Response of Creeping Bentgrass to Salinity and Mowing Management: Carbohydrate Availability and Ion Accumulation

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Abstract. Salt problems in turfgrass sites are becoming more common. The effects of mowing height on salt tolerance and associated mechanisms are not well understood. The objective of this study was to examine the effects of mowing height and the level of salinity on turf quality, canopy photosynthetic rate (Pn), total nonstructure carbohydrate (TNC) content, shoot reducing sugar content (RSC), Na+ and K+ content in shoots and roots of creeping bentgrass (Agrostis palustris Huds.). Sod pieces of ‘L-93’ were grown in a greenhouse for over 7 months. Plants were subjected to three mowing heights: 6.4, 12.7, and 25.4 mm, and to four salinity levels of irrigation water: control, 5 dS·m–1, 10 dS·m–1, and 15 dS·m–1 prepared using ocean salts. Increasing salinity resulted in reduced turf quality, increased shoot Na+, reduced K+, and reduced K to Na ratio, to a greater extent for bentgrass mowed at 6.4 mm mowing height. Reducing sugar content in shoot increased with increasing salinity level except at 15 dS·m–1 and 6.4 mm mowing regime where RSC declined. Compared to the 25.4 mm mowing height, mowing height at 6.4 mm caused 32–39% reduction in TNC, a 25% to 37% increase in Na+ content, and 45% to 51% decrease in K content in shoots, which resulted in substantial decrease in K/Na ratio. These results demonstrated that the reduction of creeping bentgrass salt tolerance under low mowing height was associated with carbohydrate depletion that reduced the plant’s genetic abilities to generate osmo-protectants (such as reducing sugar), to reduce Na+ accumulation in shoots, and to selectively uptake and transport K+. Therefore, a moderate increase in mowing height could improve salt tolerance of creeping bentgrass.

Creeping bentgrass, because of its high turf quality, is one of the most widely used species on golf course putting greens, tees, and fairways in the temperate regions of the world. Golf course managers have been challenged to reduce mowing height to increase ball rolling distance and the quality of play surfaces. Creeping bentgrass is considered to have moderate salinity tolerance (Beard, 1973; Carrow and Duncan, 1998; Harivandi et al., 1992) and variations in salinity tolerance exist among different cultivars (Marcum, 2001). Previously it has been reported that the salinity damage of creeping bentgrass becomes more severe under close mowing conditions (Fu et al., 2005).

Salinity tolerance mechanisms of plants include salt exclusion, regulation of salt transport to shoots, salt secretion, synthesis of benign osmoprotectants, organismal and cellular salt compartmentations, and damage response and repair (Tester and Davenport, 2003). Many of the salt resistance mechanisms are energy dependent (Barrett-Lennard, 2003; Tester and Davenport, 2003). For example, sodium exclusion is in part associated with the ATPase activity in root cell membranes. For cellular salt compartmentation, the function of vascular Na+/H antiport relies on ATPase to generate cytosol-directed electrochemical potential difference for H+ across membrane. The syntheses of benign osmoprotectants rely on carbohydrate substances and energy (Rhodes et al., 2002). Soluble reducing sugars are thought to serve an important role in interacting with membrane phospholipids and proteins to stabilize their structures under salinity stress. Gram (1976) suggested that, of the various organic osmotica, sugars contribute up to 50% of the total osmotic potential in glycohytes subject to saline conditions.

A positive balance of carbohydrate production and consumption would help plants maximize their ability to cope with salinity stress. Close mowing reduces canopy photosynthesis by removing large amounts of green leaves that otherwise are available for light interception (Hull, 1992), whereas nonphotosynthesizing tissues continue to respire and increase dark respiration (Krans and Beard, 1975), thereby reducing energy reserves and energy availability. Canopy photosynthesis and TNC in plants are physiological measures of energy production and energy availability under stressful conditions (Huang and Fu, 2000; Qian and Engelke, 1999; Qian et al., 1998).

