

Light Intensity and Quality from Sole-source Light-emitting Diodes Impact Growth, Morphology, and Nutrient Content of *Brassica* Microgreens

Joshua R. Gerovac and Joshua K. Craver

Department of Horticulture and Landscape Architecture, Purdue University, West Lafayette, IN 47907

Jennifer K. Boldt

USDA-ARS, Greenhouse Production Research Group, 2801 W. Bancroft Street, Mail Stop 604, Toledo, OH 43606

Roberto G. Lopez¹

Department of Horticulture, Michigan State University, 1066 Bogue Street, Room A288, East Lansing, MI 48824

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Abstract. Multilayer vertical production systems using sole-source (SS) lighting can be used for the production of microgreens; however, traditional SS lighting methods can consume large amounts of electrical energy. Light-emitting diodes (LEDs) offer many advantages over conventional light sources, including high photoelectric conversion efficiencies, narrowband spectral light quality (LQ), low thermal output, and adjustable light intensities (LIs). The objective of this study was to quantify the effects of SS LEDs of different light qualities and intensities on growth, morphology, and nutrient content of *Brassica* microgreens. Purple kohlrabi (*Brassica oleracea* L. var. *gongylodes* L.), mizuna (*Brassica rapa* L. var. *japonica*), and mustard [*Brassica juncea* (L.) Czern. ‘Garnet Giant’] were grown in hydroponic tray systems placed on multilayer shelves in a walk-in growth chamber. A daily light integral (DLI) of 6, 12, or 18 mol·m⁻²·d⁻¹ was achieved from commercially available SS LED arrays with light ratios (%) of red:green:blue 74:18:8 (R₇₄:G₁₈:B₈), red:blue 87:13 (R₈₇:B₁₃), or red:far-red:blue 84:7:9 (R₈₄:FR₇:B₉) with a total photon flux (TPF) from 400 to 800 nm of 105, 210, or 315 μmol·m⁻²·s⁻¹ for 16 hours. Regardless of LQ, as the LI increased from 105 to 315 μmol·m⁻²·s⁻¹, hypocotyl length (HL) decreased and percent dry weight (DW) increased for kohlrabi, mizuna, and mustard microgreens. With increasing LI, leaf area (LA) of kohlrabi generally decreased and relative chlorophyll content (RCC) increased. In addition, nutrient content increased under low LIs regardless of LQ. The results from this study can help growers to select LIs and LQs from commercially available SS LEDs to achieve preferred growth characteristics of *Brassica* microgreens.

Microgreens and baby greens are a relatively new specialty crop appearing in many upscale markets and restaurants. Collectively, these crops consist of vegetables and herbs consumed at a young growth stage. The main difference between the two is that microgreens are harvested at the base of the

hypocotyl when the first set of true leaves start to emerge, while baby greens are harvested after the first set of true leaves has developed, generally ≥21 d after germination (Treadwell et al., 2010). Microgreens are mainly used by chefs and consumers to enhance the flavor, color, and texture of various foods (Treadwell et al., 2010). In addition, several species of microgreens contain high concentrations of health-promoting phytochemicals (Xiao et al., 2012). Commercial greenhouse growers have recently become interested in producing microgreens because of their high market value. Wholesale prices currently range from US\$60 to \$100 per kg for microgreens packaged in clamshell containers (Resh, 2013; Treadwell et al., 2010). In comparison, the wholesale price of greenhouse-grown boston lettuce (*Lactuca sativa* L.) packaged in plastic containers was about US\$12 to \$16 per kg (United States Department of Agriculture,

2016). Specifically, microgreens of the genus *Brassica* have become a popular choice due to the ease of germination, relatively short production time (7 to 21 d), and wide offering of intense flavors and colors (Xiao et al., 2012).

Several commercial growers are currently producing microgreens in greenhouses using soilless media in trays. Microgreens are also produced hydroponically using capillary mats placed in troughs, similar to those used in a nutrient film technique system. Another technique being used is a combination of hydroponics and SS lighting in multilayer vertical growing systems (Resh, 2013). Multilayer vertical growing systems using SS lighting were first developed and implemented commercially in Japan in the early 2000s (Goto, 2012). Although fluorescent lamps were initially used as the standard light source, growers have begun replacing them with LED arrays. Several operations worldwide have implemented this technology as LEDs have become more economically viable due to increased efficiency and decreased cost (Goto, 2012).

Multilayer vertical growing operations have substantial energy costs due to the amount of electricity required for SS lighting and temperature management (Goto, 2012). Light-emitting diodes offer many advantages over conventional light sources, including high photoelectric conversion efficiencies, narrowband spectral distribution, low thermal output, and adjustable LIs (Yeh and Chung, 2009). These advantages may become even more prevalent and defined as technology and research continue to improve.

Another potential advantage of using LEDs is the ability to select light qualities and intensities that have beneficial effects on plant growth and photomorphogenesis (Goto, 2012). The ability to impact growth of microgreens has been recently investigated using SS LEDs at different LIs. Samuolienė et al. (2013) found that increasing the photosynthetic photon flux (PPF) of SS LED lighting with a red:far-red:blue light ratio (%) of 91:1:8 (R₉₁:FR₁:B₈) led to significantly reduced hypocotyl elongation of kohlrabi (*B. oleracea* var. *gongylodes* ‘Delicacy Purple’), tatsoi (*B. rapa* var. *rosularis*), and mustard (*B. juncea* L. ‘Red Lion’), and increased percent DW of red pak choy (*B. rapa* var. *chinensis* ‘Rubi F₁’) and tatsoi microgreens. In regard to LQ, Li and Kubota (2009) reported that white light supplemented with far-red light significantly increased fresh weight (FW), DW, stem length, leaf length, and leaf width, compared with white light alone, of baby leaf lettuce ‘Red Cross’. An additional study of LQ was conducted by Kopsell and Sams (2013) on broccoli (*B. oleracea* var. *italica* Plenck) microgreens 13 d after sowing. Seeds were placed under either a red:blue light ratio (%) of 88:12 (R₈₈:B₁₂) at a continuous PPF of 350 μmol·m⁻²·s⁻¹ or 0:100 (R₀:B₁₀₀) at 41 μmol·m⁻²·s⁻¹. Specifically, they found that microgreens grown under the R₀:B₁₀₀ light ratio produced significantly higher levels of all essential nutrients compared with those grown under R₈₈:B₁₂ 19 d after sowing.

