthesis in both years. One fruit per plot was then sampled for elemental analyses at 14 and 21 days after anthesis and at maturity in 1989 and at 10 and 20 days after anthesis and at maturity in 1990. Fruit maturity among and between cultivars varied from 30 to 35 days after anthesis. Fruit samples (300 g fresh weight) were taken from the blossom and stem end of fourteen 21 day old and mature fruit. Leaf samples consisting of the first fully expanded leaf (lamina and petiole) were taken at anthesis in both years.

Fruit and leaf samples were dried at 80C for 7 days, ground to pass a mesh screen with 0.16 cm<sup>2</sup> openings, and stored in air-tight jars. Before analysis, samples were redried at 80C for 24 h. Total Ca was determined using the dry ash method (Isaac and Johnson, 1975), and extractable Ca was determined using a 2% acetic acid extraction (Gallaher and Jones, 1976). Standard methods were used for analysis of K, Mg, Zn, Fe, and Mn by atomic absorption spectroscopy (Perkin-Elmer, Model 303, Norwalk Conn.).

Watermelon fruits of marketable maturity were harvested four times at 1-week intervals beginning 31 July in 1989 and 6 Aug. in 1990. The number of fruits, their respective weights, and number of BER-affected fruits were determined. A single mature watermelon from each plot was sampled for SSC and redness. Color was determined by scanning the placental tissue with a Minolta CR-200 chroma meter (Minolta, Ramsey, N.J.). Values of Hunter's L, a, and b were taken. Only the 'a' value data (redness) are presented

(Francis, 1980). Homogenate from test watermelons was filtered through Whatman no. 6 filter paper, and SSC was measured on a Fisher Scientific Co. Refractometer, Model no. ABBE-3L (Dallas) equipped with circulating water temperature control set at 20c.

## Results

There were no interactions between Ca or cultivar with years; therefore, data are pooled for years. Calcium had no significant effect on watermelon yield (31 to 37 Mt·ha<sup>-1</sup> at Bixby, 38 to 41 Mta·ha<sup>-1</sup> at Stillwater). This result contrasts with those of previous reports (Sundstrom and Carter, 1983; Walters and Nettles, 1961). In treatments where Ca was supplied at 1120 kg·ha<sup>-1</sup>, mean melon

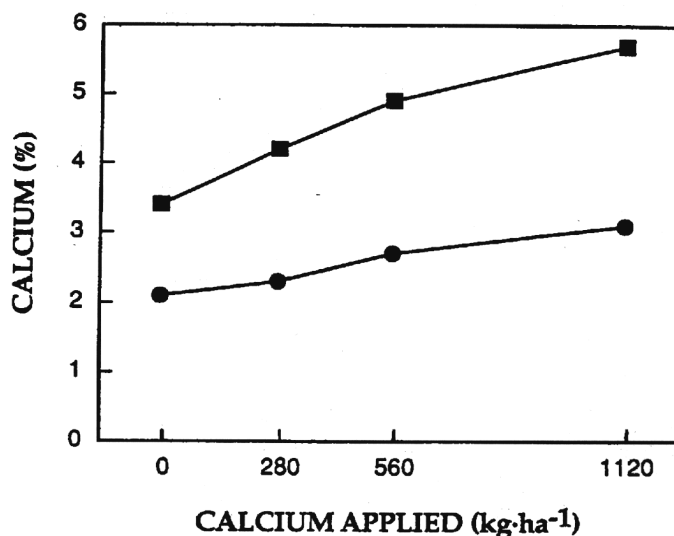


Fig. 1. Effect of soil applied Ca (CaSO<sub>4</sub>) on the concentration of total and extractable Ca in watermelon leaf tissue. Total Ca (■) and extractable Ca (●) values are means of two locations, three cultivars, and four replications (1990).

weight at Bixby but not at Stillwater decreased when compared to melons grown at the 560 kg·ha<sup>-1</sup> and lower rates (Table 1). All three cultivars yielded similarly (data not shown); however, mean melon weights, as yield, were higher in melons cultured at Stillwater than at Bixby. Soil pH values within Ca treatments were not influenced by gypsum rate.

