Seeding Growth of Catawba Rhododendron. II. Photosynthesis and Carbohydrate Accumulation and Export

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Abstract. Catawba rhododendron (Rhododendron catawbiense Michx.) seedlings of two provenances, Johnston County, N.C. (35°45’N, 78°12’W, elevation = 67 m), and Yancey County, N.C. (35°45’N, 82°16’W, elevation = 1954 m), were grown in controlled-environment chambers for 18 weeks with days at 18, 22, 26, or 30°C in factorial combination with nights at 14, 18, 22, or 26°C. Seedlings of the higher-elevation provenance generally exhibited higher net leaf photosynthetic rates (PN) than those from the lower elevation at all temperature combinations. Thus, it appears seedlings of the high-elevation provenance possess greater relative thermostolerance, expressed as net photosynthesis, than the low-elevation provenance. Eighty-seven days after initiation (DAI) of the experiment, PN showed a quadratic response to increasing day temperature, with the maximum occurring at 22°C, whereas PN decreased linearly with increasing night temperature. At 122 DAI, PN increased linearly with increasing day temperature with nights at 22 and 26°C. Highest PN were at 30/22°C and 26/22°C. Carbohydrate export increased with increasing day temperature, whereas the response to night temperature was minimal. High levels of nonstructural carbohydrates occurred at thermoperiods (22/22°C and 26/22°C) that optimize seedling growth. However, definitive trends relating seedling growth to PN, leaf carbohydrate levels, or to the amount of carbohydrate exported from the leaves were difficult to generalize due to numerous day × night interactions.

Catawba rhododendron is an outstanding ericaceous landscape plant that possesses a higher temperature optimum than most species of rhododendron (Leach, 1961; Thornton, 1990). Its southern range, however, is still limited due to factors such as high night temperatures (Harden, 1990). Thus, it would be desirable to extend the southern range of catawba rhododendron by identifying provenances possessing a higher temperature optimum for growth.

Recently, Rowe et al. (1994) compared seedling growth of a high- and a low-elevation provenance of the species exposed to 16 thermoperiods. Dry weights for the entire plant, shoots, and roots, and total leaf area of seedlings of the high-elevation provenance exceeded those of the low-elevation provenance at all temperature combinations. These results seem to indicate that the low-elevation provenance does not exhibit a higher temperature optimum for growth than the other provenance. The basis for the relatively high temperature optimum of the high-elevation provenance is intriguing and may be related to photosynthetic processes.

Doorenbos (1955) reported that plants of catawba rhododendron grown under photoperiods of 18 to 24 h grew twice as fast as plants grown under natural daylength. Nilsen (1987) investigated the effects of temperature and water stress on leaf curling of rosebay (Rhododendron maximum L.) and catawba rhododendrons. However, to our knowledge, no research has been reported to date on photosynthetic and related processes in catawba rhododendron as influenced by temperature. We, therefore, examined the effects of selected thermoperiods on photosynthetic rates, leaf carbohydrate levels, and carbohydrate export as they influence seedling growth of two provenances of catawba rhododendron.

Materials and Methods

Mature seed capsules were collected from native stands of open-pollinated plants of catawba rhododendron growing in Johnston County, N.C. (35°45’N, 78°12’W, elevation = 67 m), and Yancey County, N.C. (35°45’N, 82°16’W, elevation = 1954 m), on 31 Oct. and 20 Nov. 1989, respectively. Following collection, seeds were dried and stored as described by Rowe et al. (1994).

Seeds of both provenances, removed from storage on 10 Jan. 1990, were germinated and grown in a greenhouse for 22 months (Rowe et al., 1994). At 646 days after germination, uniform seedlings were transferred to the Southeastern Plant Environment Laboratory (Phytotron) where 16 thermoperiods were initiated the following day (day 0) using controlled-environment A-chambers (Downs and Thomas, 1983). Plants were arranged in a 4 × 4 factorial in a completely random design using nine single-plant replications per temperature treatment per provenance. The two main factors were day (18, 22, 26, and 30°C) and night (14, 18, 22, and 26°C) temperatures provided to seedlings as 9/15-h thermoperiods. Phytotron cultural conditions were described by Rowe et al. (1994).

