Growth and Nutrient Partitioning of Containerized *Malus trilobata* Schneid. and *Acer syriacum* Boiss. and Gaill. Under Two Fertigation Regimes

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Abstract. The native flora of Lebanon is threatened. As part of an ex situ conservation strategy, this study was conducted to determine if *Malus trilobata* (Schneid.) and *Acer syriacum* (Boiss. and Gaill.), two native Lebanese trees, were amenable to container production. Therefore, these species' growth and nitrogen, phosphorus, and potassium uptake efficiency and distribution under two fertilizer rates were studied. *Malus trilobata* seedlings were obtained from seeds collected from two mother trees, whereas *Acer syriacum* seeds were collected from a single tree. Two-year-old seedlings raised from these seeds were planted in 11-L containers in a 3:1 pine bark:compost substrate. Half the seedlings within each source and species were assigned to either 25 or 100 mg·L⁻¹ N from 21N–3.1P–5.9K water-soluble fertilizer treatments. Seedlings of both *Malus trilobata* sources grown under 25 mg·L⁻¹ N were taller than those grown at 100 mg·L⁻¹ N. Nitrogen loading occurred in plants of *Malus trilobata* under the high fertilizer rate, although total plant N, phosphorus, and potassium content was unaffected by fertilizer rate, because larger seedlings had lower nutrient concentrations. There were significant differences in growth among the two *Malus trilobata* seed sources, but there were no differences in mineral nutrient uptake or nutrient use efficiency. Growth, mineral nutrient uptake, and nutrient use efficiency of *Acer syriacum* seedlings were unaffected by fertilizer rate. *Malus trilobata* and *Acer syriacum* seedlings are amenable to container production. Using container production for these species with subsequent transplanting into managed landscapes might be a viable ex situ conservation method. Further studies are needed.

With the end of the civil war in Lebanon in 1990, the demand for low-maintenance landscape plants has been increasing as a result of extensive reconstruction projects. The Lebanese flora was last described by Mouterde (1966, 1970). Since then, the natural vegetation has experienced habitat fragmentation and destruction as a result of tourism and urban expansion, overgrazing, overexploitation of natural resources, and the impact of successive wars. The Lebanese government has been taking in situ conservation measures turning selected threatened habitats into "protected areas" or "nature

reserves" (UNEP, 1996), but there are no ex situ biodiversity conservation programs such as growing native plants for landscape use. Currently in Lebanon, landscape woody plants are being imported from many countries (Italy, Spain, Syria, Egypt, United States), although a large number of Lebanese taxa have potential economic value for ornamental or medicinal use (UNEP, 1996). There is a great potential for developing a native plant production industry because of the country's floristic richness estimated at 3761 vascular plant species (UNEP, 1996).

Research in the field of nursery production is very limited in Lebanon. In contrast, a considerable number of research papers discuss recommended nitrogen (N) fertilizer rates for woody ornamentals grown in container production systems in the United States (Gilliam et al., 1980, 1984; Ingestad, 1979; Jull et al., 1994; Larimer and Struve, 2002; Lumis et al., 2000; Musselwhite et al., 2004; Stubbs et al., 1997; Wright and Niemiera, 1987). The recommended N fertilizer rates ranged from 20 to 400 mg N per liter

depending on the species used and the frequency of fertilizer applications.

In a previous study, we demonstrated that another native Lebanese species, *Cercis siliquastrum*, Judas tree, was amenable to container production (Zahreddine et al., 2007a). This article describes a similar experiment with two additional native Lebanese species with ornamental traits, *Malus trilobata* (Schneid.) and *Acer syriacum* (Boiss. and Gaill.). The two species are not commonly grown in Lebanese nurseries and there are no reports on container production or nutrient partitioning of the species elsewhere.

Malus trilobata, "erect crab" or "three-lobed apple tree", belongs to the Rosaceae family. It has an upright habit with horizontal branching and a mature height of 13 m (Anonymous, 1999). The leaves are maple-like and deeply three-lobed. They turn from orange to red to deep purple in the fall. The tree blooms during April and May producing white flowers and yellow fruits. Hillier and Sons (1973) reports that the species is distributed in the eastern Mediterranean region and northeast Greece adding that the species is comparatively rare and distinct; Mouterde (1966) also reported that it is rare but endemic only to Lebanon. In Lebanon, it is found in the supra-Mediterranean zone at an altitude of 1,000 to 1,500 m in association with *Ostrya*, *Sorbus*, *Fraxinus*, and *Abies* species in sandy loam, high pH soils (Zahreddine et al., 2007b).

Acer syriacum (Boiss. and Gaill.), Syrian maple, is in the Aceraceae family. It reaches 8 m tall. The leaves are semievergreen to evergreen, glabrous, with three short, acute, or broad lobes. If fall color develops, it ranges from yellow to pink. White to yellow flowers are produced from February to March and the tree develops divergent samaras. It is also native to Syria, Cyprus, and Palestine (Mouterde, 1966). The species grows in the thermo-Mediterranean zone in Lebanon (0 to 500 m altitude). It is reported to be indifferent to soil type but is mainly found in calcareous soils (Anonymous, 1999). Its wood is commonly used for firewood and cabinet making. There is some confusion regarding its taxonomy.

