Growth and Boron Uptake of Five Pecan Cultivar Seedlings

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Abstract. Growth and B uptake of five pecan (Carya illinoensis (Wangenh.) C. Koch) seedling cultivars were evaluated in two greenhouse experiments. Seedlings were exposed for 7 to 8 months to various B-containing irrigation solutions. In one study, the growth of ‘Apache’, ‘Riverside’, and ‘Burkett’ seedlings declined significantly with a 5.0-mg B/liter application that provided 12.3 mg B/liter in the soil saturation extract. In the second study, B application of 2.5 mg·liter⁻¹ (6.4 mg·liter⁻¹ in the saturation extract) reduced growth of ‘Western’ and Wichita’ seedlings. Seedling sources differed in susceptibility to B applications. ‘Apache’ and ‘Wichita’ seedlings were more sensitive cultivars in the experiments. Leaf B concentrations increased linearly with concentrations in the saturation extract (r = 0.96 to 0.99), but did not depend on the cultivar. Boron toxicity (leaf interveinal chlorosis and tip necrosis) occurred within several weeks following B application of 1.25 to 2.5 mg·liter⁻¹ (2.8 to 6.6 mg·liter⁻¹ in the saturation extract, depending on cultivar). Three months later, chlorotic areas became necrotic in leaves containing >900 mg B/kg dry weight. Severe necrosis and some defoliation occurred when B concentrations were increased further. Leaves with no injury contained ≤325 mg B/kg.

Boron, an essential element, is recognized to cause toxic effects to many crops, including pecan. Sparks and Payne (1976) reported that application of sodium borate (to alleviate B deficiency) caused toxic symptoms on leaves of ‘Cherokee’ (from 1-year-old scion) at application rates of 50 g B or more per tree. The level of B in soils that causes foliar toxicity in pecan has, however, not been adequately studied. Eaton (1935) considered, based on the visual ranking of leaf injury symptoms of many crops grown in sand culture, that pecan (cultivar unspecified) is on the border of sensitive to semi-tolerant crops. At the time, the threshold concentration that causes toxicity was left blank for future studies. Wilcox (1960) then inserted the threshold concentration in irrigation water with little explanation (1 mg B/liter), but the same threshold concentration was later interpreted by Bernstein (1974) to be the concentration in the soil saturation extract. There seem to be no reliable data to support either guideline for pecans. This study evaluated growth and B uptake responses of seedlings from selected cultivars used by the irrigated pecan industry.

General methods. Two experiments were initiated at El Paso. Experiment 1 included ‘Apache’, ‘Riverside’, and ‘Burkett’ seedlings, while ‘Western’ and ‘Wichita’ seedlings were tested in Expt. 2. Pecan nuts were stratified for 10 weeks in perlite at 3°C, sprouted in shallow trays containing vermiculite, then planted in 20-liter plastic pots. The pots contained the A horizon of Hueco sandy loam (calcareous, coarse silty, mixed, thermic, Petrocalcic Paleargid). The lower half of the pots was filled with the soil pre-treated with dissolved boron acid in amounts to satisfy the sorption deficit according to the method of Ryan et al. (1977).

One month after planting, seedlings were thinned to an equal size, having four to five simple leaves and trunk heights between 18 and 22 cm. After thinning, each pot contained two seedlings of each cultivar. Boron treatments (supplied as boric acid in tap water) with 0.50-strength Hoagland nutrient solution no. 1 without B (Hoagland and Arnon, 1950) contained trace (<0.2 mg B/liter in tap water), 2.5, 5.0, or 7.5 mg B/liter. The pots were watered in an amount to control the leaching fraction between 0.45 and 0.55 (drainage per application volume) when the soil water storage, measured gravimetrically, was depleted to half of the water-holding capacity (0.30 m·m⁻¹). The greenhouse was maintained between 16 and 32°C.

Soils were sampled midway and at the end of B treatment periods and analyzed for B in the saturation extract (U.S. Salinity Laboratory Staff, 1954) using a plasma emission spectrometer (Applied Research Laboratories, Sunland, Calif.). The average of the two values was used to express B concentrations in the soil.

Entire seedlings were harvested, roots were washed free of soil, and trunk + root fresh weight was recorded. The growth of trunk + roots (fresh weight increase) was computed as the fresh weight at harvest minus the fresh weight when B treatments began (the latter was estimated as the average fresh weight of five seedlings per cultivar). All leaves (fully expanded with petioles) were rinsed briefly with distilled water, dried at 60°C for 24 h, and ground in a Wiley mill to pass a 40-mesh screen. Ground tissue was ignited in a muffle furnace at 550°C for 12 h, and analyzed for B (Hatcher and Wilcox, 1950), and analyzed for B as described for soils.