Eighty-seven and 122 days after initiation (DAI), leaf disk samples were taken with a cork borer (2.5 cm in diameter) (Fisher #9; Fisher Scientific, Pittsburgh) from five plants within each treatment per provenance. At the beginning of the photoperiod (0800 HR), one disk was obtained from each of three separate, recently matured leaves on the same plant (i.e., AM samples). At the end of the photoperiod (1700 HR), one additional disk was obtained from the opposite side of the midvein of each leaf sampled that morning (i.e., PM samples). A total of 240 leaf disks (16 treatments × five plants per treatment × three disks per plant = 240) was collected for each sample time within each provenance at 87 and 122 days after initiation (DAI). Disks were placed immediately in vials packed in ice and frozen at ~20°C. Samples were then freeze-dried for 3 days, weighed to the nearest 0.0001 g, and stored in desiccators for further analysis.

Between 1000 and 1400 HR on the same days that leaf disks were collected, leaf gas exchange was measured with a LI-6200 closed portable infrared gas-exchange system (LI-COR, Lincoln, Neb.). Photosynthetically active radiation, air and leaf temperatures, and relative humidity inside the leaf chamber were measured concurrently with gas exchange. Net leaf photosynthetic rates (PN) were calculated using the LI-COR 6200 measurements. Data were recorded from a recently matured leaf on each of three plants per temperature treatment per provenance using a 0.25-liter chamber for 30 sec. Vapor pressure deficits within the chamber were 0.92, 1.20, 1.31, and 1.71 kPa at 18, 22, 26, and 30°C, respectively. Measurements commenced immediately after the CO2 concentration decreased in the chamber. Average CO2 concentration was 350 µmol mol–1, and steady-state PN were assumed. Carbohydrate export from leaves was calculated by subtracting dry weight gain per leaf disk sample (i.e., leaf disk dry weight – AM leaf disk dry weight) from the gain in fixed C expected from the measured PN.

Starch content was determined for those leaf disks collected 122 DAI from plants grown at all day temperatures in combination with

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nights of 14 and 22C. These samples were chosen because these night temperatures exhibited the greatest impact on growth and dry matter accumulation (Rowe et al., 1994). Disks were ground with a mortar and pestle and extracted three times with 80% (v/v) ethanol. The remaining insoluble residue was resuspended in 1 ml 30 mM HCl and boiled for 30 min. After cooling, pH was adjusted to 4.5 using KOH. The gelatinized starch was digested for 60 min at 55C using 36 units amylglucosidase [from Aspergillus oryzae (Ahlburg) Cohn.]. The amylglucosidase had previously been dialyzed against 50 mM Na-acetate buffer (pH 4.5). Samples were boiled for 1 min to stop the reaction. After cooling and centrifugation, an aliquot of the supernatant was used to measure glucose in a solution (1 ml) containing 100 mM Hepes-NaOH (pH 8.0), 5 mM MgCl₂, 1 mM NAD, 1 mM ATP, 5 mM DTT, 2.5 units hexokinase (from baker’s yeast), and 2.5 units Glic 6-P dehydrogenase [from Leuconostoc mesenteroides (Tsengovskii) van Tiegheim]. The reaction mixture was incubated at 25C for 30 min, and its absorbance at 340 nm was measured on an ultraviolet max kinetic microplate reader (Molecular Devices Corp., Menlo Park, Calif.) to determine starch content. Data were subjected initially to analysis of variance procedures. Linear and quadratic regression equations were then fit to the data, and correlations among specified variables were examined (SAS Institute, 1990).