The purpose of this study was to determine the effects of two fertilizer rates on growth, dry weight distribution, mineral nutrient content and distribution, and mineral nutrient uptake efficiency of *Malus trilobata* and *Acer syriacum* container-grown seedlings.

Materials and Methods

A description of plant material and the numbers of seedlings used in this study are given in Table 1. The procedure used in this study is similar to the one used for *Cercis siliquastrum* (Zahreddine et al., 2007a). However, the *Cercis siliquastrum* study looked at the effect of two fertilizer rates on the growth and nutrient content of seedlings grown from six different sources or mother trees in Lebanon. Hence, the study was too

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Table 1. Description of the habitat where two *Malus trilobata* (MT) trees and one *Acer syriacum* (AS) tree used for seed collection were located in Lebanon and the number of seedlings from each source used in the study.

Species	Source	Elevation (m)	Latitude (N)	Longitude (E)	Number of plants in study
<i>Malus trilobata</i>	MT2: Ehden	1,499	34°18.776'	035°59.320'	80
	MT3: Ehden	1,457	34°18.813'	035°59.147'	80
<i>Acer syriacum</i>	AS2: Nahr Damour	43	33°41.942'	035°29.052'	80

long, making it inappropriate to combine it with the current study.

Seeds of *Malus trilobata* and *Acer syriacum* were collected from two geographical locations (Ehden and Nahr Damour) in Lebanon in Fall 2001 and brought to The Ohio State University (data not presented) where seed germination trials were carried out. In Spring 2002, germinated seeds were transplanted into 250-XL plastic containers (1.95 L; Nursery Supplies, Fairless Hills, PA) using MetroMix® 360 (Scotts Company, Marysville, OH) substrate and placed in a heated greenhouse under natural photoperiods. The greenhouse temperatures were set at 25/21 °C. The relative humidity averaged $46.6 \pm 2\%$. Plants were watered as needed to avoid water stress. When seedlings developed two true leaves, they were fertilized once per week with 100 mg·L⁻¹ N of 20N-8.3P-4.6K (20-10-10, Peters water-soluble fertilizer; O.M. Scotts Company, Marysville, OH). Plants were overwintered (from November to May) in a minimum heat (minimum temperature greater than 2 °C) polyhouse.

In May 2004, the plants (2-year-old seedlings) were removed from the containers, root pruned, transplanted into No. 3 gallon (11 L) Spin-Out® treated containers (1200 Classic; Nursery Supplies), and moved to an outdoor gravel pad. The most vigorous shoot was trained into a central leader by tying it to a 2-m bamboo stake. The plants were placed on 45.7-cm within-row spacing and 2-m between-row spacing. A 3 pine bark:1 Comtil (by volume) substrate was used. Comtil is a composted municipal sewage sludge from the city of Columbus, OH (nutrient analysis 0.45–1–0.25) and was used as a slow-release micronutrient source.

Half of the plants from each mother tree or seed source were randomly assigned to one of two N fertilizer rates: 25 mg or 100 mg·L⁻¹ N fertigation from 21N-3.1P-5.9K (21-7-7, Peters Water Soluble Fertilizer; O.M. Scotts Company) applied at 0.5 L in each of two daily irrigation cycles (1 L·d⁻¹ total). The fertility treatments were initiated on 1 June and ended on 20 Sept. Plants within a fertility treatment were arranged in a split-plot design with four replications. Each replicate constituted of 10 plants per source and fertilizer level.

An initial harvest of 10 plants per source and fertilizer treatment was done on 12 May 2004 before the start of the experiment and a second harvest on 24 Sept. 2004. At each harvest, height and caliper were measured on the harvested as well as all other trees, whereas leaf area was measured on the 10

randomly harvested trees per source and fertilizer treatment with a LI-3100 Area meter (LI-COR, Lincoln, NE). Plants were pruned at the root collar, substrate washed from the root systems, plant parts (roots, shoots, and leaves) oven dried at 82 °C for 96 h, and dry weights recorded. From the 10 plant harvest subsample, three randomly selected individuals from each source and fertilizer rate were selected for nutrient analysis. Leaves, shoots, and roots from these plants were ground to pass through a 20-mesh screen and 2-g subsamples from each tissue type were sent to the Service Testing And Research Laboratory (STAR Laboratory, OARDC, Wooster, OH) for N, P, and K nutrient analysis. Total nutrient content of each tissue type was calculated by multiplying the tissue's dry weight by the respective tissue's nutrient concentration. Total plant dry weight or nutrient content was calculated by summing the dry weights or nutrient contents of an individual's stem, roots, and leaves. Leaf, stem, and root relative dry weights, as percentages of total plant dry weight, were calculated. Relative nutrient contents of leaves, stems, and roots were determined similar to the relative dry weights.

Nitrogen use efficiency was calculated by the following equation: $NUE = [(TNC_2) - (TNC_1)/TNA] \times 100\%$ where NUE = nitrogen use efficiency, TNC₂ and TNC₁ are total plant N contents in September and May, respectively, and TNA is the total N applied between May and September. Similarly, P and K use efficiencies were calculated.