Concentrations of total and acetic acid extractable Ca in watermelon leaf tissue were positively related to soil incorporated Ca (Fig. 1). When no Ca was applied, 61% of the total Ca fraction (2.11 µg·g<sup>-1</sup>) was extractable with 2% acetic acid solution. The extractable Ca fraction remained relatively constant at all Ca rates, ranging from 55% to 61% of the total Ca in the leaf tissue. The

Table 2. Effect of cultivar on watermelon leaf elemental concentration.<sup>z</sup>

Cultivar	Elemental concn						
	Dry wt (%)			Dry wt (µg·g <sup>-1</sup> )			
	Total Ca	Ext. Ca <sup>y</sup>	K	Mg	Zn	Fe	Mn
Charleston Gray	4.69 a	2.72 a	1.68 b	0.59 b	30 ab	111 b	138 b
Crimson Sweet	5.01 a	2.91 a	0.62 b	0.70a	34 a	126 a	182 a
Tri-X-Seedless	4.13 b	2.40 b	2.01 a	0.51 b	29 b	113 b	122 b

<sup>z</sup>Means of 2 years, two locations, and four Ca treatments. Mean separation in columns by Duncan's multiple range test,  $P = 0.05$ .

acetic acid extraction results in a concentration representative of all plant Ca, except that crystallized as Ca oxalate (Gallaher and Jones, 1976).

Genotype significantly influenced the elemental concentration of watermelon leaf tissue (Table 2). 'Charleston Gray' and 'Crimson Sweet' had similar concentrations of total and extractable Ca,

Table 3. Influence of Ca rate and rind section on the concentration of total and extractable Ca in watermelon rind tissue.

Ca rate (kg·ha <sup>-1</sup> )	Rind section <sup>y</sup>	Ca concn (% dry wt)							
		Bixby				Stillwater			
		1989		1990		1989		1990	
		Total Ca	Ext. Ca <sup>x</sup>	Total Ca	Ext. Ca	Total Ca	Ext. Ca	Total Ca	Ext. Ca
0	BE	0.85	0.35	0.52	0.30	0.78	0.35	0.51	0.28
	MT	0.98	0.42	0.48	0.30	0.60	0.25	0.44	0.31
	MB	1.08	0.47	0.61	0.25	0.74	0.27	0.51	0.23
	SE	1.01	0.37	0.61	0.29	0.82	0.42	0.62	0.27
280	BE	0.76	0.31	0.51	0.29	0.81	0.39	0.54	0.28
	MT	0.90	0.35	0.47	0.26	0.67	0.28	0.52	0.41
	MB	1.05	0.45	0.58	0.22	0.79	0.29	0.69	0.27
	SE	0.92	0.34	0.65	0.30	0.98	0.52	0.65	0.29
560	BE	0.70	0.27	0.55	0.30	0.85	0.42	0.55	0.32
	MT	1.01	0.34	0.43	0.29	0.66	0.27	0.47	0.37
	MB	0.93	0.44	0.60	0.21	0.74	0.28	0.57	0.24
	SE	0.85	0.26	0.63	0.32	0.91	0.59	0.64	0.31
1120	BE	0.84	0.34	0.53	0.30	0.80	0.39	0.52	0.29
	MT	0.97	0.43	0.47	0.30	0.81	0.35	0.49	0.35
	MB	1.30	0.63	0.62	0.23	0.84	0.30	0.62	0.23
	SE	1.00	0.36	0.67	0.32	0.83	0.57	0.63	0.28
LSD 0.05									
	Section within Ca rate	0.12	0.1	0.11	0.04	0.06	0.06	0.09	0.03
	Ca rate within section								
	Linear	NS	NS	NS	NS	NS	NS	NS	NS
	Quadratic	NS	*	NS	NS	*	*	*	*

<sup>y</sup>Values mean of three cultivars and four Ca rates.

<sup>y</sup>BE = blossom-end, MT = middle top, MB = middle bottom, SE = stem-end.

<sup>NS</sup>, \*Nonsignificant or significant at *P* = 0.05.

<sup>x</sup>Acetic acid extractable Ca.

Table 4. Influence of sampling time and section on the elemental concentration of watermelon rind (1989).