### Results and Discussion

High-elevation seedlings generally exhibited higher Pₙ than low-elevation seedlings at all thermoperiods 87 DAI, but seedlings of the high-elevation provenance exhibited higher Pₙ at all 16 temperature combinations 122 DAI (data not presented). These data agree with those of Ledig and Korbobo (1983), who reported Pₙ of sugar maple (Acer saccharum Marsh.) were highest in progeny from high-elevation seedlings regardless of the growth temperature. However, Slatyer and Ferrar (1977) reported optimum temperature for photosynthesis decreased with increasing elevation for three provenances of snow gum (Eucalyptus pauciflora Sieber ex Spreng.).

### Table 1. Effect of day/night temperature on net leaf photosynthetic rate, specific leaf weight, C gain, carbohydrate export, and starch accumulation in seedlings of catawba rhododendron 87 and 122 days after initiation (DAI) of the experiment.

<table>
<thead>
<tr>
<th>Growth measurement</th>
<th>Analysis of variance</th>
<th>Regression equations</th>
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* D = day temperature, N = night temperature, D × N = interaction of day and night temperature, DL = day temperature (linear), DQ = day temperature (quadratic), NL = night temperature (linear), NQ = night temperature (quadratic).
* Response was nonsignificant (NS).
* Specific leaf weight = milligrams leaf dry weight per square centimeter.
* Carbon gain = specific leaf weight (PM) – specific leaf weight (AM).
* Carbohydrate export = milligrams carbohydrate exported from leaf per square centimeter per second.
* Carbohydrate export (%) = percent fixed CO₂ exported from leaf.
* Starch (AM) = milligrams starch per square centimeter at beginning of photoperiod.
* Starch (PM) = milligrams starch per square centimeter at end of photoperiod.
* Starch accumulation = starch (PM) – starch (AM).
* NS, *, **, *** Nonsignificant or significant at P ≤ 0.10, 0.05, or 0.01, respectively.

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lar results were reported by Fryer and Ledig (1972) for balsam fir [Abies balsamea (L.) Mill.] for which plant dry weights were correlated with \( P_N \). Although \( P_N \) are not always well correlated with growth, Friend and Nelson (1976) reported that increased growth rates at elevated day temperatures resulted from higher \( P_N \).

While the quantity of exported carbohydrate was also greater for high-elevation seedlings of catawba rhododendron at all temperature combinations 87 DAI, no such differences were detected in low-elevation seedlings 122 DAI (data not presented). Similarly, high-elevation seedlings had higher shoot and total plant dry weights at all thermoperiods (Rowe et al., 1994). Thus, it appears that progeny from high-elevation seed of catawba rhododendron possess a higher temperature optimum for growth and greater relative thermo-tolerance in respect to \( P_N \).

These results support those by Konovalov and Mikhailova (1955) who reported a strong correlation between \( P_N \) of seedlings of Persian walnut [Juglans regia (L.)] and their geographic origin. Seed collected from colder areas resulted in plants with high photosynthetic capacity and high tolerance to environmental changes. In addition, at high altitudes, which are characterized by short growing seasons, high rates of \( C \) assimilation could be favored by natural selection (Ledig and Korbobo, 1983). In climatically unfavorable years with late springs, dry summers, or early autumns, survival could depend on the capacity to produce and accumulate carbohydrates during relatively short periods (Ledig and Korbobo, 1983).

No significant interactions with provenances occurred; therefore, data were averaged over both provenances and reanalyzed by sampling time to provide information regarding photosynthetic rates, carbohydrate export, and starch accumulation.