From the dry weights and leaf area data, net assimilation rate (NAR), leaf area ratio (LAR), and relative growth rate (RGR) were calculated according to Evans (1972). To calculate NAR, LAR, and RGR, plants within the initial and final harvests were sorted by total plant dry weight and paired by ascending order. Total plant assimilation rates (g·cm⁻²·d⁻¹) were calculated as: $NAR = (TW_2 - TW_1) \times (\ln A_2 - \ln A_1) / [(T_2 - T_1) \times (A_2 - A_1)]$; leaf area ratio (g·cm⁻²) as: $LAR = (A_2 - A_1) / (TW_2 - TW_1)$; and relative growth rate (g·g⁻¹·d⁻¹) as $RGR = (\ln TW_2 - \ln TW_1) / (T_2 - T_1)$; where T₁ = initial time (12 May), T₂ = final time (24 Sept.), TW₁ and TW₂ are total plant dry weight at times 1 and 2, respectively, and A₁ and A₂ = leaf area at times 1 and 2, respectively.

For dry weight analyses, 10 single plant replications were used, whereas mineral nutrient analyses used three single plant replications. Data were analyzed using the one-way analysis of variance procedure

within SPSS (version 12.0; SPSS Institute, Chicago, IL). The linear regression procedure within SigmaPlot (version 9.0) (SPSS, Chicago) was used to develop equations describing plant height during the season; the equations were developed using height data from all the seedlings within a treatment combination.

Results

Growth and dry weight. Quadratic equations describing *Malus trilobata* and *Acer syriacum* height under both fertilizer rates were significant ($P < 0.001$, $R^2 > 0.93$, Figs. 1 and 2). *Malus* height growth was rapid; it doubled between June and September. Growth for *Acer syriacum* seedlings was not as vigorous; it increased $\approx 25\%$ between June and September.

The source by fertilizer rate interaction was significant only for *Malus trilobata* leaf area ($P = 0.043$; Table 2). Fertilizer rate did not affect any of the parameters measured for *Acer syriacum* (Table 2). Fertilizer rate affected only plant height of *Malus trilobata*; plants were taller at the low fertilizer rate (168.4 cm) than at the high rate (130.9 cm; Table 2). *Malus trilobata* plants from source 2 (MT2) had significantly higher root and total plant dry weights than those from MT3 at both fertilizer rates ($P = 0.012$ and $P = 0.027$, respectively; Table 2). Root and total plant dry weights were 60.2 g and 199.6 g, respectively, for MT2 and 34.9 g and 145.8 g, respectively, for MT3 (Table 2).

There were no significant source by fertilizer rate interaction for relative root, shoot, or leaf dry weight distribution in *Malus trilobata* plants (Table 3). Fertilizer rates affected only leaf dry weight distribution for *Malus trilobata*; it was higher at the high fertilizer rate (14.1% versus 11.6%). Fertilizer rate did not affect root, shoot, and leaf dry weight distribution in *Acer syriacum* plants (Table 3). There were no significant differences between the two *Malus trilobata* seed sources in root, shoot, and leaf dry weight distribution (Table 3). For both *Malus trilobata* sources and *Acer syriacum*, relative shoot dry weight was higher than either leaf or root dry weights at both fertilizer rates, 60% for *Malus trilobata* and 43% for *Acer syriacum* (Table 3).

There was a significant source by fertilizer rate interaction for RGR of *Malus trilobata* plants ($P = 0.025$; Table 4). For both sources, RGR was higher at the low fertilizer rate, but for MT3 plants, the difference was greater than for MT2 plants (0.0049 g·g⁻¹·d⁻¹ versus 0.0022 g·g⁻¹·d⁻¹, respectively). Fertilizer rate did not affect LAR, NAR, and RGR of *Acer syriacum* plants, but it did in *Malus trilobata* (Table 4). *Malus trilobata* plants grown at the high fertilizer rate had significantly higher LAR and lower NAR (averaging 9.8 g·cm⁻² and 0.0020 g·cm⁻²·d⁻¹, respectively) than those grown at the low fertilizer rate (averaging 7.9 g·cm⁻² and 0.0030 g·cm⁻²·d⁻¹, respectively).

Fig. 1. Height of container-grown *Acer syriacum* grown under 25 or 100 mg·L⁻¹ N from 21N-3.1P-5.9K water-soluble fertilizer. Each value is the mean of 25 plants per fertilizer rate. Plants were fertilized with 21-7-7 Peters fertilizer. Plant height at 25 mg·L⁻¹ N is predicted by the equation: $HT = -101.75 + 1.38X - 0.0025X^2$, $R^2 = 0.962$, $P < 0.0001$; whereas plant height at 100 mg·L⁻¹ N is predicted by the equation: $HT = -37.07 + 0.74X - 0.0013X^2$, $R^2 = 0.952$, $P = 0.038$; where HT = plant height (in cm) and X = Julian day between 174 and 260. Julian calendar days 174, 204, 234, and 260 refer to the dates when measurements were recorded in June, July, August, and September, respectively.

