

Production of Cuitlacoche [*Ustilago maydis* (DS) Corda] on Sweet Corn

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Abstract. In parts of central Mexico, galls of common smut, caused by *Ustilago maydis* (Syn = *Ustilago zea* Ung.), on ears of corn (*Zea mays* L.) are an edible delicacy known as cuitlacoche. Preliminary studies were done to identify methods to increase formation of ear galls on sweet corn. Of 370 sweet corn hybrids evaluated in disease nurseries, 38 hybrids were identified for which incidence of ear galls exceeded 40% in 1987 or 1988 or exceeded 12% in 1990. Inoculation techniques for inducing ear galls were: 1) spraying sporidial suspensions between leaf sheaths and stalks at the sixth to eighth nodes; 2) injecting sporidial suspensions into the sixth to eighth internodes; 3) wounding leaf sheaths at the sixth to eighth nodes with sand, followed by spraying a sporidial suspension into wounds; and 4) wounding leaf sheaths at the sixth to eighth nodes with sand in which teliospores were mixed. Only the sporidial injection technique substantially increased the incidence of smut, but it increased the incidence of stalk, tassel, and leaf galls more than ear galls. Thus, additional research is needed to determine when and how to inoculate with *U. maydis* to induce the formation of ear galls necessary to commercially produce cuitlacoche and to screen for disease resistance.

Common smut, caused by *U. maydis*, is a devastating disease of sweet corn; but in parts of central Mexico, smut galls on ears of corn (Fig. 1) are an edible delicacy known as cuitlacoche, i.e., huitlacoche (Kennedy, 1989). In Mexico, cuitlacoche is sold either fresh or canned. Recently, cuitlacoche has been marketed in urban areas of the United States as "maize mushrooms" or "Mexican truffles." Descriptions of this new, unusual food have appeared in the past 2 years in diverse popular periodicals.

Concurrent with the culinary interest in cuitlacoche, the potential of common smut as a cash crop has been noted in several popular agricultural publications. The current potential market for fresh cuitlacoche in the United States is estimated to be >45,000 kg annually (Farm Journal, 1990). Fresh corn smut cannot be imported from Mexico. Therefore, buyers have paid growers in the United States as much as \$0.50/ear for sweet corn with large ear galls. Nevertheless, one firm was able to obtain only 1400 kg of ears of cuitlacoche from growers in the eastern United States in 1989.

Reports of cuitlacoche as a cash crop gen-

erally have focused on gleaning infected ears from fields that are planted for normal production of sweet corn, but they have not considered the production of cuitlacoche as an alternative crop. To produce cuitlacoche commercially, techniques must be identified that consistently will induce the formation of ear galls. This paper reports on preliminary studies to identify methods to increase formation of ear galls on sweet corn.

Identification of susceptible hosts. Sweet corn hybrids differ in their resistance and susceptibility to *U. maydis*. A total of 370 commercial sweet corn hybrids were evaluated in Urbana, Ill., for the percent incidence of ear galls resulting from natural infection by *U. maydis* (Pataky and Headrick, 1988; Pataky et al., 1987, 1990); 150, 150, and 200 hybrids were evaluated in 1987, 1988, and 1990, respectively. Ninety hybrids were included in at least two of the three trials. The hybrids differed in maturity from ~55 to 90 days.

Hybrids were arranged in a randomized complete-block (RCB) design. Each experimental unit consisted of a single row that was 3.6 m long with ≈15 plants per row. Rows of hybrids were replicated 15 times in 1987 and 18 times in 1988 and 1990. Trials were planted 15 May 1987, 17 May 1988, and 1 May 1990. Incidence of smut resulting from natural infection was assessed during the first week of August.

Incidence of ear galls ranged from 0% to 80%, 0% to 58%, and 0% to 34%, and grand means were 17.6%, 12.5%, and 3.8% in 1987, 1988, and 1990, respectively. Incidence was <10% for more than half of the hybrids evaluated in each trial, i.e., medians were <10% (Table 1). Hybrids with an incidence of smut >40% in 1987 or 1988 or > 12% in 1990 were considered susceptible because they were more than one SD above the grand mean, i.e., z-scores were > 1.