Net leaf photosynthetic rates were influenced by day and night temperature, and a significant day \( \times \) night \( (D \times N) \) interaction occurred 122 DAI (Table 1). At 87 DAI, \( P_N \) showed a quadratic response to day temperature, with the maximum occurring at 22C, whereas \( P_N \) decreased linearly with increasing night temperature (Fig. 1A). At 122 DAI \( P_N \) increased linearly with increasing day temperature with nights at 22 and 26C (Fig. 1B) and exhibited a quadratic response at 22 and 26C days (Table 1). Kramer and Kozlowski (1979) reported that \( P_N \) of most woody plants increased with increasing temperature within moderate temperature ranges. Net leaf photosynthetic rates were not affected by day temperature at 14 or 18C nights. Highest \( P_N \) were observed at 30/22C and 26/22C (Fig. 1B), whereas shoot dry weight was highest at 26/18C and 22/22C (Rowe et al., 1994). This is not surprising since net photosynthesis on a per leaf basis is seldom correlated with growth (Friend and Nelson, 1976). However, one might expect growth to be correlated with the \( P_N \) of the entire canopy. In work with other ericaceous species, Malek et al. (1992a) reported that growth and \( P_N \) for flame azalea [Rhodo-

dendron calendulaceum (Michx.) Torr] were not correlated. However, the two variables were maximized at the same temperature for mountain laurel [Kalmia latifolia (L.)] (Malek et al., 1992b).

Except for \( P_M \) specific leaf weight at 87 DAI, \( AM \) and \( PM \) specific leaf weights (milligrams dry weight per square centimeter) increased linearly with increasing temperature at 87 and 122 DAI (Table 1, Fig. 2 A and B). However, with increasing night temperature, specific leaf weights showed a quadratic response at 87 DAI and a linear decrease at 122 DAI. Evening (\( PM \)) specific leaf weights were higher than morning (\( AM \)) specific leaf weights for all thermoperiods. We assume that increases in specific leaf weight as a result of net photosynthesis are attributable to an increase in carbohydrates.

Carbon disk \( C \) gain (of nonstructural carbohydrates) was affected by day and night temperature at 87 DAI, and a significant \( D \times N \) interaction also occurred at this time (Table 1). Carbon gain showed a quadratic response to day temperature with nights at 14C, with a maximum for days at 22C (Fig. 3). In contrast, \( C \) gain increased linearly with increasing day temperature with nights at 26C, with the maximum at 30C, and was relatively constant with nights at 18 and 22C. Carbon gain increased linearly with increasing night temperature when days were at 30C. The greatest \( C \) gain occurred at 22/14C, with the lowest at 30/14C (Fig. 3). The 30/14C thermoperiod also resulted in the lowest shoot dry weight (Rowe et al., 1994). Carbon gain was not affected by temperature 122 DAI (Table 1).

Carbohydrates produced by photosynthe-

![Fig. 1. Effects of day and night (N) temperature on net leaf photosynthetic rate of catawba rhododendron seedlings. (A) 87 and (B) 122 days after initiation (DAI) of the experiment. In A, [effects of day temperature (averaged over night temperatures) and night temperature (averaged over day temperatures)], each symbol represents a mean of 24 observations; in B, each symbol is a mean of six observations.](image)
Carbohydrate export from leaves depended primarily on day temperature (Table 1). Carbohydrate export increased linearly with increasing day temperature for both sample times (Fig. 4), whereas the response to night temperature was minimal, even though it was significant ($P \leq 0.10$) at 122 DAI (data not presented).

The percentage of fixed C exported from leaves increased linearly with increasing day temperature 87 DAI (Fig. 5) and with increasing night temperature 122 DAI. The percentage was highest at a 30/26°C thermoperiod for both sample times. Reduced export of C from leaves at lower temperatures may indicate low sink demand as well as a restriction of phloem translocation (Geiger and Sovonick, 1975; Minchin et al., 1983).