Fig. 2. Height of container-grown *Malus trilobata* (MT2 and MT3) under 25 or 100 mg·L⁻¹ N from 21N-3.1P-5.9K water-soluble fertilizer. Each value is the mean of 25 plants per fertigation treatment. Plants were fertilized with 21-7-7 Peters water-soluble fertilizer. Plant height at 25 mg·L⁻¹ N is predicted by the equation: $HT \text{ for MT2 plants} = -289.48 + 3.07X - 0.0053X^2$, $R^2 = 0.944$, $P = 0.045$. $HT \text{ for MT3 plants} = -490.15 + 4.74X - 0.0087X^2$, $R^2 = 0.933$, $P < 0.0001$. Plant height at 100 mg·L⁻¹ N is predicted by the equation: $HT \text{ for MT2 plants} = -343.96 + 3.58X - 0.0067X^2$, $R^2 = 0.939$, $P = 0.028$. $HT \text{ for MT3 plants} = -238.62 + 2.41X - 0.0038X^2$, $R^2 = 0.928$, $P < 0.0001$; where HT = plant height (in cm) and X = Julian day between 174 and 260. Julian calendar days 174, 204, 234, and 260 refer to the dates when measurements were recorded in June, July, August, and September, respectively.

Tissue nitrogen concentration, nitrogen content, and relative nitrogen distribution. There were no significant source by fertilizer rate interactions for N concentration, content,

or relative distribution in leaf, shoot, root, and total plant tissues of *Malus trilobata* (Table 5). Fertilizer rate affected root and total plant N concentrations; N concentra-

tions were higher at 100 mg·L⁻¹ N than at 25 mg·L⁻¹ N for both sources (1.49 versus 1.00 and 1.35 versus 1.12 for MT2 and MT3, respectively; Table 5). Regardless of the tissue type in *Malus trilobata*, tissue N concentration was higher at the high fertilizer rate than the low fertilizer rate (Table 5). Also, leaf N concentration was higher than that of stem and root tissue in *Malus trilobata* at both fertilizer rates. Nitrogen content in leaf and stem tissues was higher at the high fertilizer rate than at the low fertilizer rate in *Malus trilobata*, but the opposite was true for *Acer syriacum* plants (Table 5). Relative N distribution in leaf, shoot, and root tissues was unaffected by seed source or fertilizer rate in *Malus trilobata* or by fertilizer rate for *Acer syriacum*. In *Malus trilobata*, shoot tissue had the greatest relative N content, whereas in *Acer syriacum*, it was higher in shoot tissue at the low fertilizer rate but higher in leaf tissue at the high fertilizer rate (Table 5).

Tissue phosphorus concentration, phosphorus content, and relative phosphorus distribution. Neither fertilizer rate nor source affected P concentration or content or its relative distribution in leaf, shoot, and root tissues in *Malus trilobata* (Table 6). In *Acer syriacum*, plants under the low fertilizer rate had the most P in shoot tissue (44%), whereas at the high fertilizer rate, root tissue contained the most P (53%). In most cases, root P concentration was higher than that of stem and root tissue at both fertilizer rates in *Malus trilobata* and *Acer syriacum* (Table 6).

Tissue potassium concentration, potassium content, and relative potassium distribution. Neither fertilizer rate nor source affected K concentration or content or its relative distribution in leaf, shoot, and root tissues of *Malus trilobata* and *Acer syriacum* (Table 7).

Nutrient uptake and nutrient use efficiencies. For both species, there was no significant source by fertilizer rate interaction for N, P, and K uptake between May and Sept. 2004 or for mineral nutrient use efficiency (Table 8). Fertilizer rate affected potassium use efficiency in *Malus trilobata*; it was higher at the low fertilizer rate than the high rate (41.4% versus 22.7%, respectively). There were no significant differences between the *Malus trilobata* seed sources for N, P, and K uptake and use efficiencies (Table 8). Fertilizer rates did not affect N, P, or K use efficiencies in *Acer syriacum* (Table 8).

Nitrogen-phosphorus-potassium ratios. At the high fertilizer rate, the ratio of N in whole plant tissue was greatest and P was always the least; but, at the low fertilizer rate, the relative K content was greatest and P was the least (Table 9). The ratio of N-P-K in whole plant tissues (relative to P) of MT2 plants was 5.7-1-6.8 at the low fertilizer rate and 7.7-1-5.5 at the high fertilizer rate (Table 9). In plants from MT3, the ratio of N to K was lower at the low fertilizer rate and nearly equal at the high fertilizer rate. In *Acer syriacum*, the ratio of N to K was lower at both fertilizer rates (Table 9).

Table 2. Height, caliper, root, shoot, leaf and total plant dry weights, shoot/root ratio, and leaf area of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Species	Fertilizer (mg·L ⁻¹ N)	Ht (cm)	Caliper (mm)	Dry weights(g)				Shoot/root ratio	Leaf area (cm ²)
				Root	Shoot	Leaf	Total plant		
MT2	25	167.4 ^z	12.9 ^y	61.6	125.8	21.4	208.9	2.04	1,316
	100	140.3	12.2	58.8	106.2	25.4	190.4	1.80	1,693
	Avg.	153.8	12.6	60.2	116.0	23.4	199.6	1.92	1,504
MT3	25	169.5	12.5	36.7	110.0	21.2	167.9	3.00	1,459
	100	121.5	11.5	33.2	72.1	18.5	123.8	2.17	916
	Avg.	145.5	12.0	34.9	91.0	19.8	145.8	2.58	1,187
AS2	25	76.1	11.4	37.7	50.5	32.6	120.7	1.34	2,843
	100	77.6	11.0	33.7	55.1	35.1	123.9	1.63	3,021
	Avg.	76.8	11.2	35.7	52.8	33.8	122.3	1.48	2,932
ANOVA				<i>P</i> > <i>F</i> value					
<i>Malus trilobata</i>									
Source (S)		NS ^x	NS	*	NS	NS	*	NS	NS
Fertilizer rate (FR)		**	NS	NS	NS	NS	NS	NS	NS
S*FR		NS	NS	NS	NS	NS	NS	NS	*
<i>Acer syriacum</i>									
Fertilizer rate (FR)		NS	NS	NS	NS	NS	NS	NS	NS

^zEach value is the mean of 10 individual seedlings within each species and fertilizer.