Both experiments were split plots, with B treatments as the main plot and seed source (cultivar) as subplot (Little and Hills, 1978). The average value of two seedlings per pot
was used as the statistical unit with five replications. 'Apache', 'Riverside', and 'Burkett' (Expt. 1). Seedlings were planted in pots 27 June 1986, and B treatments began 25 July. By 31 Oct., control trunk height increased by 13%, and the seedlings were moved to an unheated shadehouse until 31 Jan. 1987. With our conditions, pecan seedlings had improved vigor following a chilling period (≈3 months providing 500 to 1000 h below 5°C) compared to continuous culture during decreasing natural photoperiod. However, poor vigor was not observed during B treatments under increasing natural photoperiod (see Expt. 2).

Seedlings completely defoliated during the dormant period. On 1 Feb., the seedlings were returned to the greenhouse and B treatments continued until 15 June (4 months), providing a 7½-month net treatment period. All leaves that had not fallen were collected and analyzed for B. Leaves injured by B were included in the analyses. The fresh weight of trunk + roots was recorded, and after rinsing with distilled water, B concentrations in trunk and whole roots (after grinding) were determined as described for leaves. Analyses for B in trunks and roots were made for seedlings exposed to the 5 mg·liter−1 treatment (12.3 mg·liter−1 in the saturation extract). 'Western' and 'Wichita' (Expt. 2). Seedlings were planted in pots 5 Sept. 1987. Boron treatments (1.25 mg liter−1) included with the treatment levels of Expt. 1) began 1 Nov. After 3½ months (17 Feb. 1988) all leaves were collected and analyzed for B. Seedlings were kept in the greenhouse and regrowth continued until 7 July when fresh trunk + root weight was determined. Leaf area of the three youngest, fully expanded leaves from the second growth period was measured using a portable area meter (LI-COR 3000; LI-COR, Lincoln, Neb.). Also, cumulative shoot elongation from 17 Feb. to 7 July was measured. The net treatment duration was 8 months.

**Boron in soil saturation extract.** Boron concentrations in the saturation extract were similar in Expts. 1 and 2 for trace (control) and 2.5 mg·liter−1 applications (Fig. 1). However, the saturation extract concentrations for the 5.0 and 7.5 mg·liter−1 treatments averaged 26% less in Expt. 2. This difference may be attributed to variation in leaching fractions between experiments.

**Seedling growth.** Boron concentration and cultivar significantly affected increases in seedling fresh weight (Fig. 1). The growth of 'Apache' trunk + roots dropped sharply with 6.6 mg B/liter in the saturation extract. Trunk + root growth of 'Riverside' and 'Burkett' seedlings decreased only 14% with this B concentration, but was significantly reduced at 12.3 mg·liter−1 in the saturation extract.

Trunk + root growth of 'Wichita' seedlings was less than half of controls with 2.8 mg B/liter in the saturation extract (Fig. 1). 'Western' seedling growth was greater than that of 'Wichita' seedlings with this B concentration, and did not decrease significantly until B was raised to 6.4 mg·liter−1 in the saturation extract. In Expt. 2, the B × cultivar interaction was significant.

Boron also reduced shoot elongation of 'Western' and 'Wichita' seedlings, which again depended on the cultivar (Table 1). Shoot growth of 'Wichita' was decreased by almost 30% with soil B levels of 2.8 and 6.4 mg·liter−1, whereas that of 'Western' did not decrease until B levels rose above 6.4 mg·liter−1. The percentage of 'Western' and 'Wichita' seedlings that produced new shoots from axillary and terminal buds decreased with increasing B concentration (combined averages of 96%, 61%, and 47% in trace, 2.8, and 6.4 mg·liter−1 soil concentrations, respectively). With higher B levels, all regrowth originated from roots, indicating a toxic effect of B on trunk vegetative buds.

**Boron toxicity symptoms.** Injury appeared within 2 to 3 weeks of treatment on fully expanded leaves of 'Apache', 'Riverside', and 'Burkett' seedlings exposed to 6.6 mg B/liter and higher in the saturation extract. Symptoms began as interveinal chlorosis and tip necrosis on older leaves (lower one-third of stem axes), followed by necrosis in marginal and interveinal regions ≈3 months later. Subsequently, extensive necrosis and drop of the older leaves occurred with concentrations of 12.3 and 16.1 mg·liter−1.