Hybrids for which incidence of ear galls was low were not necessarily resistant to *U. maydis* because formation of ear galls depends on the growth stage at which plants are infected (Thakur et al., 1989). For example, in a different trial in Urbana, the smut-susceptible sweet corn hybrid 'Candy Bar' was planted in adjacent plots on 17 and 29 May 1988. Incidence of ear and stalk galls for the 17 May planting was 41% and 9%, respectively, but incidence of ear and stalk galls for the 29 May planting was 0% and 24%, respectively. Thus, ear galls did not develop on the susceptible hybrid planted 29 May, probably because meristematic tissues in the ear shoot were not infected.

Although the sweet corn hybrids identified in Table 1 are relatively susceptible to *U. maydis* compared with other sweet corn hybrids that are available commercially, other



Fig. 1. Galls of common smut, *Ustilago maydis*, on ears of corn are considered an edible delicacy known as cuitlacoche in Mexico. Galls are at the most desirable stage for cuitlacoche ≈12 to 16 days after mid silk.

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Table 1. Incidence (%) of ear galls of *Ustilago maydis* on sweet corn hybrids identified as susceptible¹ to *U. maydis*, based on natural infection in field trials at Urbana, Ill., in 1987, 1988, and 1990.

1987		1988		1990	
Hybrid	Ear galls (%)	Hybrid	Ear galls (%)	Hybrid	Ear galls (%)
XPH 2628	80	Earlivee	58	Earlivee II	34
Sch 4428	78	Yankee Belle	58	Sch 4005	28
Crisp n Sweet 620	73	SsuperSweet 7210	53	HMX 8386 S	22
Sch 4020	73	Spartan	52	SsuperSweet 7410	18
Crisp n Sweet 690	72	FMX 271	51	Spartan	17
Sch 4016	63	Sch 4020	50	Sweetie Bicolor 76	17
Sch 4009	59	Sweet Dreams	45	Sch 7673	15
Yankee Belle	52	Silver Bullet	42	FMX 318	15
Sch 4026	51	Candy Bar II	41	Sch 4040	14
Sch 5170	48	Snowbelle	41	SsuperSweet 7801 W	14
Sweetie 70	47			How Sweet It Is	13
FMX 165	46			Sch 5480	13
Sweetie Bicolor 76	46			FMX 320	13
SsuperSweet 7602	45			Sweetie 70	12
Paragon	44			XPH 3030 W Sh2	12
FMX 235	40			Candy Bar II	12
Sugar Buns	40			SsuperSweet 7702 BC	12
Mean (n=150)	17.6	Mean (n = 150)	12.5	Mean (n = 200)	3.8
Median	9.3	Median	8.1	Median	1.8
Mode	1-2	Mode	0	Mode	0

¹Hybrids identified as susceptible had an incidence of ear galls that was at least one SD above the sample mean.

Table 2. Incidence (%) of *U. maydis* on sweet corn planted on four dates and inoculated with sporidia on 11 July 1989 by a leaf sheath spray or a stalk injection procedure.

Application method	Ear galls (%)					Plants with galls ² (%)				
	Planting date					Planting date				
	May		June		Mean	May		June		Mean
18	30	9	19	18		30	9	19		
Control ¹	<1	0	0	0	<1	2	<1	0	0	<1
Leaf sheath spray [*]	1	<1	0	0	<1	3	3	5	13	6
Stalk injection ^{**}	8	7	1	0	4	19	45	67	68	49
Mean (planting date)	3	2	<1	0		8	16	25	27	

¹Incidence of stalk, tassel, and/or leaf galls.

²Control = no inoculation.

^{*}Leaf sheath spray = sporidial suspension sprayed at 5.5×10^5 Pa with a hand-held gun into leaf sheaths at the sixth, seventh, and eighth nodes.