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¹Corresponding author. E-mail: rglopez@msu.edu.

Although previous reports have indicated that LI or LQ from SS LEDs had an effect on the growth of microgreens and baby greens, to our knowledge, little work has been published on the interaction between LI and LQ on the growth and nutrient content of *Brassica* microgreens. Therefore, the objective of this study was to quantify the effects of SS LEDs providing different LIs and LQs on the growth, morphology, and nutrient content of *Brassica* microgreens.

Materials and Methods

Plant material and culture. A hydroponic tray system for microgreen culture was created using polyethylene terephthalate fiber pads (50.8 cm × 24.7 cm × 0.89 cm; Sure to Grow, Beachwood, OH) placed in trays (52 cm × 26 cm × 6 cm) without drainage holes. Pads were initially hydrated with 350 mL of a calcium chloride (CaCl₂) solution in deionized water to provide 100 mg·L⁻¹ calcium (Ca) to ensure uniform germination. Nine tray systems were established for each species by sowing 25 g of purple kohlrabi (*B. oleracea* var. *gongyloides* L.), 15 g of mizuna (*B. rapa* L. var. *japonica*), or 15 g of mustard [*B. juncea* (L.) Czern. 'Garnet Giant'] (Johnny's Selected Seeds, Winslow, ME) seeds evenly onto each hydrated pad. For the first 5 d after sowing, 100 mL of CaCl₂ solution was added to each tray daily to further stimulate seedling growth. Once cotyledons were fully reflexed 5 d after sowing, ±300 mL of a 25% Hoagland's no. 1 nutrient solution (Hoagland and Aron, 1950) was added to each tray daily to provide (in mg·L⁻¹) 53 nitrogen (N), 8 phosphorus, 59 potassium (K), 50 Ca, 12 magnesium, 0.5 iron (Fe), 0.13 manganese, 0.01 zinc, 0.005 copper, 0.13 boron (B), and 0.002 molybdenum until harvest.

Growth chamber environment. Trays were placed on stainless steel shelves in three vertical layers in a walk-in growth chamber (C5 Control System; Environmental Growth Chambers, Chagrin Falls, OH) on 28 July, 18 Aug., or 11 Sept. 2014 to germinate in darkness under an average daily temperature, relative humidity (RH), and carbon dioxide (CO₂) concentration of 21 ± 0.1 °C, 80 ± 0.5%, and 500 ± 21 μmol·mol⁻¹, respectively. After a 3-d germination period, the air temperature set point was changed to 21 °C day/17 °C night (D/N; 16 h/8 h), the RH set point was changed to 55/65% D/N, and the CO₂ concentration was maintained at 500 μmol·mol⁻¹. Average D/N air temperatures, D/N RHs, and CO₂ concentrations were logged every 15 min by a data logger (DL1 Datalogger; Environmental Growth Chambers) with the mean ± SD of the combined experimental replications measuring 21.0 ± 0.1 D/17.0 ± 0.1 °C N, 65.5 ± 0.6 D/55.9 ± 1.0% N, and 504.5 ± 47.8 μmol·mol⁻¹, respectively.

SS LED lighting. Light-emitting diode arrays providing light ratios (%) of red:green:blue 74:18:8 (R₇₄:G₁₈:B₈), red:blue 87:13 (R₈₇:B₁₃), or red:far-red:blue 84:7:9 (R₈₄:FR₇:B₉) (Philips GreenPower LED production modules; Koninklijke Philips Electronics, N.V.,

Amsterdam, The Netherlands) were mounted to the underside of nine stainless steel shelves (123 cm long and 61 cm wide). Nonreflective blackout cloth was used to prevent light pollution between treatments. Average TPF, from 400 to 800 nm, of 105, 210, or 315 μmol·m⁻²·s⁻¹ was achieved by mounting 2, 4, or 6 arrays, spaced 20.3, 12.2, or 8.6 cm apart, respectively, ≈38 cm above the crop canopy. A 16-h (0600 to 2200 HR) photoperiod provided plants with a DLI of 6, 12, or 18 mol·m⁻²·d⁻¹, respectively. Light quality and TPF were measured at the beginning and confirmed at the end of each experimental replication by taking nine individual spectral scans per treatment using a spectroradiometer (PS-100; Apogee Instruments Inc., Logan, UT). The spectral LQ for each of the SS LED treatments is reported in Fig. 1. In addition, average TPF and DLI are reported in Table 1. Kohlrabi was placed under light treatments on 31 July and 21 Aug. 2014. Mizuna and mustard were placed under light treatments on 31 July and 14 Sept. 2014.

Growth and morphology measurements. Growth and morphology data were collected for kohlrabi, mizuna, and mustard, 10, 15, and 14 d after sowing, respectively. Ten seedlings of each species were randomly selected and measured to determine HL, LA, and total chlorophyll (*a* + *b*) content (i.e., RCC) for each SS LED treatment. Hypocotyl length was measured from the base of the hypocotyl to the shoot apical meristem using a digital caliper (DigiMax; Wiha, Schonach, Germany). Leaf area of cotyledons and fully expanded leaves was measured using a LA meter (LI-3100; LI-COR Inc., Lincoln, NE) by recording the average of three scans. Relative chlorophyll content of the cotyledons was measured using a SPAD meter (SPAD-502; Konica Minolta Sensing, Inc., Osaka, Japan). Ten samples, each comprised of 10 randomly selected seedlings per species, were used to determine FW and DW. After FW data were collected, each sample was dried in an oven at 70 °C for at least 4 d and DW data were recorded. Fresh weight and DW data were then used to report percent DW (DW/FW × 100) to reduce variability between the samples collected.