Sample time	Rind section <sup>y</sup>	Elemental concn													
		Bixby							Stillwater						
		Dry wt (%)		Dry wt (µg·g <sup>-1</sup> )					Dry wt (%)		Dry wt (µg·g <sup>-1</sup> )				
		Total Ca	Ext. Ca <sup>x</sup>	K	Mg	Zn	Fe	Mn	Total Ca	Ext. Ca	K	Mg	Zn	Fe	Mn
14 days <sup>w</sup>	BE	0.68	0.38	4.33	0.32	30	52	13	0.75	0.37	4.76	0.49	48	55	11
	SE	0.88	0.34	5.98	0.43	49	66	16	0.97	0.55	5.60	0.42	39	65	30
21 days	BE	0.87	0.27	6.47	0.38	37	63	20	0.75	0.35	6.42	0.42	42	43	12
	SE	0.90	0.28	6.79	0.38	42	58	26	0.93	0.42	5.70	0.40	43	53	28
Maturity	BE	0.81	0.27	6.85	0.36	34	58	14	0.78	0.27	6.72	0.39	51	38	10
	SE	1.05	0.38	9.34	0.44	47	67	25	0.94	0.45	7.09	0.43	46	46	71
LSD 0.05															
	Section within time	0.11	0.1	1.08	0.06	23	13	9	0.11	0.07	0.95	0.07	10	17	11
	Time within section	0.12	0.09	1.19	NS	NS	NS	NS	NS	0.08	0.96	NS	NS	NS	NS

<sup>y</sup>Values mean of three cultivars and four Ca rates.

<sup>y</sup>BE = blossom-end, SE = stem-end.

<sup>x</sup>Acetic acid extractable Ca.

<sup>w</sup>Days from anthesis.

but 'Tri-X Seedless' had significantly lower concentrations for both Ca fractions. This trend was reversed for K concentration, with 'Tri-X Seedless' containing more K than either of the other cultivars. These data support previous reports where K and Ca uptake were found to be inverse (Elmstrom et al., 1973). Concentrations of Mg, Zn, Fe, and Mn also varied among cultivars, with 'Crimson Sweet' containing a higher concentration of Mg, Fe, and Mn than the other two cultivars.

In contrast to leaf tissue, rind elemental Ca concentration was not affected by Ca treatment. Total Ca concentrations in fully expanded leaf tissue ranged from 100% to 500% higher than those of fruit rind tissue (Tables 2 and 3). The Ca concentration in watermelon fruit rind tissue was highly variable in relation to Ca rate. However, there was a significant Ca rate and rind section interaction on the accumulation of total and acetic acid extractable Ca in rind tissue. Calcium concentrations in watermelon fruit harvested at Stillwater in 1989 and 1990 were curvilinearly related to increasing Ca rate. Total and extractable Ca concentrations in rind tissue from either the blossom or stem-end peaked in response to the 560 kg-ha<sup>-1</sup> treatment, with a distinctive drop at the 1120

kg-ha<sup>-1</sup> Ca rate (Table 3).

The total Ca concentration in watermelon rind tissue harvested at Bixby increased during the development of the fruit (Tables 4 and 5). However, imported Ca was partitioned unequally within the melon, with the stem-end consistently maintaining a higher concentration of total Ca than the blossom-end. The acetic acid extractable Ca in the rind tissue was more variable than the total Ca fraction, however, at maturity, the stem-end had a significantly higher concentration of extractable Ca when compared to the blossom-end (Table 4).

At Stillwater in 1989, in contrast, the total Ca concentration in the blossom-end of the fruit continued to increase to maturity, while the levels of extractable Ca declined steadily during the ontogeny of the watermelon fruit. For both locations, K concentration increased over time in the stem-end of the fruit and increased in the blossom-end up to 21 days, then remained nearly unchanged through maturity (Table 4). Concentrations of Mg, Zn, Fe, and Mn differed based on the section being sampled rather than the stage of fruit development.

During both years, genotypic characteristics had a significant

Table 5. Influence of sampling time and section on the elemental concentration of watermelon rind (1990).<sup>z</sup>

		Elemental concn														
		Bixby							Stillwater							
Sample time	Rind section <sup>y</sup>	Dry wt (%)		Dry wt (µg·g <sup>-1</sup> )					Dry wt (%)		Dry wt (µg·g <sup>-1</sup> )					
		Total Ca	Ext. Ca <sup>x</sup>	K	Mg	Zn	Fe	Mn	Total Ca	Ext. Ca	K	Mg	Zn	Fe	Mn	
		10 days <sup>x</sup>	BE	0.52	0.29	3.62	0.21	25	52	7	0.48	0.26	4.24	0.21	21	61
	SE	0.54	0.26	3.49	0.20	30	48	15	0.49	0.27	4.80	0.21	29	40	27	
20 days	BE	0.54	0.32	4.48	0.21	26	39	25	0.56	0.28	5.29	0.25	24	58	49	
	SE	0.67	0.34	4.78	0.21	28	41	19	0.61	0.32	5.28	0.21	26	36	34	
Maturity	BE	0.53	0.28	4.64	0.23	37	40	30	0.52	0.30	5.94	0.25	34	55	44	
	SE	0.70	0.35	5.32	0.24	34	46	25	0.86	0.31	5.81	0.25	30	55	31	
LSD 0.05																
Section within time		0.05	0.03	0.25	0.01	4	6	4	0.04	0.03	0.20	0.01	3	4	4	
Time within section		0.04	NS	0.31	0.01	NS	7	4	0.06	0.03	0.36	NS	NS	4	5	