Active uptake of sucrose from the apoplast, thought to be the critical step in phloem loading in most species, decreases with decreasing temperature (Giaquinta, 1980). Paul et al. (1990) demonstrated that $^{14}$C exported from leaves of sunflower ($Helianthus annuus$ L.), as a proportion of $^{14}$CO$_2$ fixed, decreased when plants were transferred from 30 to 13°C, suggesting that assimilation of C exceeded utilization more at lower temperatures. Furthermore, Marowitch et al. (1986) reported that bean ($Phaseolus vulgaris$ L.) translocated a smaller proportion of C fixed in photosynthesis as temperatures decreased from 35 to 4°C, supporting the generalization that temperature affects assimilate export more than photosynthesis.

Leaf starch concentrations 122 DAI were influenced by day and night temperatures, and a D × N interaction occurred for morning (AM) starch levels (Table 1). While AM starch levels with nights at 14°C were higher than levels with nights at 22°C for all day temperatures, except 22°C (Fig. 6A), PM starch with nights at 14°C was always higher than levels with nights at 22°C (Fig. 6B). The slower rate of shoot growth (Rowe et al., 1994) and higher starch content of leaves at 18/14°C compared with 26/22°C is consistent with the high starch levels found in pangola grass ($Digitaria decumbens$ St.) when grown under cool nights (Hilliard and West, 1970). Higher carbohydrate levels would be expected at lower temperatures, as low temperatures depress respiration and new growth so that assimilates are used more slowly (Azcon-Bieto, 1983; Paul et al., 1990). Thus, carbohydrates commonly accumulate in the leaves of plants growing at low temperatures due, in part, to growth being more sensitive to reductions in temperature than photosynthesis (Farrar, 1988).

Starch accumulation during the daily 9-h photoperiod was influenced by day and night temperatures, and a D × N interaction occurred for morning (AM) starch levels (Table 1). While AM starch levels with nights at 14°C were higher than levels with nights at 22°C for all day temperatures, except 22°C (Fig. 6A), PM starch with nights at 14°C was always higher than levels with nights at 22°C (Fig. 6B). The slower rate of shoot growth (Rowe et al., 1994) and higher starch content of leaves at 18/14°C compared with 26/22°C is consistent with the high starch levels found in pangola grass ($Digitaria decumbens$ Stent.) when grown under cool nights (Hilliard and West, 1970). Higher carbohydrate levels would be expected at lower temperatures, as low temperatures depress respiration and new growth so that assimilates are used more slowly (Azcon-Bieto, 1983; Paul et al., 1990). Thus, carbohydrates commonly accumulate in the leaves of plants growing at low temperatures due, in part, to growth being more sensitive to reductions in temperature than photosynthesis (Farrar, 1988).

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respiration rates during nights at 14C than 22C may also account for higher starch levels since temperature is the most important environmental factor influencing dark respiration (Levitt, 1980). This conjecture is consistent with the findings of Pasian and Lieth (1989), who reported that rates of dark respiration increased with increasing temperature in rose (Rosa L.) species. The 30/14C thermoperiod also resulted in the lowest leaf area and shoot dry weight (Rowe et al., 1994).

In conclusion, seedlings of the higher-elevation provenance seem to possess greater relative thermotolerance, expressed as net photosynthesis, than the low-elevation provenance. Thus, seedlings from the low-elevation provenance may not be suitable for expanding the southern landscape range of catawba rhododendron. Carbohydrate export increased with increasing day temperature, but the response to night temperature was minimal. High concentrations of nonstructural carbohydrates accumulated at thermaperiods (22/22C and 26/22C) that also optimized seedling growth.

**Literature Cited**


Malek, A.A., F.A. Blazich, S.L. Warren, and J.E. Shelton. 1992b. Initial growth of seedlings of mountain laurel as influenced by day/night temper-


Fig. 6. Effects of day and night (N) temperature on starch content of catawba rhododendron seedlings 122 days after initiation of the experiment. (A) AM starch level and (B) PM starch level. Each symbol is a mean of 30 observations.

Fig. 7. Effects of day and night (N) temperature on starch accumulation of catawba rhododendron seedlings 122 days after initiation of the experiment. Each symbol is a mean of 30 observations.