Nutrient analysis. A representative sample of microgreen tissue from each treatment and replication was oven-dried at 70 °C for at least 4 d, ground into a fine powder with a mortar and pestle, and then divided into two sample fractions. Foliar N was determined using a CHN analyzer (PerkinElmer Series II CHNS/O Analyzer; PerkinElmer Instruments, Shelton, CT). For all other elements, plant tissue was microwave digested (MARS; CEM Corp., Matthews, NC) and nutrient concentration was determined using inductively coupled plasma optical emission spectroscopy (ICP-OES; Thermo iCAP 6300; Thermo Electron Corp., Waltham, MA) as described by Frantz (2013).

Experimental design and statistical analysis. The experiment was laid out in a randomized block design in a factorial arrangement with LI (three levels) and LQ (three levels) as factors. The experiment was performed twice over

time for each species and data were pooled across replications. The effects of LI and LQ were compared by analysis of variance using SAS (SAS version 9.3; SAS Institute, Cary, NC) PROC MIXED, with an additional program (Arnold M. Saxton, University of Tennessee, Knoxville, TN) that provided pairwise comparisons between treatments using Tukey's honestly significant difference test at *P* ≤ 0.05.

Results and Discussion

Hypocotyl length. Hypocotyl length of kohlrabi, mizuna, and mustard decreased progressively as LI increased (Fig. 2A–C). For example, HL of kohlrabi grown under the light ratio of R₈₄:FR₇:B₉ decreased 32% as LI increased from 105 to 315 μmol·m⁻²·s⁻¹. The role of gibberellins (GA) in the regulation of hypocotyl elongation has been well established. Increased LIs can reduce levels of endogenous GA content in *Brassica* seedlings, causing inhibition of hypocotyl elongation (Potter et al., 1999). Samuolienė et al. (2013) reported similar reductions in HL of kohlrabi grown under a light ratio of R₉₁:FR₁:B₈. The authors reported that as DLI increased from 6 to 19 mol·m⁻²·d⁻¹, HL of kohlrabi decreased 33%.

In the current study, HL of kohlrabi, mizuna, and mustard was also significantly influenced by LQ (Table 2). However, the impact of LQ in combination with LI varied by species. Hypocotyl length of mizuna and mustard was only influenced by LQ when grown at a LI of 210 or 315 μmol·m⁻²·s⁻¹, whereas HL of kohlrabi was only influenced at a LI of 105 μmol·m⁻²·s⁻¹. When differences were present, HL of kohlrabi, mizuna, and mustard was greater when grown under the light ratio of R₇₄:G₁₈:B₈ compared with those grown under R₈₇:B₁₃ or R₈₄:FR₇:B₉ (Fig. 2A–C). For example, HL of mizuna and mustard produced under a LI of 210 μmol·m⁻²·s⁻¹ and a light ratio of R₇₄:G₁₈:B₈ was 13% and 12% greater, respectively, compared with those grown under R₈₄:FR₇:B₉ (Fig. 2B and C).

Hypocotyl elongation related to LQ is generally caused by a low red:far-red ratio. Red and far-red light are absorbed by phytochrome pigments that exist in two interconvertible forms. Far-red (700 to 800 nm) and green light are transmitted through leaf tissue more efficiently than red or blue light, causing enrichment of far-red and green light in plants grown under canopies. When a low ratio of red:far-red light is absorbed by phytochrome pigments, a shade avoidance response is triggered to elongate hypocotyls (Zhang and Folta, 2012). In our study, HL of kohlrabi grown with a LI of 105 μmol·m⁻²·s⁻¹ and mustard grown with a LI of 315 μmol·m⁻²·s⁻¹ and a light ratio of R₈₄:FR₇:B₉ was 11% and 14% greater, respectively, compared with those grown under R₈₇:B₁₃ (Fig. 2A and C). The addition of far-red light in the light ratio of R₈₄:FR₇:B₉ reduced the red:far-red ratio and may have induced a shade avoidance response.

