Increased Fertilizer Levels Do Not Prevent Abscisic Acid–Induced Chlorosis in Pansy

Jong-Goo Kang¹, Rhuanito Soranz Ferrarezi², Sue K. Dove³, Geoffrey M. Weaver³, and Marc W. van Iersel³,⁴

SUMMARY. Abscisic acid (ABA) is a plant hormone involved in regulating stomatal responses to environmental stress. By inducing stomatal closure, applications of exogenous ABA can reduce plant water use and delay the onset of drought stress when plants are not watered. However, ABA can also cause unwanted side effects, including chlorosis. Pansy (Viola × wittrockiana) has been shown to be particularly susceptible to ABA-induced chlorosis. The objective of this study was to determine if fertilization rate affects the severity of ABA-induced chlorosis in this species.

‘Delta Premium Pure Yellow’ pansy seedlings were fertilized with controlled-release fertilizer incorporated at rates from 0 to 8 g L⁻¹ of substrate. When plants had reached a salable size, half the plants were sprayed with a solution containing 1 g L⁻¹ ABA, whereas the other plants were sprayed with water. Leaf chlorophyll content was monitored for 2 weeks following ABA application. Leaf chlorophyll content increased greatly as fertilizer rate increased from 0 to 2 g L⁻¹, with little increase in leaf chlorophyll at even higher fertilizer rates. ABA induced chlorosis, irrespective of the fertilizer rate. Plant dry weight was lowest when no controlled-release fertilizer was incorporated, but similar in all fertilized treatments. ABA treatment reduced shoot dry weight by ≈24%, regardless of fertilizer rate. This may be due to ABA-induced stomatal closure, which limits carbon dioxide (CO₂) diffusion into the leaves. We conclude that ABA sprays induce chlorosis, regardless of which fertilizer rate is used. However, because leaf chlorophyll concentration increases with increasing fertilizer rates, higher fertilizer rates can mask ABA-induced chlorosis.

Additional index words. ABA, chlorophyll, controlled-release fertilizer, stomatal closure, Viola × wittrockiana

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Materials and methods

Greenhouse environment. This study was performed in a glass-covered greenhouse in Athens, GA, from 21 Oct. to 20 Dec. 2013. Greenhouse environmental conditions were measured using a quantum sensor for photosynthetic photon flux (PPF) (SQ-110; Apogee Instruments, Logan, UT) and a temperature and relative humidity sensor (HMP50; Vaisala, Vantaa, Finland) connected to a datalogger (CR10; Campbell Scientific, Logan, UT). The vapor pressure deficit was calculated from the saturated and actual air vapor pressure, using the air temperature and relative humidity data. The daily light integral was calculated by integrating the PPF measurements for each day. The daily average values, from the time of ABA treatment (45 d after transplanting)
to the end of the study, were (mean ± SD) as follows: temperature 20.4 ± 0.9 °C, daily light integral 9.9 ± 5.1 mol·m⁻²·d⁻¹, and vapor pressure deficit 1.14 ± 0.45 kPa.

**PLANT MATERIAL AND FERTILIZER TREATMENTS.** ‘Delta Pure Premium Yellow’ pansy seedling plugs were transplanted into square, 4-inch pots filled with soilless substrate (Fafard 1P; Sun Gro Horticulture, Agawam, MA) on 21 Oct. 2016. Plants were grown in a greenhouse on ebb-and-flow benches (150 cm length × 90 cm width × 4 cm height; MidWest Gro-Master, St. Charles, IL) covered with commercial grade weed cloth (Weed FreePro Fabric; Du Pont, Wilmington, DE). A separate 70-L water tank and submersible pump (NK-2; Little Giant, Oklahoma City, OK) were used for each bench. Plants were watered once daily with potable water. A 19N–1.7P–5.4K controlled-release fertilizer (19–4–8 with micronutrients; Harrell’s, Lakeland, FL) was incorporated at seven different rates (0, 1, 2, 3, 4, 5, and 8 g·L⁻¹). The recommended incorporation rate for this fertilizer is 6.5 to 10 lb/yard³ (≈4 to 6 g·L⁻¹).