The hypothesis of the current study is that the decline in bentgrass’s ability to tolerate salinity under close mowing conditions is associated with the carbohydrate availability, the effectiveness of Na+ exclusion from shoots, and the ability to selectively uptake and transport K+. To test the hypothesis, the experiment was conducted to determine turf quality, canopy photosynthetic rate, total non-structure carbohydrate content (TNC), shoot reducing sugar content (RSC), Na+ and K+ content in shoots and roots under several combinations of mowing regimes and salinity levels.

Materials and Methods

Plant materials and growth conditions. Sod pieces (10 cm in diameter) of ‘L-93’ were collected from a U.S. Golf Association putting green seeded 4 years ago at the Horticulture Research Center, Fort Collins, Colo. After removing soil through hand washing, sod pieces were transplanted to polyvinyl chloride (PVC) tubes (45 cm long and 10 cm in diameter) filled with sandy clay loam soil (Aridic Argustoll) and kept in a greenhouse. A 10-cm-inside diameter PVC cap with five 1-cm-diameter holes drilled at the bottom was positioned at the bottom of the PVC tube for drainage. The soil pH and EC were 7.4 and 1.2 dS·m–1, respectively. Soil columns were packed to a bulk density of about 1.3 g·cm–3.

Plants were grown in the PVC tubes for 3 months, allowing root and canopy to establish before treatments were imposed. During this period, plants were watered on alternate days using tap water (EC = 0.2 dS·m–1) until water drained freely from the holes at the bottom of each tube and fertilized weekly with Peters Excel solution (15N–2P–12K, Scotts-Sierra Horticultural Products Company Marysville, Ohio) to provide 24 kg N/ha per month. Turfgrass was hand clipped every 2 d at 6.4 mm. The daytime greenhouse temperature ranged from 19.5 to 28.5 °C and nighttime temperature ranged from 13.5 to 20.5 °C. Ambient photosynthetically active radiation just above the turf canopy at 10:00 h averaged about 1000 μmol·m–2·s–1.

Treatments. Salinity treatments were applied by irrigating with different concentrations of saline water. Irrigation waters of different salinity levels were prepared by the addition of ocean salts (Aquarium Syst., Mentor, Ohio) to tap water to obtain EC values of 5.0, 10.0, and 15.0 dS·m–1. The tap water without addition of sea salts is used as the control (EC = 0.2 dS·m–1). Concurrent to the initiation of salinity treatments, mowing treatments (that included clipping three times weekly at 25.4 mm, four times at 12.7 mm, and daily at 6.4 mm) started. The irrigation waters were applied every 2 d with the amount equal to 120% of evapotranspiration (ET) measured with an extra PVC container (served as a lysimeter) of bentgrass mowed at 25.4 mm. The amount of water applied was the same for all salinity and mowing treatments. Salinity and mowing treatments continued for 4 months.

Measurements. Data were collected for turf quality, Pn, soil salinity, shoot TNC, shoot RSC, and tissue Na+ and K+ content for each treatment.

Visual turf quality was rated biweekly based on color, density, and uniformity using a scale of 0 (brown, dead turf) to 9 (optimum color, dense, and uniform turf), with a rating of 6.0 or higher indicating acceptable quality.

Canopy net photosynthetic rate was measured biweekly between 10:00 and 15:00 HR.
with a portable CO2 gas analyzer system (model CI-301; QC CD Inc., Vancouver, Wash.). During each measurement, the whole turf canopy of individual PVC columns was enclosed in a transparent plexiglass chamber (7.6 cm in height and 10 cm in diameter) for 2 min. The canopy chamber was attached to the CO2 gas analyzer system to determine CO2 uptake. Canopy net photosynthetic rate was expressed as CO2 uptake per unit canopy area.