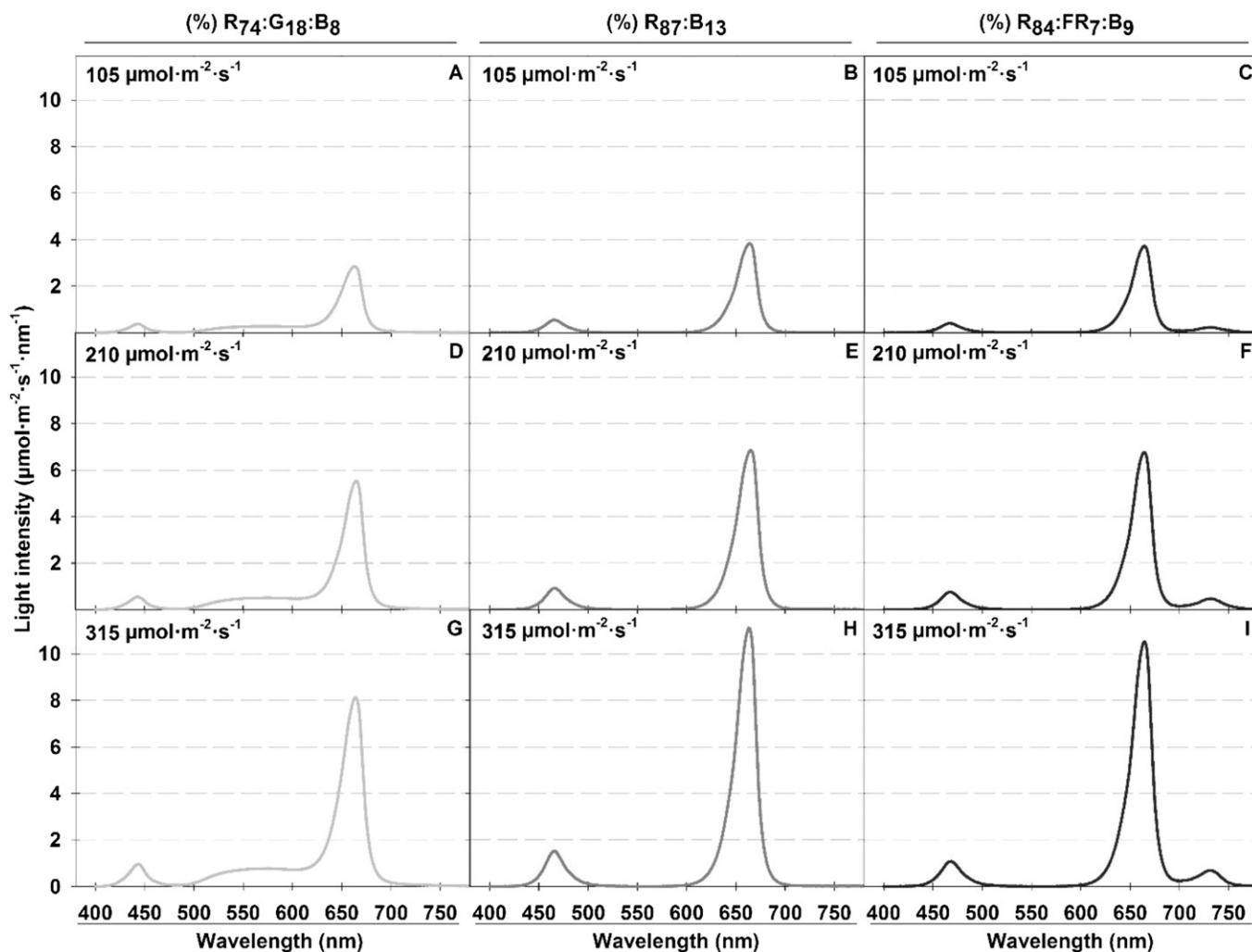


Fig. 1. Spectral quality delivered from sole-source light-emitting diode arrays with light ratios (%) of red:green:blue 74:18:8 ($R_{74}:G_{18}:B_8$), red:blue 87:13 ($R_{87}:B_{13}$), or red:far-red:blue 84:7:9 ($R_{84}:FR_7:B_9$) at a total photon flux from 400 to 800 nm of 105, 210, and 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ at canopy level.

Table 1. Average total photon flux (TPF) from 400 to 800 nm \pm SD delivered from sole-source light-emitting diodes with light ratios (%) of red:green:blue 74:18:8 ($R_{74}:G_{18}:B_8$), red:blue 87:13 ($R_{87}:B_{13}$), or red:far-red:blue 84:7:9 ($R_{84}:FR_7:B_9$) to achieve target light intensities (LIs) of 105, 210, and 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$. The average daily light integrals (DLIs), measured from 400 to 800 nm, under a 16-h photoperiod (0600 to 2200 HR) are also reported. Mean values reported are the average of nine spectral scans across three replications.

Light intensity treatment ($\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$)	Light quality treatment (%)	Avg TPF ($\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$)	Avg DLI ($\text{mol}\cdot\text{m}^{-2}\cdot\text{d}^{-1}$)
105	$R_{74}:G_{18}:B_8$	108.2 ± 23.7	6.2 ± 1.4
	$R_{87}:B_{13}$	110.8 ± 25.2	6.4 ± 1.4
	$R_{84}:FR_7:B_9$	108.4 ± 22.8	6.2 ± 1.3
210	$R_{74}:G_{18}:B_8$	215.1 ± 33.6	12.4 ± 1.9
	$R_{87}:B_{13}$	210.2 ± 31.4	12.1 ± 1.8
	$R_{84}:FR_7:B_9$	207.6 ± 33.2	12.0 ± 1.9
315	$R_{74}:G_{18}:B_8$	312.9 ± 54.4	18.0 ± 3.1
	$R_{87}:B_{13}$	313.2 ± 53.9	18.0 ± 3.1
	$R_{84}:FR_7:B_9$	310.6 ± 50.9	17.9 ± 2.9

In addition to far-red light, the proportion of light in the green waveband could be a possible explanation for hypocotyl elongation in our study. Light in the blue spectral range in combination with red light has been reported to inhibit extension growth of many species (Wollaeger and Runkle, 2014). However, green light in combination with red and blue light has been shown to reverse blue-light

inhibition of hypocotyl elongation. The mechanisms responsible for this are believed to be mediated through cryptochrome blue-light receptors (Zhang and Folta, 2012). In addition, Folta (2004) reported that hypocotyl elongation of *Arabidopsis thaliana* grown under red, blue, or far-red light was suppressed within minutes compared with dark-grown seedlings. However, seedlings grown under

monochromatic green light had increased hypocotyl elongation compared with dark-grown seedlings (Folta, 2004). The proportion of green light in the light ratio of $R_{74}:G_{18}:B_8$ may have resulted in a similar response. Green light absorbed by cryptochrome has been shown to cause responses similar to the shade avoidance response, although the mechanisms are not fully understood (Zhang and Folta, 2012). Although there have been extensive studies conducted concerning the influence of LQ on HL of individual seedlings, limited work has been published on *Brassica* microgreens, which are typically grown in high densities.

Leaf area. In our study, the interaction between LI and LQ was not significant for LA in any of the three species. However, LA of kohlrabi and mustard was significantly influenced by the main effect of LI (Table 2). Specifically, LA of kohlrabi and mustard decreased as LI increased. For example, as LI increased from 105 to 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, LA of kohlrabi decreased from 1.52 to 1.14 cm^2 (25% decrease), and mustard decreased from 1.50 to 1.30 cm^2 (13% decrease). This finding

