

# Chlorophyll Fluorescence as a Potential Indicator of Controlled-atmosphere Disorders in 'Marshall' McIntosh Apples

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**Abstract.** Chlorophyll fluorescence was evaluated as a rapid and nondestructive technique to detect low-O<sub>2</sub> or high-CO<sub>2</sub> stress in apples (*Malus domestica* Borkh.) during storage. 'Marshall' McIntosh apples were held for 5, 10, 15, 20, or 25 days at 3C in 1) standard O<sub>2</sub> (2.5% to 3%) and low CO<sub>2</sub> (<1%), 2) low O<sub>2</sub> (1% to 1.5%) and low CO<sub>2</sub> (<1%), 3) standard O<sub>2</sub> (2.5% to 3%) and standard CO<sub>2</sub> (4% to 4.5%), or 4) standard O<sub>2</sub> (2.5% to 3%) and high CO<sub>2</sub> (11% to 12%). Only 10% of the apples had skin discoloration after 5 days in 1% to 1.5% O<sub>2</sub>; 80% developed skin discoloration after 20 days in low O<sub>2</sub>. Small desiccated cavities in the cortex, associated with CO<sub>2</sub> injury, developed in 10% of the apples after 20 days in 11% to 12% CO<sub>2</sub>. Five days in 1% to 1.5% O<sub>2</sub> or 11% to 12% CO<sub>2</sub> caused variable fluorescence (Fv) of apple fruit to decrease compared to those held in standard atmospheres. Additional exposure did not significantly affect Fv in either the low-O<sub>2</sub> (1% to 1.5%) or high-CO<sub>2</sub> (11% to 12%) treatment. Our results suggest that chlorophyll fluorescence techniques can detect low-O<sub>2</sub> and high-CO<sub>2</sub> stress in apples before the development of associated disorders.

During the past decade, interest has developed in the practical application of chlorophyll fluorescence as a rapid and nondestructive method to detect stress in plants. Stress or injury to plant tissue that disrupts photosynthesis changes the characteristic fluorescence pattern of that tissue. There have been numerous studies using chlorophyll fluorescence to detect environmental, chemical, and biological stresses in plants (Lichtenhaler, 1988). However, most studies have used leaves, and there has been little application of this technique to postharvest studies of fruit. Chlorophyll fluorescence has been used to follow the development of chilling injury in banana (*Musa*, AAA Group, Cavendish subgroup) and mango (*Mangifera indica* L.) (Smillie et al., 1987) and to detect chilling injury in cucumbers (*Cucumis sativus* L.) (van Kooten et al., 1992) and green peppers (*Capsicum annuum* L.) (Lurie et al., 1994).

Apples are often stored in a controlled atmosphere (CA) in which the O<sub>2</sub> is reduced to 1% to 3% and CO<sub>2</sub> is elevated to 1.5% to 5%

(Lidster et al., 1988). However, if the O<sub>2</sub> concentration is too low or the CO<sub>2</sub> concentration is too high, apples may become injured (Meheriuk et al., 1994). Our objective was to evaluate chlorophyll fluorescence as a potentially rapid and nondestructive method to detect low-O<sub>2</sub> or CO<sub>2</sub> stress in stored apples before the development of associated disorders. This report gives only preliminary results on the potential applicability of this technique to apples held under low-O<sub>2</sub> or high-CO<sub>2</sub> stress.

## Materials and Methods

**Plant material.** 'Marshall' McIntosh apples were harvested from orchards in the Annapolis Valley, Nova Scotia, Canada, in late Sept. 1993 at optimum maturity for long-term storage (based on internal ethylene concentrations and starch index). Apples were cooled to 0C within 12 h of harvest.

**Chlorophyll fluorescence measurements.** Chlorophyll fluorescence was determined using a plant productivity fluorometer (model SF-20; Richard Brancker Research, Ottawa, Ontario, Canada). Apples were dark-adapted for at least 4 h and warmed to 20C before fluorescence measurements were taken. With a light intensity of 20  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ , the fluorometer probe was placed firmly on the blush side of the apple. Each apple was numbered and marked where fluorescence was measured to ensure that subsequent readings would be from the same apple at the same location. The peak (P) and terminal (T) values of the fluores-

cence induction curve were recorded from the fluorometer digital display. The T value was obtained after 50 secs. Variable fluorescence (Fv) was calculated as (P - T)/P, which was used in the statistical analyses.

**Low-O<sub>2</sub> and high-CO<sub>2</sub> treatments.** Apples were held in ambient air at 0C for  $\approx$ 1 month or in standard CA (2.5% O<sub>2</sub> and 4.5% CO<sub>2</sub>) at 3C for 4 months. On removal from storage, apples were warmed to 20C and divided into twenty 10-apple samples; initial chlorophyll fluorescence measurements were taken. Each 10-apple sample then was placed in a 4-liter jar and attached to a gas mixing board at 3C. Appropriate gas mixtures from bottled N<sub>2</sub>, CO<sub>2</sub>, and medical air were flushed through the jars every 3 h for 3 min at a rate of 0.7 liter $\cdot\text{min}^{-1}$  to ensure gas concentrations were maintained. Gas concentrations flowing through the jars were monitored using a gas analyzer (Nova Analytical Systems, Hamilton, Ontario, Canada); concentrations within the jars were monitored daily by withdrawing 1-ml gas samples through septa attached to the jars and injecting them into a gas chromatograph (model 3400; Varian Canada, Georgetown, Ontario).