Soil salinity, TNC, RSC, and tissue Na+ and K+ content were determined at the termination of the experiment. To determine soil salinity, the PVC tubes containing soil were cut into four 10-cm-deep sections. One fourth of the soil within each section was air-dried for the measurement of soil salinity (and the rest of the soil was used to collect roots). The air-dried soil was well mixed, crushed and passed through a 2-mm soil sieve. Deionized water was added to ground and sieved soil and mixed uniformly until a saturated paste was obtained. Electrical conductivity of saturation extract was determined with soil salinity appraisal kit (model CO150; Hach, Loveland, Colo.). Detailed information on salinity and root mass at different depths was presented in Fu et al. (2005). Data on average soil salinity over four depths was used in the current paper.

At the termination of the experiment, aboveground tissues (including shoots and stolons) were harvested and washed with cold water to remove plant debris for carbohydrate analysis. The samples were freeze-dried (Genesis 25 L; Lyophilizer; Virtis, Gardiner, N.Y.). After freeze-drying, samples were ground in a Wiley mill, sieved through a screen with 80-mµm openings, and kept in airtight vials at –20 °C. Total nonstructural carbohydrate content was measured using the method described by Chatterton et al. (1987). Briefly, 25 mg freeze-dried samples were transferred to 5 mL 0.1% clarase solution and incubated at 38 °C for 24 h. Then 0.5 mL of hydrochloric acid (50%, by volume) was added to the incubation solution. After the solution was incubated at room temperature for 18 h, the pH value of the solution was adjusted to between 5 and 7 with 10 and 1 N NaOH. This solution was used to determine TNC content (the reducing sugar in the solution was considered as TNC content) using a spectrophotometer at 515 nm wavelength (model DU640; Beckman).

To measure free reducing sugar, 25 mg of the freeze-dried, ground, and sieved sample was extracted with 10 mL 0.1 M phosphate buffer (pH = 5.4) for 24 h at room temperature. Aliquot of extraction (0.2 mL) was used to determine the reducing sugar content by using method of measuring TNC.

To measure ion content, shoots and roots were harvested, washed with deionized water, and dried at 70 °C for 24 h. Dried shoots and roots were ground in a Wiley mill to pass through a screen with 425-µm openings. About 1 g of screened and dried sample was weighed and ashed for 7 h at 500°C. Ash was dissolved in 10 mL of 1 N HCl and diluted with deionized water. Solution aliquots were analyzed for Na+ and K+ by inductively coupled plasma atomic emission spectrophotometry (ICP–AES) (model 975 Plasma Atomcomp; Thermo Jarrell Ash Corp., Franklin, Mass.).

Statistical analysis. As split-plot experimental design was used with mowing regimes being the main plot treatments, and salinity levels of irrigation water being the subplot treatments. Each treatment had four replications. Effects of mowing, salinity, and their interaction were determined using analysis of variance according to the general linear procedure of the Statistical Analysis System (SAS, 2001) (Table 1). Because of a significant mowing regime and salinity level interaction on most of the measured variables, comparisons among mowing regimes within each salinity level were presented and means were separated by least significant difference at the 0.05 level of probability. Linear regression analysis was performed to determine the relationship between the measured parameters and soil salinity, where salinity level was the independent variable and each measured parameter was the dependent variable. Since turf quality and Pn measurements were taken on the same experimental units biweekly from week 10 to week 12, a repeated measures analysis was performed to determine effects of mowing and salinity on turf quality and Pn (SAS, 2001).

Results and Discussion

Soil salinity and turf quality. A linear relationship of increasing soil salinity with increasing irrigation water salinity was observed for each mowing regime (R2 = 0.92 to 0.99) (Table 2). Since all treatments received the same amount of water, the 6.4 mm mowing regime exhibited lower soil salinity than the 25.4 mm mowing regime under all salinity treatments. At 5 and 10 dS·m–1 salinity treatments, the 12.7 mm mowing regime exhibited lower soil salinity than the 25.4 mm mowing regime. Likely, the 6.4 mm mowing regime exhibited lower soil salinity than the 12.7 mm mowing regime. The trend of lower soil salinity with reduced mowing height was because a lower mowing height resulted in a higher leaching fraction; as we observed that greater volume of water was drained from the low mowing regimes. The higher leaching fractions from the low mowing treatments likely have occurred because turf mowed lower possessed lower ET rates.