<sup>x</sup>Values mean of three cultivars and four Ca rates.

<sup>y</sup>BE = blossom-end, SE = stem-end.

<sup>x</sup>Acetic acid extractable Ca.

<sup>x</sup>Days from anthesis.

Table 6. Influence of cultivar on the elemental concentration of watermelon rind.<sup>z</sup>

		Elemental concn														
		Bixby							Stillwater							
Cultivar		Dry wt (%)		Dry wt (µg·g <sup>-1</sup> )					Dry wt (%)		Dry wt (µg·g <sup>-1</sup> )					
		Total Ca	Ext. Ca <sup>y</sup>	K	Mg	Zn	Fe	Mn	Total Ca	Ext. Ca	K	Mg	Zn	Fe	Mn	
		1989														
Charleston Gray		0.95 a	0.40 a	6.84 c	0.43 a	60 a	65 a	26 a	0.91 a	0.44 a	5.81 a	0.43 a	51 a	57 a	26 a	
Crimson Sweet		0.93 a	0.38 a	7.31 b	0.41 a	63 a	63 a	22 a	0.83 a	0.39 a	6.24 a	0.41 a	50 a	57 a	22 a	
Tri-X Seedless		0.83 b	0.28 b	8.00 a	0.40 a	67 a	62 a	25 a	0.71 b	0.29 b	6.37 a	0.40 a	42 a	50 a	17 a	
1990																
Charleston Gray		0.62 a	0.32 a	4.23 b	0.23 a	31 b	45 a	22 a	0.61 a	0.33 a	4.96 c	0.25 a	30 a	54 a	37 a	
Crimson Sweet		0.61 a	0.31 a	4.89 a	0.21 a	36 a	44 a	21 a	0.58 a	0.30 a	5.49 a	0.23 a	31 a	51 a	36 a	
Tri-X		0.49 b	0.24 b	4.67 a	0.20 a	29 b	41 a	20 a	0.52 b	0.26 b	5.20 b	0.22 a	23 b	46 b	33 a	

<sup>z</sup>Means of four Ca rates and two sections. Mean separation within column and location by Duncan's multiple range test, *P* = 0.05.

<sup>y</sup>Acetic acid extractable Ca.

Table 7. Incidence of BER in 'Charleston Gray' melons as affected by Ca applied to soil (gypsum).

Ca applied (kg·ha <sup>-1</sup> )	No. of BER affected fruits			
	1989		1990	
	Bixby	Stillwater	Bixby	Stillwater
0	9 <sup>z</sup>	8	13	12
280	6	11	11	11
560	5	4	7	4
1120	2	2	4	3
Linear	*	NS	*	*
Quadratic	NS	*	NS	NS

<sup>NS</sup>, \*Nonsignificant or significant at  $P = 0.05$ .

impact on the elemental concentration in watermelon rind tissue (Table 6). For both years and locations 'Tri-X Seedless' had total Ca concentrations lower than 'Charleston Gray' or 'Crimson sweet'. This was also true for the extractable Ca fraction. 'Tri-X Seedless' rind tissue also had a lower percentage of the total Ca in the acetic acid extractable fraction than the other two cultivars. In the 1989 growing season, 37% of the total Ca fraction in 'Tri-X Seedless' was extractable Ca. For 'Charleston Gray' and 'Crimson Sweet', the extractable Ca fraction constituted 45% of the total Ca. Concentrations for most elements were higher in 1989 than they were in 1990 (Table 6).

Tissue K concentration differed among the three cultivars, except at Stillwater in 1989. 'Tri-X Seedless' and 'Crimson Sweet' had higher K concentrations than 'Charleston Gray' (Table 6). Magnesium, Zn, Fe, and Mn concentrations were usually not significantly different among cultivars.