**ABA SPRAY SOLUTIONS.** Stock solution of s-ABA, the biologically active isomer of ABA [10% s-ABA (VBC-30101; Valent BioSciences Corporation, Long Grove, IL)] was diluted with a surfactant solution (Brij 98 Surfactant; Valent BioSciences Corporation). The final solution contained 1 g·L⁻¹ ABA and 0.1 mg·L⁻¹ surfactant. This concentration was chosen because previous studies indicated that this is an effective rate to induce chlorosis in pansy (Weaver and van Iersel, 2014). Half of the plants of each fertilization rate were sprayed with the ABA solution once all plants had reached a salable size, 45 d after transplanting. Control plants were sprayed with water with 0.1 mg·L⁻¹ surfactant.

**MEASUREMENTS.** To monitor the effect of fertilizer rate and plant available nutrients, pore water electrical conductivity (EC) was measured at four different times during the study, using a sensor that measures the temperature, dielectric, and bulk EC of the substrate (WET-2; Delta-T Devices, Burwell, Cambridge, UK). Those data are used to calculate the pore water EC (Hilhorst, 2000). Leaf chlorophyll concentration index (CCI) was measured on uppermost fully expanded leaves using a leaf chlorophyll meter (CCM-200; Apogee Instruments). The CCI is closely, but not linearly, related to leaf chlorophyll concentrations (Parry et al., 2014). All plants were measured at 0, 1, 3, 5, 7, 10, and 15 d after spray treatment. Above-ground parts of all plants were harvested 15 d after spray treatment. Samples were dried in a drying oven at 80 °C for 4 d and then weighed.

**STATISTICS.** Plants were arranged in a randomized complete-block design with three replications. Each experimental unit had eight plants. Statistical analysis of pore water EC and leaf CCI was performed using repeated measures analysis of variance (ANOVA) (SAS version 9.2; SAS Institute, Cary, NC). In the case of a significant ABA-treatment × fertilizer interaction (i.e., CCI), the slice option of SAS was used to test for differences between the ABA-treated and control plants at each fertilization rate (SAS Institute, 2010). Dry weights were analyzed using ANOVA (SAS version 9.2). Plant responses to different fertilizer rates were analyzed using nonlinear regression (SigmaPlot 11.0; Systat Software, San Jose, CA).

**RESULTS AND DISCUSSION.**

**PORE WATER EC.** Pore water EC of the substrate was similar on all four sampling dates and unaffected by ABA application. Pore water EC increased nonlinearly with increasing fertilizer rates, from 1 dS·m⁻¹ without any fertilizer to 3.2 dS·m⁻¹ with a fertilizer rate of 8 g·L⁻¹ [P < 0.0001 (Fig. 1)]. This increase in EC was expected since increased fertilizer rates result in higher nutrient ion concentrations in the solution, detected by the pore water EC readings. Similarly, Klock-Moore and Broschat (1999) saw an increase in leachate EC with increasing controlled-release fertilizer rates and little change in leachate EC over time.

**LEAF CHLOROPHYLL.** There was a significant interactive effect of ABA-treatment × fertilizer rate (P = 0.03) and a main effect of measurement day on the leaf CCI (P < 0.0001). Leaf CCI increased near the end of the study and was 6.6 higher on the last than on the first measurement day, regardless of ABA or fertilizer treatment. The application of ABA significantly decreased leaf CCI below the levels of the control plants in the 2, 4, 6, and 8 g·L⁻¹ fertilizer treatments. A similar, but nonsignificant trend was seen at the other fertilizer levels (Fig. 2). This is consistent with previous findings that exogenous ABA spray applications cause chlorosis in pansy (Waterland et al., 2010a, 2010b; Weaver and van Iersel, 2014). Leaf CCI increased asymptotically with fertilization rate in both the control and ABA-treated plants [P < 0.0001 (Fig. 2)]. Nitrogen and other nutrient deficiencies are well known to cause leaf chlorosis (Marschner, 2012), so the increase in CCI with increasing fertilizer levels is expected.