^{**}Stalk injection = 2 ml of a sporidial suspension injected with a modified Vaco Pistol Grip Syringe into stalks at the sixth, seventh, and eighth internodes.

germplasm may be more suitable for the production of cuitlacoche. Most sweet corn breeders discard genotypes or do not release experimental hybrids that are extremely susceptible to *U. maydis*. If available, these extremely susceptible genotypes would be ideal for production of cuitlacoche. Similarly, extremely susceptible genotypes may exist among dent, flint, floury, and other types of corn. Ear galls from these types of corn may be equally well suited for cuitlacoche, although buyers prefer ear galls from sweet corn. Bojanowski (1969) identified a susceptible flint corn inbred, U12, for which incidence of smut from natural infection was 100% in most seasons in Poland. Also, several investigators have observed that for some corn genotypes, gall formation is specific for certain tissues, such as ears, tassels, or nodal shoots (Christensen, 1963). Genotypes susceptible to gall formation on ears would be well suited for cuitlacoche.

Inoculation procedures and time of inoculation. Although the pathogenesis of *U. maydis* on corn is well studied (Christensen, 1963), the epidemiology of common smut is

not understood completely. Davis (1936) noted that ear infections were prevalent in some years and rare in others, varying considerably in neighboring fields and in different sections of the same field. Investigations of environmental factors favoring smut infection have resulted in lists of variables that are contradictory (Smith and White, 1988). Meristematic tissue of above-ground plant parts can be infected. Galls may form on stalks, axillary buds, ears, leaves, and tassels. Infection occurs from sporidia that have been wind-disseminated or produced by germinating teliospores. Either teliospores or sporidia must be deposited in suitable infection sites on the host to induce infection. An early report indicated that systemic infection was rare, and that injury to plants was conducive to infection (Piemeisel, 1917). Higher incidence of smut at low plant populations also was reported (Wilcoxson, 1975).

Various inoculation methods to induce galls have been attempted to screen for smut resistance. Reviewing inoculation techniques evaluated in the early part of the century, Walter (1935) concluded that injury to young,

rapidly growing plants increased severity of smut, but application of large amounts of inoculum in water did not. Likewise, Christensen (1963) noted there was no real evidence that spraying or dusting plants or seed with inoculum increased the incidence of infection; whereas injection of sporidial suspensions into leaf whorls resulted in a high incidence of plants with galls. Similarly, recent trials in Australia indicated that incidence of boil smut (i.e., common smut) on plants inoculated in the whorl was only 1.4% higher than on noninoculated plants (Nuberg et al., 1986). Conversely, Thakur et al. (1989) reported 100% incidence of tassel galls resulting from sporidial injection techniques, and ~50% incidence of ear galls in greenhouse trials. Incidence of galls was related to the growth stage of the host at inoculation. Ear galls were induced at a high frequency when the sporidial suspension was injected between the leaf sheath and stalk at the sixth, seventh, and eighth nodes (the nodes at which ears were most likely to form) 0 to 8 days before tassel formation.

In 1989 and 1990, various inoculation procedures were evaluated in field trials in Urbana to identify methods that consistently induced formation of ear galls but were not as labor-intensive as the sporidial injection techniques of Christensen (1963) and Thakur et al. (1989). Such techniques would be useful for the commercial production of cuitlacoche and for screening germplasm in breeding programs for resistance to *U. maydis*.

In 1989, the treatment design was $14 \times 4 \times 3$ factorial of hybrids, planting dates, and inoculation procedures. There were three replications arranged in a split-split plot of a RCB with 15 to 22 plants per experimental unit. Twelve susceptible and two relatively resistant hybrids were main plots. Subplots were four planting dates: 18 and 30 May and 9 and 19 June. Three inoculation procedures were applied on 1 July to sub-subplots. Inoculum was produced, and inoculation procedures were slightly modified from methods described previously (Thakur et al., 1989). Plants inoculated by the leaf sheath spray procedure were sprayed with a suspension of sporidia using a Spraymiser pistol-grip hand-held spray gun (FMC Corp., Jonesboro, Ark.) operated at $\approx 5.5 \times 10^5$ Pa. Sprays were directed at leaf sheaths from the sixth to eighth nodes. Plants inoculated by the stalk injection procedure were injected with ~2 ml of the sporidial suspension in the sixth or eighth internodes using a modified 50 ml Vaco Pistol Grip Rubber Plunger Syringe (Ideal Instruments, Chicago). Plants in the control treatment were not inoculated.