^yCaliper was measured at 1 inch (2.5 cm) above the root–shoot junction.

^xNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

Table 3. Distribution of root, shoot, and leaf dry weights of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Species	Fertilizer (mg·L ⁻¹ N)	Dry weight distribution (%)		
		Root	Shoot	Leaf
MT2	25	29.5 ^z	60.2	10.6
	100	30.9	55.8	13.3
	Avg.	30.2	58.0	11.9
MT3	25	21.9	65.5	12.6
	100	26.8	58.2	14.9
	Avg.	24.3	61.8	13.7
AS2	25	31.2	41.8	27.0
	100	27.2	44.5	28.3
	Avg.	29.2	43.1	27.6
ANOVA		<i>P</i> > <i>F</i> value		
<i>Malus trilobata</i>				
Source (S)		NS ^y	NS	NS
Fertilizer rate (FR)		NS	NS	*
S*FR		NS	NS	NS
<i>Acer syriacum</i>				
Fertilizer rate (FR)		NS	NS	NS

^zEach value is the mean of 10 individual seedlings within each species and fertilizer.

^yNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

Table 4. Leaf area ratio (LAR), net assimilation rate (NAR), and relative growth rate (RGR) of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Species	Fertilizer (mg·L ⁻¹ N)	LAR (g·cm ⁻²)	NAR (g·cm ⁻² ·d ⁻¹ × 10 ⁻³)	RGR (g·g ⁻¹ ·d ⁻¹ × 10 ⁻³)
MT2	25	7.2 ^z	3.7	25.7
	100	10.5	2.5	23.5
	Avg.	8.8	3.1	24.6
MT3	25	8.6	2.2	17.4
	100	9.7	1.4	12.5
	Avg.	9.1	1.8	14.9
AS2	25	25.7	0.8	20.3
	100	27.7	0.7	19.0
	Avg.	26.7	0.7	19.6
ANOVA		<i>P</i> > <i>F</i> – value		
<i>Malus trilobata</i>				
Source (S)		NS ^y	**	**
Fertilizer rate (FR)		*	**	**
S*FR		NS	NS	*
<i>Acer syriacum</i>				
Fertilizer rate (FR)		NS	NS	NS

^zEach value is the mean of 10 individual seedlings within each source and fertilizer.

^yNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

Discussion

The lower fertilizer rate, 25 mg·L⁻¹ N, resulted in greater growth than the high fertilizer rate, 100 mg·L⁻¹ N, in *Malus trilobata* seedlings. They were taller; had greater caliper, shoot, root, and leaf dry weights; and higher NAR at the lower fertilizer rate. However, the higher fertilizer rate increased LAR. Similar results were found for *Cercis siliquastrum* (Zahreddine et al., 2007a). In contrast, *Acer syriacum* growth was unaffected by fertilizer rate. Therefore, when using 21N–3.1P–5.9K water-soluble fertilizer, we recommend that both species be fertilized at 25 mg·L⁻¹ N rather than 100 mg·L⁻¹ N, which would result in a 75% reduction in fertilizer use without reducing growth. Additional testing is needed to determine the optimum fertilizer rate for these species in container-production systems. The 25 mg·L⁻¹ N fertilizer rate is lower than that typically recommended for other container-grown perennial woody species. *Malus trilobata* growth was vigorous at a lower rate than that reported by others for other genera (Gilliam et al., 1980, 1984; Ingstad, 1979; Jull et al., 1994; Larimer and Struve, 2002; Lumis et al., 2000; Musselwhite et al., 2004; Stubbs et al., 1997; Wright and Niemiera, 1987). For example, red maple growth was greatest at 200 to 400 mg·L⁻¹ N (Gilliam et al., 1980; Larimer and Struve, 2002). However, 20 mg·L⁻¹ N was optimal for *Cupressus arizonica* var. *glabra* ‘Carolina Sapphire’ (Arizona cypress ‘Carolina Sapphire’). In that study, higher fertilizer rates did not affect height and stem diameter, although N concentration in shoots and leaves increased with increased N rate (Stubbs et al., 1997). Height growth of *Malus trilobata* seedlings was similar to that of a container-grown crabapple (Stoven, 2004). Height growth of *Acer syriacum* was less than that of container-grown *Acer* ‘Autumn Flame’ (Larimer and Struve, 2002), not an