Boron injury was also observed with 'Western' and 'Wichita' seedlings, even with 2.8 mg B/liter in the saturation extract (1.25 mg·liter−1 application; Fig. 2). At this concentration, however, the youngest leaves were uninjured. Chlorosis, necrosis, and leaf cupping occurred with 6.4 mg B/liter, while 8.8 mg·liter−1 caused severe necrosis. With 12.3 mg B/liter, leaves were much smaller, with distorted margins. Soil B concentrations ≥6.4 mg·liter−1 reduced leaf area by at least 29% (Fig. 2), but there were no cultivar differences in this response (data not shown).

Tissue B concentration. Boron concentra-

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**Table 1. Cumulative shoot elongation (17 Feb. to 7 July 1988) of 'Western' and 'Wichita' pecan seedlings in relation to applied B and that in the soil saturation extract (SSE).**

<table>
<thead>
<tr>
<th>Applied Boron concn (mg·liter−1)</th>
<th>SSE</th>
<th>Shoot elongation (% control)</th>
<th>Western</th>
<th>Wichita</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.25</td>
<td>2.8</td>
<td>120</td>
<td>71</td>
<td></td>
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<tr>
<td>2.5</td>
<td>6.4</td>
<td>115</td>
<td>72</td>
<td></td>
</tr>
<tr>
<td>5.0</td>
<td>8.8</td>
<td>71</td>
<td>68</td>
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<td>7.5</td>
<td>12.2</td>
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LSD columns (21) rows (25)

<table>
<thead>
<tr>
<th>Source</th>
<th>df</th>
<th>Mean square</th>
</tr>
</thead>
<tbody>
<tr>
<td>Boron</td>
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<td>5049.9**</td>
</tr>
<tr>
<td>Cultivar</td>
<td>1</td>
<td>5392.4***</td>
</tr>
<tr>
<td>Boron × cultivar</td>
<td>3</td>
<td>1685.8*</td>
</tr>
</tbody>
</table>

*Significant at P = 0.05 or 0.01, respectively.

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**Fig. 2. Boron-induced leaf necrosis and size reduction of 'Western' and 'Wichita' pecan seedlings.** Boron concentrations are listed in mg·liter−1 in irrigation solutions. The respective soil saturation extract concentrations were 0.5, 2.8, 6.4, and 8.8 mg·liter−1. Average leaf area of controls (0 or trace B) ranged from 30 to 40 cm².
tion in leaf tissue increased linearly with that in the saturation extract. Leaves of ‘Apache’, ‘Riverside’, and ‘Burkett’ seedlings with no visible injury contained <325 mg B/kg dry weight (Fig. 3A). There were no cultivar effects on leaf B concentrations, although ‘Apache’ seedlings (Expt. 1) and ‘Wichita’ seedlings (Expt. 2) seemed to be more sensitive to high B than seedlings of the other cultivars (Fig. 1, Table 1).

Leaf injury symptoms observed in pecan leaves (Fig. 2) are different from chloride injury, which starts as leaf tip necrosis, then develops to marginal necrosis. Boron injury begins as tip burn, interveinal chlorosis, and cupping of leaves before widespread necrosis. The development of leaf injury appeared to be associated with significant growth reduction of ‘Apache’ and ‘Wichita’ seedlings, but not necessarily of the others.

Blackmon and Winsor (1946) found that marginal leaf necrosis of 17-year-old ‘Stuart’ trees occurred when leaves accumulated ~500 to 700 mg B/kg dry weight (1 to 2 months after B application). Marginal necrosis began to appear at ~300 mg kg⁻¹ on several seedlings of our study, but we recorded generally higher toxic leaf B concentrations. On average, B-affected leaves contained a minimum of ~900 mg kg⁻¹, whether after 3½ months (Fig. 3B) or ½ months (data not shown). Although leaf B concentrations of the pecan seedlings varied between Expts. 1 and 2, the close correlations (Fig. 3) indicate that pecan seedling B status can be estimated using saturation extract B or leaf B measurements in the same experimental conditions.

Despite some growth differences among ‘Apache’, ‘Riverside’, and ‘Burkett’ seedlings, B uptake characteristics were similar. Several authors suggest that B uptake is chiefly passive and movement to leaves is driven by transpiration (Oertli and Kohl, 1961; Raven, 1980). If there were a pecan rootstock having low B uptake characteristics, leaf injuries of pecan cultivars could be reduced. Such a possibility was demonstrated for grapefruit by Cooper et al. (1955). However, this possibility does not seem to exist in these particular seedling cultivars, which are commonly used as rootstock in the irrigated pecan industry. An alternative to rootstock selection would be to use pecan cultivars having higher leaf B tolerance. Based on the current results, ‘Western’ may be able to maintain growth better than ‘Wichita’ at equivalent leaf B concentrations.

**Literature Cited**