Data were arcsin transformed to make variance more homogeneous before analysis of variance (ANOVA). The ANOVA for incidence of ear galls and galls on other plant tissues indicated significant differences among inoculation treatments and planting dates and an inoculation treatment \times planting date interaction. Fewer than 1% of plants in the control treatment had galls (Table 2). Incidence of ear galls from the leaf sheath spray

Table 3. Incidence (%) of *Ustilago maydis* on 14 sweet corn hybrids planted on four dates and inoculated on 11 July 1989 by a stalk injection procedure¹.

Hybrid ^a	Ear galls (%)					Plants with galls ^b (%)				
	Planting date				Mean	Planting date				Mean
	May		June			May		June		
18	30	9	19	18	30	9	19			
Earlivee	5 ^w	0 ^w	8 ^v	0	2	22 ^w	13 ^w	84 ^v	100	53
Spartan	13 ^w	10 ^w	0	0	7	43 ^w	13 ^w	42	100	52
Sch 5170	7 ^w	14 ^v	0	0	9	7 ^w	31 ^v	100	100	64
Sweetie 70	13 ^w	13 ^v	0	0	6	12 ^w	31 ^v	100	100	58
XPH 2623s	8 ^w	6 ^v	0	0	4	17 ^w	28 ^v	86	83	45
Sch 4009	0 ^w	13 ^v	0	0	4	0 ^w	38 ^v	100	100	54
Sweetie Bicolor 76	13 ^w	0 ^v	0	0	3	13 ^w	46 ^v	100	80	57
Yankee Belle	20 ^w	7 ^v	0	0	6	20 ^w	50 ^v	75	33	42
Candy Bar II	13 ^w	5 ^v	0	0	4	13 ^w	20 ^v	75	50	36
FMX 271	7 ^w	0 ^w	0	0	2	29 ^v	40 ^v	83	85	54
SsuperSweet 7210	0 ^v	0 ^v	0	0	0	6 ^v	36 ^v	71	75	42
Platinum Lady	3 ^v	0 ^v	0	0	1	15 ^v	100 ^v	83	67	62
Miracle	11 ^v	9 ^v	10	0	7	16 ^v	27 ^v	60	33	29
Florida Staysweet	0 ^v	10	0	0	4	33 ^v	100	0	0	41
Mean (planting date)	8	6	1	0		18	41	76	72	
BLSD ^d (k = 100)	4.2					15.3				

¹Two milliliters of a sporidial suspension injected with a modified hog vaccinator into the sixth, seventh, or eighth internodes (or at the most recently formed internode for younger plants).

²Incidence of stalk, tassel, and leaf galls.

³Hybrids listed according to maturity in Urbana, Ill., in 1989.

⁴Silks and tassels emerged by 11 July.

⁵Tassels emerged by 11 July.

⁶BLSD = Wallen-Duncan Bayesian LSD test, $P = 0.05$.

Table 4. Incidence (%) of ear galls 21 days after mid silk on three sweet corn hybrids wounded by sandblasting at the early or full-tassel growth stages in 1990 and inoculated with a sporidial suspension or teliospores of *Ustilago maydis*.

Hybrids	Incidence of (%) of ear galls					Mean (hybrid) ^c
	Control	Early tassel stage		Full-tassel stage		
		Sporidia	Teliospores	Sporidia	Teliospores	
Snowbelle	24	30	29	26	28	27 b
SsuperSweet 8701	24	29	33	30	29	29 b
Supersweet Jubilee	13	17	22	24	15	18 a
Mean (treatment)	20	25	28	27	24	

^aHybrid means followed by different letters are significantly different according to the Waller-Duncan Bayesian LSD test (k = 100), $P = 0.05$.

and stalk injection procedures was only 0.4% and 4%, respectively, but incidence of plants with galls was 6% and 49%, respectively (Table 2). Planting date, which affected the growth stage of plants, affected host tissues on which galls formed from stalk injections (Table 3). Ear galls were more common for the two plantings in May than for the two plantings in June. Except on the hybrids 'Miracle' and 'Florida Staysweet', ear galls formed only on plants with tassels formed or forming at inoculation (Table 3). However, incidence of galls on other plant tissues resulting from the stalk injection procedure was ~70% for plants on which tassels had not formed (Table 3).