Increasing soil salinity reduced turf quality, more severely and rapidly when mowed at 6.4 mm than at 12.7 or 25.4 mm (Fig. 1). Regressions analysis of soil salinity and turf quality suggested that turf quality of creeping bentgrass mowed to 6.4, 12.7, and 25.4 mm fell to an unacceptable level when soil salinity reached 4.1, 12.5, and 13.9 dS·m–1, respectively.

Canopy photosynthetic rate (Pn). The interaction of salinity and mowing treatments on Pn was not significant (Table 1). Increasing salinity and reducing mowing height additively decreased canopy photosynthesis of creeping bentgrass (Table 3). As irrigation water salinity increased from control to 15 dS·m–1, Pn reduced an average of 40%. Although Pn is less sensitive to salinity compared to shoot growth (Fu et al., 2005), our results indicated that the reduction in Pn by close mowing (mowed to 6.4 mm) and high salinity stress could lead to limited carbohydrate availability. The adverse effects of salinity on photosynthesis have been determined in studies in other species and creeping bentgrass (Balibrea et al., 2000; Chartzoulaki et al., 2002; Liu and Cooper, 2002). Using in a hydroponic system, Liu and Cooper (2002) reported that increasing salinity to 16 dS·m–1 decreased net photosynthesis of creeping bentgrass by 20%. In tomato plant,

Table 1. Analysis of variance of turf quality, total non-structure carbohydrate content (TNC), shoot reducing sugar content (RSC), shoot Na+ and K+ content, canopy photosynthetic rate (Pn), and soil salinity with mean squares and treatment significance levels.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Mowing ht</th>
<th>Block</th>
<th>Mowing ht × block</th>
<th>Salinity level</th>
<th>Mowing regime × salinity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Turf quality</td>
<td>7.3**</td>
<td>0.1</td>
<td>0.7</td>
<td>67.8***</td>
<td>3.4*</td>
</tr>
<tr>
<td>TNC</td>
<td>9243***</td>
<td>301*</td>
<td>283*</td>
<td>815***</td>
<td>392**</td>
</tr>
<tr>
<td>RSC</td>
<td>2922***</td>
<td>15.8*</td>
<td>23.2*</td>
<td>108**</td>
<td>77</td>
</tr>
<tr>
<td>Root Na+ content (mg kg–1)</td>
<td>2.5 × 104</td>
<td>25 × 10–4</td>
<td>6.2 × 10–4</td>
<td>58 × 10–6</td>
<td>8.5 × 10–6</td>
</tr>
<tr>
<td>Root K+ content (mg kg–1)</td>
<td>7.0 × 104</td>
<td>8.9 × 10–4</td>
<td>1.5 × 10–4</td>
<td>46 × 10–6</td>
<td>9.4 × 10–6</td>
</tr>
<tr>
<td>Shoot Na+ content (mg kg–1)</td>
<td>56.7 × 10–4</td>
<td>4.3 × 10–4</td>
<td>6.6 × 10–4</td>
<td>403.4 × 10–6</td>
<td>11.3</td>
</tr>
<tr>
<td>Shoot K+ content (mg kg–1)</td>
<td>142.8 × 10–4</td>
<td>9.8 × 10–3</td>
<td>1.73 × 10–1</td>
<td>174 × 10–3</td>
<td>1.35</td>
</tr>
<tr>
<td>Pn</td>
<td>19.7*</td>
<td>1.0</td>
<td>2.2*</td>
<td>78.4***</td>
<td>0.5</td>
</tr>
<tr>
<td>Soil salinity</td>
<td>23.0*</td>
<td>0.02</td>
<td>1.04</td>
<td>415.7***</td>
<td>2.16*</td>
</tr>
</tbody>
</table>