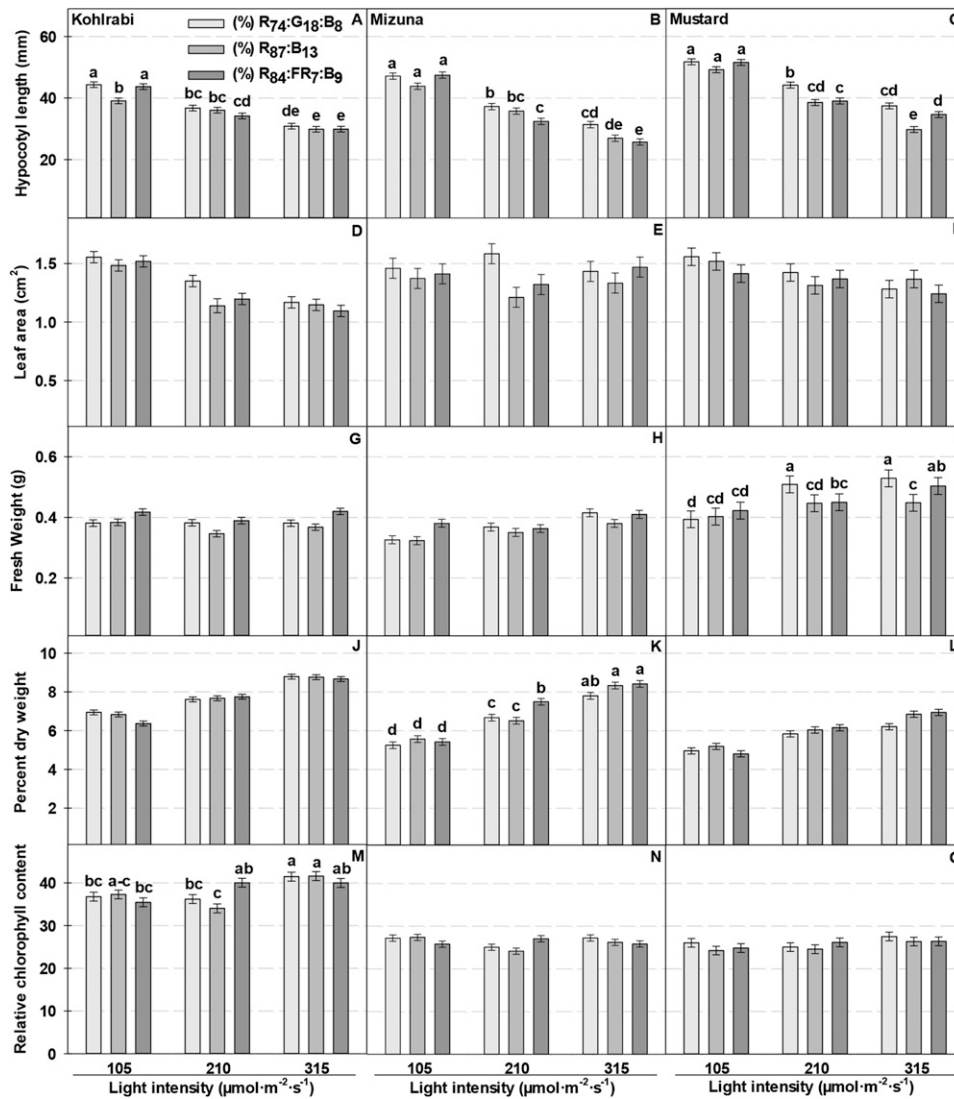


Fig. 2. Hypocotyl length (A–C), leaf area (D–F), fresh weight (G–I), percent dry weight (dry weight/fresh weight \times 100) (J–L), and relative chlorophyll content (M–O) of kohlrabi (*Brassica oleracea* L. var. *gongylodes* L.), mizuna (*B. rapa* L. var. *japonica*), and mustard [*B. juncea* (L.) Czern. ‘Gamet Giant’] microgreens placed under light intensities (LIs) of 105, 210, or 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ delivered from sole-source light-emitting diodes with light ratios (%) of red:green:blue 74:18:8 (R₇₄:G₁₈:B₈), red:blue 87:13 (R₈₇:B₁₃), or red:far-red:blue 84:7:9 (R₈₄:FR₇:B₉). Error bars indicate \pm SE. Means sharing a letter are not statistically different by Tukey’s honestly significant difference test at $P \leq 0.05$. Figures with no letter were found to have no significant interaction between LI and light quality.

Table 2. Analysis of variance for the effects of light intensity (LI), light quality (LQ), or LI \times LQ from sole-source light-emitting diodes on growth and morphology of kohlrabi (*Brassica oleracea* L. var. *gongylodes* L.), mizuna (*Brassica rapa* L. var. *japonica*), and mustard [*Brassica juncea* (L.) Czern. ‘Gamet Giant’] microgreens.

Data	Kohlrabi			Mizuna			Mustard		
	LI	LQ	LI \times LQ	LI	LQ	LI \times LQ	LI	LQ	LI \times LQ
Hypocotyl length	***	**	**	***	***	**	***	***	*
Leaf area	***	*	NS	NS	*	NS	**	NS	NS
Fresh weight	*	***	NS	***	**	NS	***	***	**
Percent dry weight	***	NS	NS	***	**	*	***	*	NS
Relative chlorophyll	***	NS	**	NS	NS	NS	NS	NS	NS

NS, *, **, ***Nonsignificant or significant at $P \leq 0.05$, 0.01, or 0.001, respectively.

coincides with previous research that reported the LA of seedlings generally increased when grown under low LIs (Jarvis, 1964).

In addition, the LA of kohlrabi and mizuna was significantly impacted by the main effect of LQ (Table 2). The LA of kohlrabi and mizuna microgreens grown under a light ratio

of R₇₄:G₁₈:B₈ increased from 1.26 to 1.36 cm^2 (7% increase) and from 1.31 to 1.49 cm^2 (12% increase), respectively, compared with those grown under R₈₇:B₁₃. This finding contrasts what was observed for HL under the light ratio of R₇₄:G₁₈:B₈, as the green light-induced shade avoidance mechanism observed for

HL can elicit reductions in LA (Zhang and Folta, 2012). Thus, if the addition of green light was indeed initiating a similar response to that observed with decreasing red:far-red ratios, a decrease in LA would have been expected. Although green light is believed to elicit photomorphogenic responses similar to shade avoidance, further research is needed to better understand this mechanism and the associated changes in plant growth (Zhang and Folta, 2012).