In 1990, the treatment design was a 3 x 5 factorial of hybrids and inoculation treatments. There were four replications arranged in a split-plot of a RCB design. Each experimental unit consisted of 12 rows, 3 m long, with 12 to 15 plants per row. The five inoculation treatments were applied to main plots. Three hybrids, 'Snowbelle', 'SsuperSweet 8701', and 'Supersweet Jubilee', were planted in the 12-row subplots on 1 May. One of the five inoculation treatments was a noninoculated control. The other

four inoculation treatments were a 2 x 2 factorial consisting of two times of inoculation and two types of inoculum. Plants were inoculated as tassels began to emerge (6 July) or at the full tassel stage (13 July) with a sporidial suspension or with teliospores. The leaf sheaths from the sixth to eighth nodes of all plants were wounded immediately before inoculation. To wound plants, sand was discharged from a Sear's Heavy Duty Sandblasting System Model no. 916808 (Sears Roebuck, Chicago) operated at 4.1×10^5 Pa. For one method of inoculation, a sporidial suspension was prepared as described by Thakur et al. (1989) and sprayed directly into the wounds created by sandblasting. For the other inoculation method, teliospores were obtained from galls collected in the greenhouse, dried, and thoroughly mixed with sand before the wounding procedure. Incidence of ear galls was assessed 13 Aug. Arcsin transformation of percentage data did not result in variances that were more homogeneous than nontransformed data; therefore, analyses were done on nontransformed data.

The ANOVA for incidence of ear galls indicated significant differences among hybrids, but inoculation techniques and the in-

oculation by hybrid interaction were not significant. Incidence of ear galls was higher for 'Snowbelle' and 'SsuperSweet 8701' than for 'Supersweet Jubilee' (Table 4). Mean incidence of ear galls was 5% to 8% higher for inoculated treatments than the noninoculated treatment, but this difference was not significant as there was considerable variation among replications. Differences in incidence of ear galls on plants inoculated at the early tassel or full-tassel stages or with inocula of sporidia or teliospores were not significant.

The "labor-saving" inoculation techniques (leaf sheath sprays and sandblasting) performed poorly at inducing formation of ear galls in these trials. The stalk inoculation technique increased incidence of ear galls; however, incidence was too low to be acceptable for commercial production of cuitlacoche or for evaluation of sweet corn hybrids for reactions to *U. maydis*. Conversely, incidence of galls on leaves, stalks, and tassels was extremely high following stalk injection, thus indicating that inoculation of plants with *U. maydis* increased infection. Additional research is needed to determine when and how to inoculate with *U. maydis* to consistently induce formation of ear galls that are necessary to commercially produce cuitlacoche and to screen for disease resistance. Recently, Pope and McCarter (1991) presented a preliminary report on a relatively successful technique in which sporidia were injected into cob tissue at the mid silk stage. Also, further research is needed to identify ways by which successful inoculation techniques can be made less labor intensive.

Other considerations. In addition to identifying susceptible corn genotypes and developing inoculation techniques that insure consistent formation of ear galls, several other factors should be considered before cuitlaco-

coche can be produced commercially as an alternative crop. Some of these include: time and method of harvest, postharvest storage and handling, and build-up of inocula of *U. maydis*.

Yield and quality of cuitlacoche appear to be inversely related during the "harvest" period. Ear galls are relatively small and spongy and usually are protected by husk leaves until ≈8 to 12 days after the mid silk stage (50% of ears with silk). Galls continue to enlarge until 19 to 21 days after mid silk, when sweet corn is harvested for fresh market. Smut galls usually erupt through husk leaves ≈12 to 18 days after mid silk. Sporulation of *U. maydis* also increases during this time, and most ear galls are too mature and are of unacceptable quality for cuitlacoche 19 days after mid silk. Also, species of *Aspergillus*, *Fusarium*, *Mucor*, *Penicillium* and other genera can colonize mature galls (Christensen, 1963), rendering them unacceptable and possibly even harmful if eaten.