Table 5. Nitrogen concentration and content in leaf, shoot, root, and total plant tissues and nitrogen distribution in leaf, shoot, and root tissues as percent of total plant N content of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Source	Fertilizer rate (mg·L ⁻¹ N)	Nitrogen concn (%)				Nitrogen content (g)				Nitrogen distribution (%)		
		Leaf	Shoot	Root	Total plant	Leaf	Shoot	Root	Total plant	Leaf	Shoot	Root
MT 2	25	2.16 ^z	0.71	1.09	0.98	0.45	1.00	0.94	2.40	19	42	39
	100	2.32	1.13	1.49	1.41	0.51	1.06	0.75	2.32	22	46	32
MT 3	25	2.05	0.73	1.16	1.02	0.57	0.92	0.57	2.06	28	45	27
	100	2.27	0.90	1.49	1.26	0.80	1.10	0.63	2.53	32	43	25
AS 2	25	1.29	0.83	1.39	1.09	0.46	0.50	0.38	1.35	34	37	28
	100	1.68	0.83	1.30	1.22	0.38	0.26	0.35	1.00	38	27	35
ANOVA												
<i>Malus trilobata</i>												
Source (S)		NS ^y	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
Fertilizer rate (FR)		NS	NS	*	*	NS	NS	NS	NS	NS	NS	NS
S*FR		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
<i>Acer syriacum</i>												
Fertilizer rate (FR)		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS

^zEach value is the mean of three plants for each source and fertilizer rate.

^yNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

Table 6. Phosphorus concentration and content in leaf, shoot, root, and total plant tissues and P distribution in leaf, shoot, and root tissues as percent of total plant P content of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Source	Fertilizer rate (mg·L ⁻¹ N)	Phosphorus concn (µg·g ⁻¹)				Phosphorus content (g)				Phosphorus distribution (%)		
		Leaf	Shoot	Root	Total plant	Leaf	Shoot	Root	Total plant	Leaf	Shoot	Root
MT 2	25	2,056 ^z	964	2,341	1,634	0.04	0.13	0.25	0.42	9	31	60
	100	2,169	1,400	2,383	1,822	0.05	0.13	0.13	0.30	17	42	42
MT 3	25	1,894	95	1,954	1,354	0.05	0.13	0.10	0.28	18	46	36
	100	2,175	1,178	2,511	1,639	0.07	0.14	0.11	0.32	22	44	34
AS 2	25	1,408	1,922	3,375	2,102	0.05	0.12	0.10	0.26	19	44	37
	100	1,829	2,002	4,403	2,740	0.04	0.06	0.12	0.22	18	29	53
ANOVA												
<i>Malus trilobata</i>												
Source (S)		NS ^y	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
Fertilizer rate (FR)		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
S*FR		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
<i>Acer syriacum</i>												
Fertilizer rate (FR)		NS	NS	NS	NS	NS	NS	NS	NS	NS	*	*

^zEach value is the mean of three plants for each source and fertilizer rate.

^yNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

Table 7. Potassium concentration and content in leaf, shoot, root, and total plant tissues and K distribution in leaf, shoot, and root tissues as percent of total plant K content of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Source	Fertilizer rate (mg·L ⁻¹ N)	Potassium concn (µg·g ⁻¹)				Potassium content (g)				Potassium distribution (%)		
		Leaf	Shoot	Root	Total plant	Leaf	Shoot	Root	Total plant	Leaf	Shoot	Root
MT 2	25	17,064 ^z	11,883	9,698	12,143	0.35	1.66	0.84	2.85	12	58	30
	100	17,130	8,835	10,242	10,561	0.37	0.72	0.55	1.64	23	44	33
MT 3	25	15,984	1,225	9,425	12,072	0.45	1.56	0.47	2.48	18	3	19
	100	17,425	11,736	1,537	12,722	0.57	1.37	0.52	2.47	23	56	21
AS 2	25	10,422	13,746	15,298	13,367	0.37	0.84	0.46	1.67	22	50	27
	100	10,043	13,798	17,620	13,920	0.22	0.42	0.47	1.11	20	38	42
ANOVA												
<i>Malus trilobata</i>												
Source (S)		NS ^y	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
Fertilizer rate (FR)		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
S*FR		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
<i>Acer syriacum</i>												
Fertilizer rate (NFR)		NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS

^zEach value is the mean of three plants for each source and fertilizer rate.

^yNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

unexpected finding because *Acer syriacum*'s mature height is less than 'Autumn Flame'.

There were only two source by fertilizer rate interactions for *Malus trilobata* seedlings, leaf area and RGR. Fertilizer rate changed the rank order for leaf area of the two *Malus* sources; at the high rate, leaf area increased in MT2 but decreased in MT3. Both *Malus* sources had lower RGR at the higher fertilizer rate, but the decrease was

greater for MT3 than MT2. There was also a source by fertilizer rate interaction for RGR among six *Cercis siliquastrum*. For *Cercis*, four sources had lower RGR at the higher fertilizer rate, one was unchanged, and one had higher RGR (Zahreddine et al., 2007a).