FW and percent DW. In our study, FW and percent DW of all species generally increased as LI increased (Fig. 2G–L). Fresh weight of mustard grown under light ratios of R₇₄:G₁₈:B₈ and R₈₄:FR₇:B₉ increased 34% and 19%, respectively, as LI increased from 105 to 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ (Fig. 2I). In addition, FW of mizuna increased 15% as LI increased from 105 to 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, regardless of LQ. Similarly, percent DW of kohlrabi and

mustard increased from 6.7% to 8.7% and from 5.0% to 6.7%, respectively, as LI increased from 105 to 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, regardless of LQ. Samuoliéné et al. (2013) also reported similar increases in percent DW of tatsoi and red pak choi microgreens as DLI increased from 6 to 19 $\text{mol}\cdot\text{m}^{-2}\cdot\text{d}^{-1}$. Plant growth, defined as an irreversible increase in plant size, is a function of biomass production driven by photosynthesis (Heins et al., 2000). Thus, the response observed in this study was expected as the treatments with the higher LIs provided additional light for photosynthetic activity and, as a result, biomass accumulation.

The main effect of LQ on FW was significant for kohlrabi and mizuna. Generally, FW was greater when grown under a light ratio of R₈₄:FR₇:B₉ compared with those grown under R₈₇:B₁₃ (Fig. 2G–I). For example, FW of kohlrabi and mizuna was 9.8% and 7.9% greater, respectively, under the ratio R₈₄:FR₇:B₉ compared with R₈₇:B₁₃. In addition, FW of mustard grown with a light ratio of R₇₄:G₁₈:B₈ and a LI of 210 or 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ was 14% or 18% greater, respectively, compared with those grown under R₈₇:B₁₃ (Fig. 2I), but not different across LQ at a LI of 105 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$.

In our study, percent DW of kohlrabi was not influenced by LQ. However, the impact of LQ on percent DW of mizuna varied with LI. Light quality did not affect DW at low or high LI (105 or 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$), but percent DW of mizuna grown under a LI of 210 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ with a light ratio of R₈₄:FR₇:B₉ was 1.0% and 0.8% higher than those grown under R₈₇:B₁₃ and R₇₄:G₁₈:B₈, respectively (Fig. 2K). In addition, the main effect of LQ for percent DW of mustard was significant. The light ratio of R₈₇:B₁₃ produced 6.02% DW, 0.36% higher than those grown under a light ratio of R₇₄:G₁₈:B₈ (5.66% DW). Limited work has been published on *Brassica* microgreens to investigate the effects of LQ on biomass accumulation. Kopsell et al. (2014) observed that broccoli microgreens displayed no significant changes in DW under light ratios of R₉₅:B₅, R₈₅:G₁₀:B₅, R₈₀:B₂₀, R₇₀:G₁₀:B₂₀, or fluorescent/incandescent lamps providing a PPF of 250 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$. However, the FW of broccoli microgreens produced under light ratios of R₈₅:G₁₀:B₅ and R₈₀:B₂₀ were significantly higher than those produced under R₇₀:G₁₀:B₂₀ or fluorescent/incandescent lamps. Li and Kubota (2009) reported that LQ had significant effects on FW and DW of leaf lettuce harvested 25 d after germination at the ‘baby green’ growth stage. The FW and DW of baby leaf lettuce significantly increased when grown under SS fluorescent white light supplemented with far-red LEDs compared with fluorescent white light as a control or fluorescent white lights supplemented with ultraviolet-A or red LEDs. In that study, while the PPF was 305 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ for all treatments, the far-red LEDs provided an additional 160 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ of light. Recent in vitro studies using spinach (*Spinacia oleracea* L.) and in vivo studies using sunflower (*Helianthus annuus* L.) and bean (*Phaseolus vulgaris* L.) have reported that

far-red light, up to 790 nm, can drive photosynthetic activity in photosystem II (PSII) (Pettai et al., 2005; Thapper et al., 2009). Increased photosynthetic activity of baby leaf lettuce may have occurred due to the additional 160 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ of far-red light, resulting in increased FW and DW. In the present study, TPF was measured from 400 to 800 nm to include light qualities in the far-red range to ensure LI was consistent among SS LED light treatments. Our results and those by Li and Kubota (2009) indicate that more studies are needed to clarify how far-red light in SS lighting scenarios impacts biomass accumulation of *Brassica* microgreens and baby leaf lettuce.

Relative chlorophyll content. Relative chlorophyll content of mizuna and mustard was not significantly influenced by LI or LQ (Table 2). However, RCC of kohlrabi under a light ratio of R₇₄:G₁₈:B₈ and a LI of 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ increased by 11% or 13%

compared with those provided a LI of 105 or 210 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, respectively (Fig. 2M). In addition, RCC of kohlrabi grown with a LI of 210 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ and a light ratio of R₈₄:FR₇:B₉ increased 15% compared with those under R₈₇:B₁₃ (Fig. 2M). Samuoliéné et al. (2013) reported similar results in terms of LI with an 11% increase in RCC of mustard grown with a light ratio of R₉₁:FR₁:B₈ and a PPF of 545 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ compared with those grown under the same light ratio with a PPF of 220 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$. However, the majority of the results presented from the present study show relatively stable RCC levels across treatments.

Nutrient content. Generally, for all three *Brassica* species in the current study, nutrient concentration of both macro- and micronutrients decreased as LI increased (Tables 3 and 4). This trend may be the result of a dilution of nutrients due to the increased percent DW observed at higher LIs (Fig. 2J–L). This

Table 3. Macronutrient concentration [percent dry weight (DW)] of kohlrabi (*Brassica oleracea* L. var. *gongylodes* L.), mizuna (*Brassica rapa* L. var. *japonica*), and mustard [*Brassica juncea* (L.) Czern. ‘Garnet Giant’] microgreens placed under light intensities (LIs) of 105, 210, or 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ delivered from sole-source light-emitting diodes (LEDs) with light quality (LQ) ratios (% of red:green:blue 74:18:8 (R₇₄:G₁₈:B₈), red:blue 87:13 (R₈₇:B₁₃), or red:far-red:blue 84:7:9 (R₈₄:FR₇:B₉).