Mean red flesh Hunter 'a' color values ranged from 19.8 to 24.9 and were not significantly affected by Ca fertilization or cultivar. Fruit heart tissue SSC ranged from 9.1% to 11.1% and were independent of treatment.

BER is a physiological disorder of watermelon commonly attributed to inadequate Ca concentrations in the affected tissue (Foroughi and Kloke, 1974, Walters and Nettles, 1961). Increasing rates of soil-incorporated Ca reduced the incidence of BER in 'Charleston Gray' watermelons (Table 7). Only five 'Crimson Sweet' fruit were affected with BER in each of the two years. The 'Tri-X Seedless' watermelon was completely resistant to the disorder.

#### Discussion

The positive relation between elemental concentrations for total and extractable Ca of leaves and soil-incorporated Ca rates (Fig. 1) indicates that gypsum is an adequate source of Ca for field mineral nutrition of watermelon, at least in cases where the pH adjusting benefit of lime is not required. Root systems of melon plants grown with plastic mulch tend to be confined near the surface and within the polyethylene mulch bed (Bhella, 1985).

Recognizable foliar symptoms of Ca deficiency are seldom observed in field-grown fruit or vegetable crops. Calcium becomes limiting due to an inefficient distribution of Ca by the transpirative stream to growing tissue rather than low Ca uptake (Kirkby and Pilbeam, 1984). As a result of different Ca allocation and remobilization, Ca concentrations in watermelon leaves and fruits were not directly related. These results agree with conclusions drawn from studies conducted on Ca nutrition of cucumber (*Cucumis sativus* L.) (Engelkes et al., 1990).

Elemental concentrations in the stem-end of rind tissue increased during fruit development (Tables 4 and 5). These data indicate that

imported Ca is first deposited at the stem-end and that transport to the blossom-end may be impeded, at least late in development. This gradient in Ca concentration from the stem to blossom-end also has been observed in cucumber and hypothesized as Ca depletion from the xylem solution during the transport (Frost and Kretschman, 1989). Tissue from the blossom-end consistently had a lower total Ca concentration (0.68% to 0.87%, 1989; 0.48% to 0.56%, 1990) than tissue in the stem-end (0.88% to 1.05%, 1989; 0.49% to 0.86%, 1990).

Transport of Ca into the stem-end of watermelon fruit continues to maturity, while accumulation in the blossom-end slows significantly 7 to 10 days before harvest (Tables 4 and 5). This trend may be due to reduced movement of Ca into the blossom-end and/or dilution of the existing concentrations of the elements as the fruit grew. Whatever the cause, all three cultivars studied exhibited the same trend.

Cultivar differences were evident for the accumulation of Ca in watermelon rind tissue (Table 6). Differences in resistance of tomato cultivars to BER have been shown to be due to differences in efficiency of Ca uptake and accumulation in the fruit or to differences in the Ca concentration in the fruit (Greenleaf and Adams, 1969).

This study did not elucidate the basis for the variation among cultivars in regard to Ca nutrition. However, 'Tri-X Seedless' fruit consistently had lower concentrations of total and extractable Ca in leaf and rind tissue than the other two test cultivars. Despite maintaining a lower Ca concentration, 'Tri-X Seedless' demonstrated a complete resistance to BER. However, 'Charleston Gray' with a relatively high concentration of total Ca in the rind had a high incidence of BER. Further investigation is needed to adequately describe the histological and physiological differences among these cultivars. These findings are in agreement with work conducted previously (Sundstrom and Carter, 1983).

Cultivar selection and environmental influences all combine to influence the elemental status of plant tissues. This study clearly shows that the cultivar chosen is nearly as important as external pressures applied by the environment, at least relative to soil Ca levels (pH not being limiting).

Elemental concentrations in the leaf and rind tissue were significantly influenced by the environment within each year. Accumulation of nearly all elements was higher during the 1989 than the 1990 growing season (Table 6). In 1989 the watermelons were infected by a severe outbreak of the foliar disease anthracnose [*Colletotrichum orbiculare* (Berk. & Mont.) Arx]. The disease reduced biomass production, which in turn may have concentrated the elements resulting in increased elemental concentrations. Precipitation was significantly higher in 1989 than in 1990. Although

all plots were maintained at adequate soil moisture levels and monitored using tensiometers, in 1989 the soil profile adjacent to the plots was wetter, and this condition may have led to increased uptake of nutrients.

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