Each of five jars received one of the following treatments: 1) standard O<sub>2</sub> (2.5% to 3%) and low CO<sub>2</sub> (<1%), 2) low O<sub>2</sub> (1% to 1.5%) and low CO<sub>2</sub> (<1%), 3) standard O<sub>2</sub> (2.5% to 3%) and standard CO<sub>2</sub> (4% to 4.5%), and 4) standard O<sub>2</sub> (2.5% to 3%) and high CO<sub>2</sub> (11% to 12%). The low-O<sub>2</sub> (1% to 1.5%) treatment examined the effects of low-O<sub>2</sub> stress on chlorophyll fluorescence, using standard O<sub>2</sub> (2.5% to 3%) as a control, whereas the high-CO<sub>2</sub> (11% to 12%) treatment examined the effects of high-CO<sub>2</sub> stress on fluorescence, using standard CO<sub>2</sub> (4% to 4.5%) as a control. Although apples stored in <3% O<sub>2</sub> may develop symptoms of low-O<sub>2</sub> injury (Autio et al., 1990), no low-O<sub>2</sub> injury occurred in 'Marshall' McIntosh after long-term CA storage in 2.5% O<sub>2</sub> (DeEll and Prange, unpublished data).

One jar from each treatment was removed after 5, 10, 15, 20, or 25 days, and chlorophyll fluorescence measurements were repeated. Individual apples also were examined for external and internal symptoms of low-O<sub>2</sub> or CO<sub>2</sub> injury on removal.

**Statistical analyses.** The data were tested by the analysis of variance (ANOVA) procedure on the statistical program Genstat 5 (Payne, 1993). The factors and levels used in the analyses were atmosphere (four treatments), removal time (5, 10, 15, 20, and 25 days), and two replications, which were confounded within time (after 1 month in air at 0C and after 4 months in standard CA at 3C) because of space constraints of the gas board. In the ANOVA, there was no significant difference between the two replications in initial fluorescence values (data not presented).

## Results and Discussion

Only 10% of the apples had skin discoloration after 5 days in 1% to 1.5% O<sub>2</sub> (Table 1), but 80% developed skin discoloration after 20 days in low O<sub>2</sub>. Strong off-flavors also were

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detected (subjective tasting) in apples after 20 days in 1% to 1.5% O<sub>2</sub>. No visual symptoms of CO<sub>2</sub> injury were observed in apples held for 15 days in 11% to 12% CO<sub>2</sub>. However, six of the 40 apples evaluated after 20 and 25 days in high CO<sub>2</sub> developed small desiccated cavities in the cortex—a symptom of CO<sub>2</sub> injury. Strong

off-flavors also were detected (subjective tasting) in apples after 20 days in 11% to 12% CO<sub>2</sub>.

The Fv of apple fruit decreased from 11.5 to 7.1 after 5 days in low O<sub>2</sub>; the Fv of apples in standard O<sub>2</sub> increased from 9.9 to 12.5 (Table 1). The Fv of apple fruit decreased from 11.0 to 3.3 after 5 days in high CO<sub>2</sub>, and the Fv of apples in standard CO<sub>2</sub> decreased from 10.4 to 8.7. Exposure for >5 days did not significantly affect Fv in either the low-O<sub>2</sub> or high-CO<sub>2</sub> treatment. Low-O<sub>2</sub> or high-CO<sub>2</sub> stress was detected by changes in chlorophyll fluorescence before any major physical symptoms of low-O<sub>2</sub> injury or CO<sub>2</sub> injury developed.

Based on our studies, chlorophyll fluorescence may have potential as a rapid and non-destructive method to screen for stress tolerance in apples. For example, the relative tolerance of different cultivars and strains to low-O<sub>2</sub> or high-CO<sub>2</sub> concentrations in storage could be determined before storage using chlorophyll fluorescence techniques. However, more research is needed to determine if this technique works on other apple cultivars and strains and if low O<sub>2</sub> or high CO<sub>2</sub> affects fluorescence within hours or days of exposure.

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Table 1. Incidence of low-O<sub>2</sub> or CO<sub>2</sub> injury and chlorophyll fluorescence (Fv<sup>2</sup>) values of 'Marshall' McIntosh apples after 5 days in standard O<sub>2</sub> (2.5% to 3%), low-O<sub>2</sub> (1% to 1.5%), standard CO<sub>2</sub> (4% to 4.5%), and high-CO<sub>2</sub> (11% to 12%) atmospheres at 3C.

Atmosphere	Injury incidence (%)	Fv	
		Interval (days)	
		0	5
Low O <sub>2</sub>			
Standard	0	9.9	12.5***
Low	10	11.5	7.1
High CO <sub>2</sub>			
Standard	0	10.4	8.7***
High	0	11.0	3.3
SEM (n = 20, df = 399) <sup>3</sup>			1.52

<sup>2</sup>Fv = [(P - T)/P] × 100.

<sup>3</sup>To compare the effects of treatment × time. The statistical analysis was on Fv data from evaluations after 5, 10, 15, 20, and 25 days. However, because Fv did not change after 5 days, only these data are presented.

\*\*\*Difference significant at P < 0.001.