LI	LED	Macronutrients (percent DW)					
		Nitrogen	Phosphorus	Potassium	Sulfur	Calcium	Magnesium
<i>Kohlrabi</i>							
105	R ₇₄ :G ₁₈ :B ₈	5.25 ^z	1.17	1.22	1.75	2.01	0.56
	R ₈₇ :B ₁₃	5.41	1.21	1.26	1.83	2.02	0.57
	R ₈₄ :FR ₇ :B ₉	5.41	1.18	1.33	1.78	2.13	0.58
210	R ₇₄ :G ₁₈ :B ₈	4.83	1.10	0.89	1.62	1.72	0.49
	R ₈₇ :B ₁₃	4.75	1.11	0.93	1.58	1.73	0.50
	R ₈₄ :FR ₇ :B ₉	4.69	1.08	0.95	1.57	1.89	0.51
315	R ₇₄ :G ₁₈ :B ₈	4.30	0.97	0.84	1.46	1.77	0.48
	R ₈₇ :B ₁₃	4.38	0.97	0.78	1.43	1.69	0.48
	R ₈₄ :FR ₇ :B ₉	4.23	0.96	0.97	1.42	1.86	0.49
LQ		NS	NS	**	NS	**	NS
LI		***	***	***	***	***	***
LQ × LI		NS	NS	NS	NS	NS	NS
<i>Mizuna</i>							
105	R ₇₄ :G ₁₈ :B ₈	4.30 a ^y	1.01	1.88 a	1.19	2.88 a	0.73
	R ₈₇ :B ₁₃	3.88 a	0.96	1.77 a	1.12	2.75 a	0.70
	R ₈₄ :FR ₇ :B ₉	4.00 a	0.97	1.98 a	1.17	2.89 a	0.71
210	R ₇₄ :G ₁₈ :B ₈	3.23 b	0.76	1.44 b	0.91	2.17 b	0.58
	R ₈₇ :B ₁₃	3.20 b	0.75	1.42 b	0.89	2.13 b	0.57
	R ₈₄ :FR ₇ :B ₉	2.58 cd	0.69	1.20 d	0.83	2.15 b	0.54
315	R ₇₄ :G ₁₈ :B ₈	2.69 c	0.62	1.41 bc	0.78	2.16 b	0.54
	R ₈₇ :B ₁₃	2.56 cd	0.59	1.21 cd	0.74	2.03 bc	0.50
	R ₈₄ :FR ₇ :B ₉	2.21 d	0.60	1.18 d	0.76	1.89 c	0.48
LQ		***	NS	**	NS	**	**
LI		***	***	***	***	***	***
LQ × LI		**	NS	**	NS	**	NS
<i>Mustard</i>							
105	R ₇₄ :G ₁₈ :B ₈	4.63	0.94	1.27 b	1.20	2.32 a	0.71 ab
	R ₈₇ :B ₁₃	4.29	0.86	1.24 b	1.17	2.32 a	0.66 b
	R ₈₄ :FR ₇ :B ₉	4.50	0.93	1.44 a	1.25	2.44 a	0.74 a
210	R ₇₄ :G ₁₈ :B ₈	3.53	0.71	1.13 bc	0.93	1.94 b	0.56 c
	R ₈₇ :B ₁₃	3.41	0.66	1.01 cd	0.87	1.79 bcd	0.53 cd
	R ₈₄ :FR ₇ :B ₉	3.32	0.68	1.01 cd	0.87	1.80 bcd	0.53 cd
315	R ₇₄ :G ₁₈ :B ₈	3.26	0.62	1.15 bc	0.81	1.94 bc	0.52 cd
	R ₈₇ :B ₁₃	3.04	0.61	0.92 d	0.76	1.74 cd	0.50 cd
	R ₈₄ :FR ₇ :B ₉	2.85	0.56	0.91 d	0.72	1.72 d	0.48 d
LQ		*	*	**	NS	*	NS
LI		***	***	***	***	***	***
LQ × LI		NS	NS	***	NS	**	*

^zMean values are based on a representative sample from each treatment across two experimental replications.

^yMeans sharing a letter are not statistically different by Tukey’s honestly significant difference test at $P \leq 0.05$. Means with no lettering were found to have no significant interaction between LI and LQ.

NS, *, **, ***Nonsignificant or significant at $P \leq 0.05, 0.01, \text{ or } 0.001$, respectively.

Table 4. Micronutrient concentrations (mg·kg⁻¹) of kohlrabi (*Brassica oleracea* L. var. *gongylodes* L.), mizuna (*Brassica rapa* L. var. *japonica*), and mustard [*Brassica juncea* (L.) Czern. 'Garnet Giant'] microgreens placed under light intensities (LIs) of 105, 210, or 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ delivered from sole-source light-emitting diodes (LEDs) with light quality (LQ) ratios (%) of red:green:blue 74:18:8 (R₇₄:G₁₈:B₈), red:blue 87:13 (R₈₇:B₁₃), or red:far-red:blue 84:7:9 (R₈₄:FR₇:B₉).