The relative distribution of *Malus trilobata*'s total plant dry weight was altered by fertilizer rate; at the higher fertilizer rate, relatively more dry weight was in leaves and

roots and less in shoots, but shoot tissue still represented the greatest relative percentage of total plant dry weight. In contrast, relative shoot dry weight of *Cercis siliquastrum* (Zahreddine et al., 2007a) and 'Autumn Blaze' red maple (Larimer and Struve, 2002) plants increased with higher N addition rates. *Acer syriacum* growth was unaffected by fertilizer rate; it also had the highest relative percentage of total plant dry weight

Table 8. Nitrogen, P, and K uptake and mineral nutrient use efficiency of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Source	Fertilizer rate (mg·L ⁻¹ N)	Mineral nutrient uptake (g)			Mineral nutrient use efficiency (%)		
		N	P	K	N	P	K
MT 2	25	2.24 ^z	0.41	2.42	10	2	45
	100	2.17	0.29	1.21	7	1	18
MT 3	25	1.78	0.25	2.20	8	1	38
	100	2.26	0.29	2.19	7	1	27
AS 2	25	1.13	0.24	1.40	5	1	24
	100	0.77	0.20	0.84	2	1	11
ANOVA							
<i>Malus trilobata</i>							
Source (S)		NS ^y	NS	NS	NS	NS	NS
Fertilizer rate (FR)		NS	NS	NS	NS	NS	**
S*FR		NS	NS	NS	NS	NS	NS
<i>Acer syriacum</i>							
Fertilizer rate (FR)		NS	NS	NS	NS	NS	NS

^zEach value is the mean of three plants for each source and fertilizer rate.

^yNS, *, and ** indicate no statistical difference at $\alpha = 0.05$ level of significance or statistical difference at $\alpha \leq 0.05$ or $\alpha \leq 0.01$, respectively.

Table 9. Whole plant N–P–K ratios (expressed relative to P) of seedlings raised from two *Malus trilobata* trees and one *Acer syriacum* tree when grown under either 25 or 100 mg·L⁻¹ N water-soluble fertilizer.

Source	Fertilizer rate (mg·L ⁻¹ N)	Ratio		
		N	P	K
MT 2	25	5.7 ^z	1	6.8
	100	7.7	1	5.5
MT 3	25	7.3	1	8.8
	100	7.9	1	7.7
AS 2	25	5.2	1	6.1
	100	4.5	1	5.0

^zEach value is the mean of three plants per source and fertilizer treatment.

in the shoot system. The relative percentage of *Malus* shoot dry weight was high compared with North American *Quercus* (oak) species, *Malus* ‘Prairifire’ (Prairifire crabapple), *Acer rubrum* ‘Autumn Blaze’ (Autumn Blaze red maple), and *Nyssa sylvatica* (Black gum) plants grown under similar conditions (Drunasky and Struve, 2005; Larimer and Struve, 2002; Stoven, 2004; Struve, 1995). The relative percentage of *Acer syriacum* shoot dry weight was similar to *Cercis canadensis* and *C. siliquastrum* (judas tree), *Acer rubrum* ‘Autumn Blaze’, *Malus* ‘Prairifire’, and *Quercus prinus* (chestnut oak) (Drunasky and Struve, 2005; Larimer and Struve, 2002; Stoven, 2004; Zahreddine et al., 2007a). The significance of the relatively high *Malus trilobata* stem dry weight is unknown.

Nitrogen loading occurred in the *Malus trilobata* plants grown at the high fertilizer rate, but not in *Acer syriacum* seedlings (Table 5). Nutrient loading is defined as an increase in plant tissue nutrient concentration without a significant increase in plant dry weight (Malik and Timmer, 1995). Nutrient loading was observed in *Cercis siliquastrum* sources grown at 100 mg·L⁻¹ N in a study by Zahreddine et al. (2007a). Studies by Xu and Timmer (1999) and by Malik and Timmer (1995) showed that nutrient-loaded conifer seedlings performed better when outplanted

in nutrient-poor sites than those not nutrient-loaded. In addition, nutrient loading increases the competitive ability for resources of the crop species in weed-prone or heavily vegetated sites (Malik and Timmer, 1995), which could be useful for conservation purposes (reforestation or reintroduction of species to their native habitats). The possible benefits of nutrient loading nursery stock during production in *Malus trilobata* plants require experimental confirmation. If these benefits are proven, nursery managers could offer higher-quality plants, assuming the benefits of nutrient loading are not offset by decreased plant quality (i.e., smaller caliper, smaller total plant dry weight, and higher shoot to root ratio).

Similar to *Cercis siliquastrum* (Zahreddine et al., 2007a), neither mineral nutrient uptake nor mineral nutrient uptake efficiency between May and September were affected by fertilizer rate in *Malus trilobata* and *Acer syriacum* seedlings (Table 8). Nutrient contents of *Malus trilobata* seedlings were similar under the two fertilizer rates because although seedlings had higher dry weights at the lower fertilizer rate, nutrient concentrations were lower than for those grown at the high rate.

Although comparisons are limited, nitrogen use efficiency of both species was lower at the higher fertilizer rate in this study and others (Table 8; Larimer and Struve, 2002; Struve, 1995; Zahreddine et al., 2007a). Nitrogen use efficiency ranged between 7% and 11% in *Malus* and 3% and 5% in *Acer* compared with a range of 7% to 15% in *Cercis siliquastrum* (Zahreddine et al., 2007a). Also similar to *Cercis siliquastrum*, P use efficiency in *Malus trilobata* and *Acer syriacum* was lower than N and K use efficiencies. Phosphorus use efficiency at both fertilizer rates ranged between 1% and 2% in *Malus trilobata* and 1% in *Acer syriacum*; in *Cercis siliquastrum*, phosphorus use efficiency was 2% to 4%.