**Significant level at P ≤ 0.05, 0.01, 0.001, or <0.0001, respectively.
salinity increased, the mowing TNC by 20% in comparison with turf mowed treatment, mowing height at 6.4 mm reduced caused by close mowing. Without salinity high salinity escalated the TNC depletion that removes photosynthetic tissues. However, reduced mowing height and increased mowing each salinity treatment, total nonstructural carbohydrates may interact with membrane iccation (Popp and Smirnoff, 1995). Soluble reducing sugars are thought to serve an important role in salinity, drought, and low temperature tolerance via osmotic adjustment and act as protectants as they prevent cell desiccation (Popp and Smirnoff, 1995). Soluble carbohydrates may interact with membrane phospholipids and proteins to stabilize their structures under salinity stress. Previous studies have demonstrated that salinity increases glucose and fructose content in olive (Olea europaea L.) (Gucci et al., 1998; Tattini, et al., 1996), wheat seedlings (Triticum aestivum L.) (Kerepesi and Galiba, 2000; Hu et al., 2000), and tomato (Lycopersicon esculentum L.) (Balibrea et al., 1996, 2000; Gao et al., 1998). Several studies have attempted to relate the magnitude of changes in soluble carbohydrate to salinity tolerance. Ashraf (1994) found that salt-tolerant wide populations of Melilotus indica and Erura sativa had significantly higher soluble sugars in their leaves than the salt sensitive populations at varying salt levels of the growth medium.

The reducing sugar in grass species mainly consists of glucose and fructose (Ball et al., 2002; Shahba et al., 2003). When mowed at 25.4 and 12.7 mm, increasing salinity increased reducing sugar content in shoots of creeping bentgrass, despite a significant decrease in Pn (Table 4). Under 6.4 mm mowing height, however, increasing salinity to 15 dS·m⁻¹ reduced RSC at P < 0.1.

Nonstructural carbohydrates are reserved energy sources in turfgrass plants. Soluble reducing sugars are thought to serve an important role in salinity, drought, and low temperature tolerance via osmotic adjustment and act as protectants as they prevent cell desiccation (Popp and Smirnoff, 1995). Soluble carbohydrates may interact with membrane phospholipids and proteins to stabilize their structures under salinity stress. Previous studies have demonstrated that salinity increases glucose and fructose content in olive (Olea europaea L.) (Gucci et al., 1998; Tattini, et al., 1996), wheat seedlings (Triticum aestivum L.) (Kerepesi and Galiba, 2000; Hu et al., 2000), and tomato (Lycopersicon esculentum L.) (Balibrea et al., 1996, 2000; Gao et al., 1998). Several studies have attempted to relate the magnitude of changes in soluble carbohydrate to salinity tolerance. Ashraf (1994) found that salt-tolerant wide populations of Melilotus indica and Erura sativa had significantly higher soluble sugars in their leaves than the salt sensitive populations at varying salt levels of the growth medium.

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changes in Na⁺ and K⁺ resulted in a decrease in the K⁺ to Na⁺ ratio from 22 to <1. Wyn Jones et al. (1979) suggested a threshold K⁺ to Na⁺ ratio of 1 for normal growth of plants subjected to salinity. Shannon (1978) studied salt tolerance of 32 lines of tall wheatgrass and found that salinity tolerance was associated with maintenance of K⁺ to Na⁺ ratio >1. Regression analysis predicted that shoot K⁺ to Na⁺ ratio decreased to 1 at 6.0, 7.1, and 9.5 dS·m⁻¹ when bentgrass was mowed to 6.4, 12.7, and 25.4 mm, respectively.

Our data on Na and K content indicated that bentgrass had the highest selectivity of K⁺ over Na⁺ when maintained at 25.4 mm mowing height. Close mowing reduced the root selectivity for K under high salinity conditions and increased the uptake and transport of Na to shoots. Membrane proteins play a significant role in the selective distribution of ions within the plant and cell. These proteins include 1) primary H⁺-ATPases which generate the H⁺ electrochemical gradient that drives ion transport through the plasma membrane with high selectivity for K over Na, 2) Na/H antiports in the plasma membrane for pumping excess Na out of the cell, 3) Na/H antiports in the tonoplast for extruding Na into the vacuole (Ashraf and Harris, 2004; Tester and Davenport, 2003). All these are energy dependent processes. The selectivity of K over Na in the root uptake and transport are likely regulated to a substantial degree by H⁺ gradients across the plasma membrane. These H⁺ gradients are maintained by H⁺-ATPase activity. Low mowing height depletes TNC reserves that induce ATP deficit, resulting in the lack of energy to power active ion selectivity in uptake and transport.