Because ear galls are spongy and succulent, they are difficult to harvest and ship without damage. Likewise, smut galls deteriorate very rapidly in cold storage. Freezing appears to be a relatively easy method of maintaining gall integrity, but additional research on methods of harvest and post-harvest handling and storage is necessary.

Finally, inoculation of plants to induce ear galls for a cash crop of cuitlacoche could be detrimental to subsequent crops of sweet corn. *Ustilago maydis* overwinters in the soil as teliospores. Although relationships between levels of initial inoculum and incidence of smut are not known, unharvested smut galls could potentially increase initial inoculum for subsequent crops of sweet corn planted in that field, or possibly, in adjacent fields. Therefore, cuitlacoche probably should not be grown as an alternative crop in fields that will be used for production of sweet corn, popcorn, or dent corn in subsequent years.

Literature Cited

- Bojanowski, J. 1969. Studies of inheritance of reaction to common smut in corn. *Theor. Applied Genet.* 39:32-42.
- Christensen, J.J. 1963. Corn smut caused by *Ustilago maydis*. *Amer. Phytopathol. Soc. Monogr.* 2.
- Davis, G.N. 1936. Some of the factors influencing the infection and pathogenicity of *Ustilago zae* (Beckm.) Unger on *Zea mays* L. *Iowa Agr. Expt. Sta. Res. Bul.* 199.
- Farm Journal. 1990. Cashing in on smut. *Farm J.* 114(6):29.
- Kennedy, D. 1989. *The art of Mexican cooking.* Bantam, New York.
- Nuberg, I.K., R.N. Allen, J.M. Colless, and R.E. Darnell. 1986. Field reactions of maize varieties commonly grown in Australia to boil smut caused by *Ustilago zae*. *Austral. J. Expt. Agr.* 26:481-488.
- Pataky, J.K. and J.M. Headrick. 1988. Illinois sweet corn disease nursery-1988, p. 100-107. In: J.E. Simon et al. (ed.) *Midwest vegetable variety trial report for 1988.* Purdue Univ. Agr. Expt. Sta. Bul. 551.
- Pataky, J.K., J.M. Headrick, and Suparyono. 1987. 1987 sweet corn hybrid disease nursery, p. 10-16. In: R.K. Lindstrom (ed.). *Illinois vegetable research report. I11.* Agr. Expt. Sta. Hort. Ser. 68.
- Pataky, J.K., P. Fallah Moghaddam, and J.W. Gantz. 1990. Illinois sweet corn disease nursery-1990, p. 170-179. In: J.E. Simon et al. (ed.). *Midwest vegetable variety trial report for 1990.* Purdue Univ. Agr. Expt. Sta. Bul. 600.
- Piemeisel, F.J. 1917. Factors affecting the parasitism of *Ustilago zae*. *Phytopathology* 7:294-307.
- Pope, D.D. and S.M. McCarter. 1991. The effect of inoculation method on disease incidence and severity of the corn smut caused by *Ustilago maydis*. *Phytopathology* 81:814.
- Smith, D.R. and D.G. White. 1988. Diseases of corn, p. 687-766. In: G.F. Sprague and J.W. Dudley (eds.). *Corn and corn improvement.* 3rd ed. Amer. Soc. Agron., Madison, Wis.
- Thakur, R.P., K.J. Leonard, and J.K. Pataky. 1989. Smut gall development in adult corn plants inoculated with *Ustilago maydis*. *Plant Dis.* 73:921-925.
- Walter, J.M. 1935. Factors affecting the development of corn smut, *Ustilago zae* (Beckm.) Unger. *Minn. Agr. Expt. Sta. Tech. Bul.* 111.
- Wilcoxson, R.D. 1975. The relationship between corn plant population and smut infections. *Plant Dis. Rptr.* 59:678-680.