The extent of ABA-induced leaf chlorosis in this study was less than that has been previously observed with pansy (Waterland et al., 2010a, 2010b; Weaver and van Iersel, 2014). This study differs from previous research that reported more severe chlorosis at similar rates of ABA because controlled-release fertilizer was used, whereas in earlier experiments plants were fertigated daily (Waterland et al., 2010a, 2010b; Weaver and van Iersel, 2014). However, the use of controlled-release fertilizer resulted in a stable pore water EC over the course of the study, suggesting a steady nutrient supply for the plants. It thus seems unlikely that the use of controlled-release fertilizer reduced the severity of the chlorosis.

**SHOOT DRY WEIGHT.** Final shoot dry weight was affected by fertilizer rate and ABA application, but not by their interaction. At all fertilization rates, ABA spray treatment reduced...
Fig. 2. Leaf chlorophyll concentration index of ‘Delta Premium Pure Yellow’ pansy grown with different controlled-release fertilizer rates in a peat:perlite substrate. Plants were sprayed with a 1 g L\(^{-1}\) abscisic acid (ABA) solution or with water (no ABA). * indicates significant differences between control and ABA-treated plants within a specific fertilizer rate (\(P<0.05\), \(n=3\)). Data are averaged over seven measurement days; 1 g L\(^{-1}\) = 0.1335 oz/gal.

the shoot dry weights by \(\approx 24\%\) \((P < 0.001\) (Fig. 3)). Similarly, Agehara and Leskovar (2015) reported that ABA sprays reduced root and shoot dry weight of bell pepper (\textit{Capsicum annuum}) transplants. The ABA-induced reduction in dry matter accumulation is likely caused by restricted diffusion of CO\(_2\) into the leaf tissue due to ABA-induced stomatal closure (Buckley, 2005; Chaves et al., 2003; Pantin et al., 2012). Weaver and van Iersel (2014) previously showed that ABA spray applications reduce stomatal conductance and net photosynthesis of pansy in a rate-dependent manner for 2 weeks after application.

Within the ABA-treated and control groups, shoot dry weights were similar in all fertilized treatments (1.65 and 2.16 g/plant, respectively), but lower in the treatments without fertilizer (Fig. 3). This indicates that even the lowest fertilizer rate was sufficient to support maximal pansy shoot growth. Little information is available concerning controlled-release fertilization rates in subirrigated ornamentals, because water-soluble fertilizers are typically used in subirrigation systems. Klock-Moore and Broschat (1999) showed that shoot growth of ‘Ultra Red’ petunia (\textit{Petunia xhybrida}) and ‘Super Elfin Violet’ impatiens (\textit{Impatiens walleriana}) increased linearly as controlled-release fertilizer rate increased from 1.25 to 7.5 g L\(^{-1}\). Richards and Reed (2004) reported that dry weight of subirrigated ’Illusion’ new guinea impatiens (\textit{Impatiens hawkeri}) increased with increase in controlled-release fertilizer rates from 3.56 to 10.67 g L\(^{-1}\), but decreased at higher rates. Thus, our finding that shoot dry weight of pansy was similar with fertilizer rates from 1 to 8 g L\(^{-1}\) differs from previous findings. This may be related to differences among species, fertilizers, or environmental conditions in the various studies.

Conclusions

Spray applications of ABA lowered leaf CCI of pansy. Although higher fertilizer rates increased leaf CCI, they did not negate the effect of ABA sprays on CCI. Nonetheless, higher fertilizer rates could be used to mask the chlorosis induced by ABA applications. In addition to reducing chlorosis, ABA also reduced shoot dry weight by 24%, regardless of the fertilizer application. The effect of ABA on dry weight cannot be negated by using higher fertilizer rates, since rates of 1 to 8 g L\(^{-1}\) all resulted in similar shoot dry weights.

Literature cited