Our results on Pn, TNC, RSC, and Na and K content support our hypothesis, i.e., salinity tolerance of creeping bentgrass is appreciably

<table>
<thead>
<tr>
<th>Carbohydrate type</th>
<th>Mowing ht (mm)</th>
<th>Salinity level (dS·cm⁻¹)</th>
<th>Regression (TNC or RSC vs. soil salinity)</th>
<th>( R^2 )</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNC (mg g⁻¹ dry wt)</td>
<td>6.4</td>
<td>756.5 a</td>
<td>695.6 a</td>
<td>688.5 b</td>
</tr>
<tr>
<td></td>
<td>12.7</td>
<td>549.3 a</td>
<td>715.3 a</td>
<td>983.9 a</td>
</tr>
<tr>
<td></td>
<td>25.4</td>
<td>485.6 a</td>
<td>646.0 a</td>
<td>1052.7 a</td>
</tr>
<tr>
<td>K (mg g⁻¹ dry wt)</td>
<td>6.4</td>
<td>885.10 a c</td>
<td>226.23 b</td>
<td>190.83 b</td>
</tr>
<tr>
<td></td>
<td>12.7</td>
<td>614.07 a b</td>
<td>443.45 a</td>
<td>529.06 a</td>
</tr>
<tr>
<td></td>
<td>25.4</td>
<td>625.54 a b</td>
<td>51.9 a</td>
<td>55.5 a</td>
</tr>
</tbody>
</table>

Table 4. Total nonstructural carbohydrates (TNC) and reducing sugar content (RSC) in shoots of creeping bentgrass subjected to three mowing heights and at four levels of irrigation salinity.

<table>
<thead>
<tr>
<th>Ion</th>
<th>Mowing ht (mm)</th>
<th>Salinity level (dS·cm⁻¹)</th>
<th>Regression (Na or K vs. soil salinity)</th>
<th>( R^2 )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Na (mg g⁻¹ dry wt)</td>
<td>6.4</td>
<td>123.8 b</td>
<td>102.7 b</td>
<td>68.6 b</td>
</tr>
<tr>
<td></td>
<td>12.7</td>
<td>142.5 ab</td>
<td>145.5 a</td>
<td>137.3 a</td>
</tr>
<tr>
<td></td>
<td>25.4</td>
<td>157.2 a</td>
<td>150.5 a</td>
<td>152.2 a</td>
</tr>
<tr>
<td>K (mg g⁻¹ dry wt)</td>
<td>6.4</td>
<td>34.7 b</td>
<td>35.1 b</td>
<td>34.0 b</td>
</tr>
<tr>
<td></td>
<td>12.7</td>
<td>41.5 a</td>
<td>48.8 a</td>
<td>52.2 a</td>
</tr>
<tr>
<td></td>
<td>25.4</td>
<td>44.8 a b</td>
<td>51.9 a</td>
<td>55.5 a b</td>
</tr>
</tbody>
</table>

Table 5. Sodium and potassium content in roots of ‘L-93’ creeping bentgrass subjected to three mowing heights under different levels of salinity.

Values followed by the same letters within a column are not significantly different (\( P = 0.05 \)) based on a LSD test. **Significant at \( P \leq 0.1, 0.05, \text{ or } 0.01.\)
lowered under close mowing conditions and this reduction is associated with reduced carbohydrate availability that leads to reduced ability to effectively produce osmo-protectants (such as glucose and fructose), minimize Na accumulation in shoots, and selectively uptake and transport K.

**Literature Cited**