Similarly, potassium use efficiency in *Malus trilobata* was lower at the high fertilizer rate than at the low fertilizer rate and was

higher than nitrogen use efficiency or phosphorus use efficiency. It should be noted that at the low fertilizer rate, the compost contributed 85%, 88%, and 94%; and at the high rate, it contributed 58%, 95%, and 76% of the total N, P, and K applied to the plants, which contributed to the relatively low nutrient use efficiency. Plants under the low fertigation regime (25 mg·L⁻¹ N) received 3.3, 0.5, and 0.8 g N, P, and K, respectively, between 12 May and 24 Sept.; those under the high fertigation regime (100 mg·L⁻¹ N) received 13.3, 2.0, and 3.2 g N, P, and K, respectively. Comtil alone supplied 18.0, 39.8, and 9.9 g N, P, and K, respectively, during the growing season. An explanation for the high K efficiency could be genetic ability of *Malus trilobata* to store K for to be used in osmotic adjustment during drought. Deficiency in K has been associated with reduced ability to withstand stress and maintain high transpiration rates (Tisdale et al., 1998). Another possible explanation for why potassium use efficiency was higher than nitrogen use efficiency and phosphorus use efficiency maybe that K promotes the conversion of inorganic P to nucleic acids and phosphoproteins that would otherwise cause plant toxicity. Toxicity effects resulting from elevated inorganic phosphates has been reported in *Ilex crenata* when P was applied at concentrations higher than 10 ppm (Wright and Niemiera, 1987).

The fertilizer used in this study had an N–P–K ratio of 6.8–1–1.6 when N and K were expressed relative to P. However, whole plant mineral nutrient ratios at the end of the growing season did not match the fertilizer N–P–K ratios. Plant tissue contained more N than P or K under the high fertilizer rate, but K content was highest when plants were grown at the low fertilizer rate. This was true for both species. Nutrient use efficiency can be increased by matching the fertilizer nutrient ratio to plant N–P–K tissue ratios.

There were differences in growth between the two *Malus trilobata* sources as there was among six *Cercis siliquastrum* sources (Zahreddine et al., 2007a). *Malus trilobata*, like other apple species, is reported to be crosspollinated by insects (http://www.ibiblio.org/pfaf/cgi-bin/arr_html?Malus+trilobata&CAN=LATIND). Although both mother trees of *Malus trilobata* came from the same stand, there were significant differences in growth between the half-sib families. However, there were no differences among the *Malus trilobata* sources in mineral nutrient uptake efficiency or mineral nutrient partitioning, but there were significant differences among the six *Cercis siliquastrum* sources in N, P, and K concentrations and contents (Zahreddine et al., 2007a). Interestingly, whole plant tissue from MT3 contained more N and K than MT2 tissue at both fertilizer rates. The N–P–K ratio of MT3 was 7.6–1–8.25 compared with that of MT2, which was equal to 6.7–1–6.15.

As was previously mentioned, *Malus trilobata* is endemic to Lebanon and is endangered, whereas *Acer syriacum*’s wood is often used by local people for firewood and

cabinet making. These species have an economic potential as a result of their ornamental attributes. One way to conserve them is by encouraging the establishment of ex situ facilities and in situ conservation efforts. Establishing botanic gardens, seed banks in Lebanon, or conserving entire ecosystems by turning them into "protected areas" will require an extensive amount of time, research effort, and huge capital. A more practical and feasible alternative will be to encourage the cultivation of the species. Cultivation practices will satisfy both commercial and conservation purposes. Nursery managers can introduce both species into the Lebanese landscape market by propagating seedlings and selling them to local municipalities for street plantings and to people who want to plant those trees in their home gardens. Encouraging the cultivation of native trees will also reduce the dependence of the local market on introduced landscape species; it will also reduce the potential of introducing non-native pests and pathogens that negatively affect the environment. Moreover, reintroducing the trees into their native habitats and other reforestation projects will take place once those trees are made available in the market.

Conclusions

Acer syriacum and *Malus trilobata* seedlings are amenable to container production. Seedlings of both species can be grown at 25 mg·L⁻¹ N using 21N-3.1P-5.9K water-soluble fertilizer. Additional fertilizer rates need to be tested to determine the optimum fertilizer rate. Nutrient uptake was unaffected by fertilizer rate in *Malus trilobata* and *Acer syriacum*, but N concentration in *Malus trilobata* was. Nitrogen loading occurred in *Malus trilobata* plants grown under the high fertilizer rate, but any possible benefits need to be confirmed by field trials to determine whether the nutrient-loading benefits out-

weigh the decrease in plant quality (smaller plants and lower dry weights). There were significant differences in growth between the two *Malus trilobata* half-sib families although both trees were growing in the same stand. We recommend that seeds be collected from additional tree sources for both species and tested for growth and nutrient partitioning under container production to identify genetically superior seed trees of these threatened species. Container nursery production and transplanting to managed landscapes might be a practical ex situ genetic conservation method.

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