LI	LED	Micronutrients (mg·kg ⁻¹)				
		Boron	Copper	Iron	Manganese	Zinc
<i>Kohlrabi</i>						
105	R ₇₄ :G ₁₈ :B ₈	45.49 ^c	5.87	120.57 a	57.26	67.28
	R ₈₇ :B ₁₃	48.35 ab	5.91	111.78 abc	57.49	64.89
	R ₈₄ :FR ₇ :B ₉	49.49 a	5.94	119.76 ab	56.32	62.10
210	R ₇₄ :G ₁₈ :B ₈	41.92 d	5.87	102.83 cde	50.03	56.67
	R ₈₇ :B ₁₃	46.40 bc	5.89	99.96 de	51.71	56.22
	R ₈₄ :FR ₇ :B ₉	39.69 de	5.47	108.89 bcd	51.94	52.42
315	R ₇₄ :G ₁₈ :B ₈	45.12 c	4.85	109.52 bcd	49.18	45.94
	R ₈₇ :B ₁₃	39.38 e	5.01	107.78 cd	48.02	46.94
	R ₈₄ :FR ₇ :B ₉	39.08 e	4.45	93.87 e	51.35	43.91
LQ		**	NS	NS	NS	*
LI		***	***	***	***	***
LQ × LI		***	NS	***	NS	NS
<i>Mizuna</i>						
105	R ₇₄ :G ₁₈ :B ₈	80.40	8.15	159.98	96.63	51.27 ab
	R ₈₇ :B ₁₃	75.47	7.55	129.14	93.21	54.07 a
	R ₈₄ :FR ₇ :B ₉	60.06	7.64	127.26	88.73	48.88 abc
210	R ₇₄ :G ₁₈ :B ₈	58.79	6.69	103.36	73.69	50.41 abc
	R ₈₇ :B ₁₃	62.07	5.76	98.46	74.08	39.62 cde
	R ₈₄ :FR ₇ :B ₉	46.81	6.14	106.89	70.91	42.15 bcd
315	R ₇₄ :G ₁₈ :B ₈	53.21	5.69	114.82	63.73	30.66 e
	R ₈₇ :B ₁₃	45.27	5.84	97.48	60.93	35.66 de
	R ₈₄ :FR ₇ :B ₉	26.09	5.71	80.20	58.87	40.31 bcde
LQ		***	NS	**	NS	NS
LI		***	***	***	***	***
LQ × LI		NS	NS	NS	NS	**
<i>Mustard</i>						
105	R ₇₄ :G ₁₈ :B ₈	86.03 b	9.08	132.32	48.11	42.48 ab
	R ₈₇ :B ₁₃	70.72 d	7.67	106.95	47.10	42.24 ab
	R ₈₄ :FR ₇ :B ₉	95.37 a	8.24	189.88	47.15	47.10 a
210	R ₇₄ :G ₁₈ :B ₈	74.51 cd	6.84	84.67	42.67	33.26 bc
	R ₈₇ :B ₁₃	76.84 c	6.72	81.40	39.92	35.95 bc
	R ₈₄ :FR ₇ :B ₉	58.49 e	8.31	100.22	39.13	38.88 ab
315	R ₇₄ :G ₁₈ :B ₈	57.08 e	5.98	91.99	38.16	41.91 ab
	R ₈₇ :B ₁₃	48.23 f	5.58	98.44	38.53	27.40 c
	R ₈₄ :FR ₇ :B ₉	51.83 f	5.58	84.42	35.84	33.60 bc
LQ		***	NS	NS	NS	*
LI		***	***	*	***	***
LQ × LI		***	NS	NS	NS	**

^aMean values are based on a representative sample from each treatment across two experimental replications.

^bMeans sharing a letter are not statistically different by Tukey's honestly significant difference test at $P \leq 0.05$. Means with no lettering were found to have no significant interaction between LI and LQ.

NS, *, **, ***Nonsignificant or significant at $P \leq 0.05, 0.01, \text{ or } 0.001$, respectively.

nutrient dilution was previously investigated in chrysanthemum (*Chrysanthemum ×morifolium* Ramat. 'Fiesta') by Kuehny et al. (1991). Specifically, they found that nutrient concentration of chrysanthemum was higher under elevated CO₂ and increased irradiance. However, most of the differences observed between treatments were removed when data were expressed on a starch-free DW basis. Thus, they proposed that the increased CO₂ and irradiance led to a dilution of nutrient content due to the associated increase in growth (Kuehny et al., 1991). This finding may help to explain the higher nutrient levels we observed with all three species of *Brassica* microgreens under lower LIs. However, given that microgreens are typically sold on a FW basis, production under lower LIs may result in crops that are more nutrient dense.

At a LI of 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, mizuna grown with a light ratio of R₇₄:G₁₈:B₈ accumulated

22%, 20%, and 14% more N, K, and Ca, respectively, compared with those grown under R₈₄:FR₇:B₉ (Table 3). Similarly, mustard accumulated 26%, 13%, and 10% more K, Ca, and B, respectively, when grown under a light ratio of R₇₄:G₁₈:B₈ compared with those grown under R₈₄:FR₇:B₉ when provided a LI of 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ (Tables 3 and 4). Kohlrabi accumulated 15% and 17% more B and Fe, respectively, when grown under a light ratio of R₇₄:G₁₈:B₈ compared with those grown under R₈₄:FR₇:B₉ under a LI of 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ (Table 4). Lastly, while differences were not always significant, there was a trend for microgreens grown with a LI of 315 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ under the light ratio of R₇₄:G₁₈:B₈ to consistently display higher nutrient content compared with those under R₈₇:B₁₃ for the same nutrients and species. As previously stated, Kopsell and Sams (2013) found that broccoli microgreens grown under a light ratio of R₀:B₁₀₀ produced significantly higher concentrations

of all essential elements compared with those under R₁₂:B₈₈ at a PPF of 41 and 350 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, respectively. Further research by Kopsell et al. (2014) validated the hypothesis that nutrient-dense microgreens could be produced under high percentages of blue light. Specifically, blue wavelengths of light have been shown as a dominant means of regulating the processes involved in nutrient content, such as proton pumping, ion channel activities, and membrane permeability (Kopsell et al., 2014). However, results from the present study display that, under higher LIs, the content of specific macro- and micronutrients may benefit from the presence of green wavelengths as well. However, it is uncertain whether the increased nutrient content under the light ratio of R₇₄:G₁₈:B₈ was due to the presence of green wavelengths or the absence of far-red wavelengths. Thus, future research should look further into the implications of increasing green light and its potential impact on nutrient content for these microgreens species.

Results from this study on growth, morphology, and nutrient content can be used by growers to select the LI and LQ required to achieve preferred growth characteristics of *Brassica* microgreens. Now, recommendations on light recipes for the production of *Brassica* microgreens will depend strongly on the market targeted for consumption and the morphological attributes desired by the grower. The data presented should provide adequate information to allow those interested in using these technologies to make informed decisions regarding the production of microgreens based on market needs.

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